



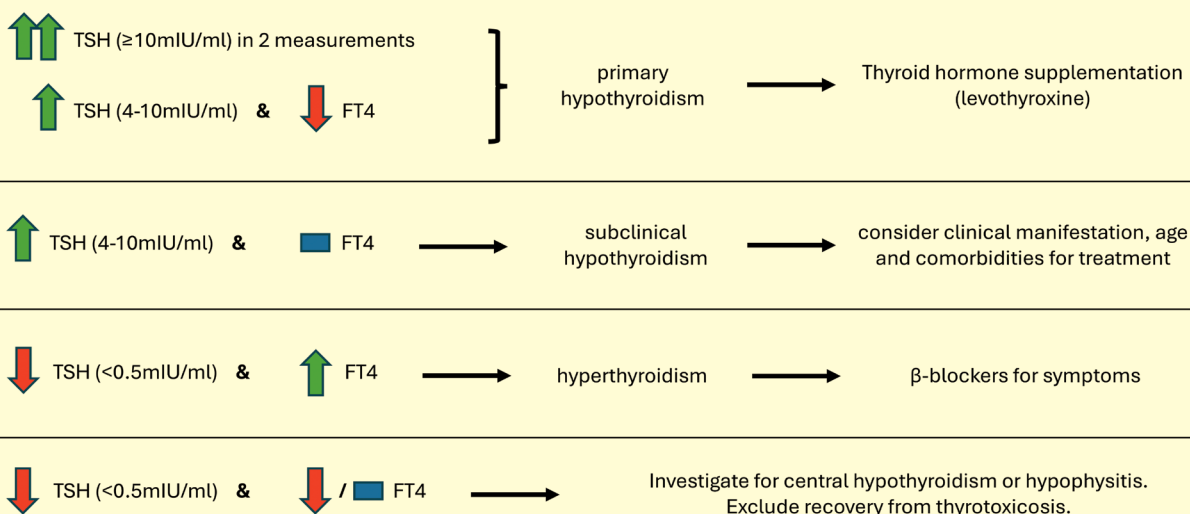
VOLUME 44 • ISSUE 2 • APRIL - JUNE 2025

Achaiki Iatriki

OFFICIAL PUBLICATION OF THE MEDICAL SOCIETY OF WESTERN GREECE AND PELOPONNESUS

Summary of monitoring the thyroid function of patients on ICI

Every 4-6 weeks for all patients under ICI therapy
or
At each cycle of immunotherapy



ACHAIKI IATRIKI

OFFICIAL JOURNAL OF THE MEDICAL SOCIETY OF WESTERN GREECE AND PELOPONNESUS (IEDEP)

GENERAL INFORMATION

ISSN Print Edition: 1106-3319

Journal Homepage: <https://achaiki-iatriki.gr/>

ISSN Electronic Edition: 1792-3018

NLM Unique ID: 9802550

Journal citation: *Achaiki iatriki* is published on behalf of the Journal of the Medical Society of Western Greece and Peloponnesus (IEDEP), representing the Society's official Journal. Please cite articles of the Journal as: Author names. Title of article. Ach Iatriki year;volume:pages.

Aims and scope: The journal publishes original papers on clinical and basic research from all areas of the health sciences including healthcare. *Achaiki iatriki*

is an open access journal. It provides immediate free access to its scientific contents and authors are not charged for submission, processing or publication of the manuscripts.

Copyright: © 2020 Medical Society of Western Greece and Peloponnesus (IEDEP)

Abstracting and indexing services: *Achaiki iatriki* is abstracted/indexed in the following databases: Google Scholar and Index Copernicus.

GOVERNING BOARD OF THE MEDICAL SOCIETY OF WESTERN GREECE AND PELOPONNESUS

President: P. Dousdampanis

Vice-President: S. Assimakopoulos

Secretary - General: M. Michalaki

Secretary - Special: I. Maroulis

Treasurer: G. Merekoulis

Members: K. Akinosoglou
E. Jelastopulu
D. Karokis
N.G. Kounis
I. Ntouvas
C. Triantos
I. Tsolakis
S. Fouzas

MEDICAL SOCIETY OF WESTERN GREECE AND PELOPONNESUS

42 Votsi Street, Patras 26221, Greece

Tel: +30 2610 279579, Fax: +30 2610 220518

email: iede_pel@yahoo.gr

Publisher

Medical Society of the Western Greece
and Peloponnesus

Editor-in-Chief

Christos Triantos
email: achaiki.iatriki@gmail.com

ACHAIKI IATRIKI

OFFICIAL JOURNAL OF THE MEDICAL SOCIETY OF WESTERN GREECE AND PELOPONNESUS (IEDEP)

Editor In-Chief

Associate Professor Christos Triantos
Department of Medicine, School of Health Sciences
University of Patras, Patras, 26504, Greece, E-mail: chtriantos@hotmail.com

Associate Editor-in-Chief

Professor Charalampos Gogos, *University of Patras, Patras, Greece*

Associate Editors

Associate Professor Stelios Assimakopoulos, *University of Patras, Patras, Greece*
Dr. Periklis Dousdampanis, *Hemodialysis Unit Kyanos Stavros Patras, Achaia, Greece*
Professor Spilios Manolakopoulos, *National and Kapodistrian University of Athens, Athens, Greece*
Professor Athanasia Mouzaki, *University of Patras, Patras, Greece*
Associate Professor Emmanouil Sinakos, *Aristotle University of Thessaloniki, Thessaloniki, Greece*

Editor-in-Chief Emeritus

Professor Emeritus Nicholas G Kounis, *University of Patras, Patras, Greece*

Emerity Editors

Professor Emeritus Konstantinos Chrysanthopoulos, *University of Patras, Patras, Greece*
Professor Emeritus Ioannis Tsolakis, *University of Patras, Patras, Greece*

EDITORIAL BOARD

Associate Professor Karolina Akinosoglou, *University of Patras, Patras, Greece*
Associate Professor Panagiotis Alexopoulos, *University of Patras, Patras, Greece*
Associate Professor Georgios Androutsopoulos, *University of Patras, Patras, Greece*
Professor Georgios Adonakis, *University of Patras, Patras, Greece*
Professor Dimitrios Apostolopoulos, *University of Patras, Patras, Greece*
Associate Professor Martha Assimakopoulou, *University of Patras, Patras, Greece*
Professor Anastasios Athanasopoulos, *University of Patras, Patras, Greece*
Associate Professor Vasiliki Bravou, *University of Patras, Patras, Greece*
Associate Professor Angelos Daniilidis, *Aristotle University of Thessaloniki, Thessaloniki, Greece*
Associate Professor Dimitrios Daoussis, *University of Patras, Patras, Greece*
Assistant Professor Foteinos Dimitrakopoulos, *University of Patras, Patras, Greece*
Associate Professor Theodoros Dimitroulas, *Aristotle University of Thessaloniki, Thessaloniki, Greece*
Professor George Dimopoulos, *National and Kapodistrian University of Athens, Athens, Greece*
Assistant Professor Stefanos Foinitsis, *Aristotle University of Thessaloniki, Thessaloniki, Greece*
Associate Professor Foteini Fligkou, *University of Patras, Patras, Greece*
Associate Professor Sotirios Fouzas, *University of Patras, Patras, Greece*
Associate Professor Georgios Germanidis, *Aristotle University of Thessaloniki, Thessaloniki, Greece*
Professor Evangelos J. Giamarellos-Bourboulis, *National and Kapodistrian University of Athens, Athens, Greece*

ACHAIKI IATRIKI

OFFICIAL JOURNAL OF THE MEDICAL SOCIETY OF WESTERN GREECE AND PELOPONNESUS (IEDEP)

Assistant Professor Despoina Gkentzi, *University of Patras, Patras, Greece*

Professor Georgios Glantzounis, *University of Ioannina, Ioannina, Greece*

Professor Kostantinos Gyftopoulos, *University of Patras, Patras, Greece*

Professor Eleni Jelastopulu, *University of Patras, Patras, Greece*

Professor George Kagadis, *University of Patras, Patras, Greece*

Professor Stavros Kakkos, *University of Patras, Patras, Greece*

Professor Christina Kalogeropoulou, *University of Patras, Patras, Greece*

Dr. Katerina Karaivazoglou, *Day Centre for Children with Autism Spectrum and other Developmental Disorders, Messolonghi, Greece*

Assistant Professor Kiriakos Karkoulas, *University of Patras, Patras, Greece*

Professor Dimitrios Karnabatidis, *University of Patras, Patras, Greece*

Assistant Professor Nikolaos Karydis, *University of Patras, Patras, Greece*

Professor Konstantinos Katsanos, *University of Ioannina, Ioannina, Greece*

Associate Professor Konstantinos Katsanos, *University of Patras, Patras, Greece*

Associate Professor Efstratios Koletsis, *University of Patras, Patras, Greece*

Associate Professor Aggelos Koutras, *University of Patras, Patras, Greece*

Associate Professor Evangelia Lampri, *University of Ioannina, Ioannina, Greece*

Associate Professor Vasiliki Labropoulou, *University of Patras, Patras, Greece*

Assistant Professor Maria Lagadinou, *University Hospital of Patras, Patras, Greece*

Professor Evaggelos Liatsikos, *University of Patras, Patras, Greece*

Professor Stamatis-Nick Liossis, *University of Patras, Patras, Greece*

Professor Markos Marangos, *University of Patras, Patras, Greece*

Professor Ioannis Maroulis, *University of Patras, Patras, Greece*

Professor Nikolaos Mastronikolis, *University of Patras, Patras, Greece*

Assistant Professor Marina Michalaki, *University of Patras, Patras, Greece*

Professor Haralampos Milionis, *University of Ioannina, Ioannina, Greece*

Associate Professor Konstantinos G. Moulakakis, *University Hospital of Patras, Patras, Greece*

Assistant Professor Konstantina Nika, *University of Patras, Patras, Greece*

Dr. Ioannis Ntouvas, *University Hospital of Patras, Patras, Greece*

Assistant Professor Marios Papasotiriou, *University of Patras, Patras, Greece*

Professor George Papatheodoridis, *National and Kapodistrian University of Athens, Athens, Greece*

Associate Professor Aikaterini Patsatsi, *Aristotle University of Thessaloniki, Thessaloniki, Greece*

Associate Professor Charalampos Pontikoglou, *University of Crete, Heraklion, Greece*

Professor George Skroubis, *University of Patras, Patras, Greece*

Professor Elena Solomou, *University of Patras, Patras, Greece*

Professor Alexandros Spiridonidis, *University of Patras, Patras, Greece*

Professor Anargiros Simeonidis, *University of Patras, Patras, Greece*

Assistant Professor Vasiliki Stamatopoulou, *University of Patras, Patras, Greece*

Dr. Ioulia Syrokosta Stathopoulou, *University Hospital of Patras, Patras, Greece*

Professor Stavros Taraviras, *University of Patras, Patras, Greece*

Professor Konstantinos Thomopoulos, *University of Patras, Patras, Greece*

ACHAIKI IATRIKI

OFFICIAL JOURNAL OF THE MEDICAL SOCIETY OF WESTERN GREECE AND PELOPONNESUS (IEDEP)

Associate Professor Vasiliki Tzelepi, *University of Patras, Patras, Greece*

Associate Professor Maria Tsironi, *University of Peloponnese, Tripoli, Greece*

Assistant Professor Sofia Tsabouri, *University of Ioannina, Ioannina, Greece*

Assistant Professor Grigorios Tsigkas, *University of Patras, Patras, Greece*

Assistant Professor Efstratios Vakirlis, *Aristotle University of Thessaloniki, Thessaloniki, Greece*

Professor Apostolos Vantarakis, *University of Patras, Patras, Greece*

Associate Professor Dimitrios Velissaris, *University of Patras, Patras, Greece*

INTERNATIONAL EDITORIAL BOARD

Professor Shomron Ben-Horin, *Sheba Medical Center, Tel-Aviv, Israel*

Professor Emeritus Nick Bouras, *King's College, London, UK*

Consultant in Internal Medicine and Gastroenterology and Senior Visiting Lecturer Pierre Ellul, *University of Malta, Malta*

Professor Vicent Hernandez, *Complejo Hospitalario Universitario de Vigo, Vigo, Spain*

Professor Konstantinos N. Lazaridis, *Mayo Clinic College of Medicine, Rochester, MN, USA*

Consultant Hepatologist and Honorary Senior Lecturer Pinelopi Manousou, *St Mary's Hospital,*

Imperial College Healthcare, NHS Trust, London, UK

Senior Consultant, Giulia Roda, IBD Center, Dept. of Gastroenterology, *Humanitas Research Hospital, Rozzano, Milan, Italy*

Associate Professor, Gerasimos Sykiotis, *Lausanne University Hospital (CHUV), Lausanne, Switzerland*

Professor Theoharis C Theoharides, *Tufts University School of Medicine, Boston, MA, USA*

Professor Christos Toumpanakis, *Royal Free Hospital, London, UK*

Professor and Honorary Consultant Emmanouil Tsochatzis, *Royal Free Hospital, London, UK*

Professor Andreas Tzakis, *Cleveland Clinic Florida, Florida, United States*

Acknowledgments

We would like to thank Dr. Ioanna Aggeletopoulou for scientific editing of the manuscripts

ACHAIKI IATRIKI

Quarterly Official Journal of the
Medical Society of Western Greece And Peloponnesus (IEDEP)

C O N T E N T S

Letter from the editor.....	66
Editorial	
Understanding and Management of Autoimmune Pancreatitis	67
Ploutarchos Pastras, Ioanna Aggeletopoulou, Christos Triantos	
Reviews	
New Trends in the Management of Soft Tissue Sarcoma	71
Anastasios Kyriazoglou, Anna Boulouta	
Difficulties in treating Rheumatoid Arthritis until nowadays	76
Alexandra Filippopoulou	
Immunotherapy - Induced Colitis	84
Gerasimia-Marina Chardalia, Angelos Koutras	
Immunotherapy-induced thyroid dysfunction	93
George C. Kyriakopoulos	
Case Report	
Childhood vitiligo: Report of a case associated with psychiatric morbidity in the patient's mother and an approach to differential diagnosis	102
Daniel A. Ndidiamaka Onyiriuka	

Dear colleagues,

In the current issue, the editorial by Pastras et al. highlights recent advances in the understanding, diagnosis, and treatment of Autoimmune Pancreatitis (AIP). The article particularly emphasizes on emerging therapeutic strategies and the shift toward pathophysiology-driven, personalized care.

The current issue features four review articles. The first, authored by Kyriazoglou et al., discusses recent advances in the management of soft tissue sarcomas, focusing on the integration of neoadjuvant therapies, immunotherapy, and precision medicine strategies to improve patient outcomes. The review by Filippopoulou et al. examines the key factors contributing to difficult-to-treat rheumatoid arthritis and highlights the critical role of early identification in enhancing disease management and improving patient outcomes. The review by Chardalia et al. underscores the clinical importance of early diagnosis of immune checkpoint inhibitor-induced colitis emphasizing management strategies aligned with the latest evidence-based guidelines, while also

exploring emerging therapeutic approaches. Finally, the review by Kyriakopoulos et al. studies the significance of immune checkpoint inhibitor-induced thyroid dysfunction, highlighting its clinical patterns, underlying mechanisms, diagnostic methods, and management strategies.

Lastly, this issue features a case report by Ndidiamaka Onyiriuka Daniel, which discusses the psychosocial impact of childhood vitiligo on both the patient and her family. It emphasizes the importance of addressing the concerns of parents and reviews the recommended treatment guidelines for childhood vitiligo, along with certain challenges associated with their application.

Yours sincerely,

C. Triantos
Associate Professor in Internal Medicine
and Gastroenterology Faculty of Medicine,
School of Health Sciences, University of Patras
Editor-in-Chief of the journal "ACHAIKI IATRIKI"

Understanding and Management of Autoimmune Pancreatitis

Ploutarchos Pastras, Ioanna Aggeletopoulou, Christos Triantos

INTRODUCTION

Autoimmune pancreatitis (AIP) is a chronic, benign inflammatory disease of the pancreas, characterized by autoimmune mechanisms. Unlike other pancreatic diseases, AIP responds dramatically to glucocorticoid therapy [1]. Although it is considered rare, it accounts for approximately 2% of chronic pancreatitis cases [2].

The clinical presentation of AIP varies widely, ranging from asymptomatic cases to symptoms such as obstructive jaundice, weight loss, chronic abdominal pain, fever, and acute pancreatitis episodes. AIP can present as a diffuse pancreatic process, which is pathognomonic, or as a focal pancreatic mass that can mimic pancreatic cancer. Histological confirmation may be necessary, with findings typically including pancreatic lymphoplasmacytic infiltration and fibrosis.

Diagnosing autoimmune pancreatitis is based on clinical features, serological markers, imaging findings from computed tomography (CT) or magnetic resonance imaging (MRI), and the classification of AIP type. Endoscopic ultrasound (EUS) with pancreatic biopsy is often crucial to rule out pancreatic cancer [3] and may also assist in determining the AIP subtype.

AIP is classified into two types: type I (lymphoplasmacytic sclerosing pancreatitis) and type II (idiopathic duct-centric pancreatitis) [1]. Each subtype differs in clinical presentation, histological features, and associated systemic involvement, necessitating tailored diagnostic and therapeutic approaches.

Autoimmune Pancreatitis Subtypes

Type I AIP (Lymphoplasmacytic Sclerosing Pancreatitis - LPSP)

Type I AIP is the most common subtype, accounting for approximately 80% of AIP cases. It is observed more frequently in Asia, particularly Japan, but its distribution is global. The mean age of onset is 60–70 years, and males are three times more likely to be affected than females. Clinically, type I AIP most often presents with painless obstructive jaundice (75%) and, less frequently, with abdominal pain. Imaging findings may include a diffuse pancreatic process or a focal pancreatic mass [4]. The hallmark feature is an elevated level of immunoglobulin G4 (IgG4)-positive plasma cells in the blood (≥ 2 times the upper limit of normal) and within the pancreatic parenchyma. Type I AIP is histologically defined as lymphoplasmacytic sclerosing pancreatitis (LPSP), according to the International Consensus Diagnostic Criteria (ICDC) [5]. This subtype often involves other organs beyond the pancreas and may present either as an isolated condition or as part of IgG4-related disease (IgG4-RD). Unlike type II AIP, type I has no association with inflammatory bowel disease (IBD). Relapses occur in up to 60% of cases following corticosteroid therapy [4].

Type II AIP (Idiopathic Duct-Centric Pancreatitis - IDCP)

Type II AIP is more prevalent in Europe and the United States compared to Asia, but it represents a minority of cases (approximately 20% of AIP patients). The mean age of onset is 40–50 years, and there is no significant gender predominance.

Patients with type II AIP commonly present with either obstructive jaundice or symptoms of acute pan-

creatitis at similar rates. Imaging frequently reveals a focal pancreatic mass (85%). Histologically, type II AIP is characterized as idiopathic duct-centric pancreatitis (IDCP), which features granulocyte epithelial lesions (GELs). Unlike type I, blood IgG4 levels are typically normal, and pancreatic tissue has very few IgG4-positive plasma cells. Type II AIP does not involve other organs but is strongly associated with IBD. Relapse is rare, occurring in less than 10% of cases [4].

Table 1 provides an overview of the key characteristics, prevalence, demographic trends, clinical features, and relapse rates associated with the different subtypes of AIP, highlighting the distinctions between type I and type II.

Other recorded disorders

Two additional disorders have been reported in the literature without being the primary classification of autoimmune pancreatitis. More specifically, AIP Not Otherwise Specified (AIP-NOS) refers to cases of AIP that do not meet the criteria for either type I or type II AIP. These patients typically lack elevated serum IgG4 levels, other organ involvement, IBD association, or histological confirmation. Evidence suggests that AIP-NOS shares similar clinical features with type II AIP, and a significant proportion of these patients are reclassified as type II AIP upon further follow-up [6].

Furthermore, Immune Checkpoint Inhibitor-Induced Pancreatic Injury is a recently described disorder, associated with immune-related adverse events caused by immune checkpoint inhibitors (ICIs). This iatrogenic pancreatic injury occurs due to a nonspecific immune response triggered by regulatory checkpoint blockade in patients undergoing treatment for advanced malignancies [7]. That condition typically manifests four–six months after initiating ICI therapy. While asymptomatic elevation of pancreatic enzymes (e.g., lipase) is the most common finding, a minority of patients may develop

symptoms of chronic pancreatitis. Diagnosis relies on excluding alternative causes of pancreatitis and establishing a temporal correlation with ICI exposure, as radiological findings are often absent. Disorder is usually seronegative, and it lacks pathognomonic histopathological lesions [7].

Therapeutic Management and Follow-Up of Autoimmune Pancreatitis

It is not necessary to initiate treatment for all patients with AIP immediately. The decision to begin therapy is guided by specific clinical indications. Patients presenting with obstructive jaundice, abdominal pain, back pain, or other symptoms involving the pancreas or extrapancreatic organs (e.g., jaundice due to bile duct strictures in overlapping IgG4-related disease) should receive prompt treatment.

Additionally, asymptomatic patients with persistent pancreatic masses on imaging, irreversible pancreatic exocrine or endocrine dysfunction, persistent abnormalities in liver function tests, or progressive subclinical lesions in vital organs associated with (overlapping) IgG4-related disease should also be treated.

Conversely, patients without these characteristics typically do not require initial therapy. This approach is supported by evidence showing recurrence rates without a definitive cure in 25–55% of AIP cases [8].

The primary treatment for AIP patients with clinical indications is prednisone at an initial dose of 40 mg per day for four–six weeks. After this induction period, repeat clinical evaluation and imaging studies (e.g., CT or MRI) are performed. Patients who demonstrate clinical and radiological improvement undergo a gradual tapering of prednisone over two months until discontinuation. Most patients with AIP (80–99%) respond to this initial corticosteroid therapy.

In cases where patients fail to respond to corticosteroids, it is crucial to re-evaluate the diagnosis and

Table 1. Summary Table of Autoimmune Pancreatitis Subtypes.

Subtype	Prevalence	Age Range	Gender Predominance	Key Features	Relapse Rate
Type I	~80%	60–70 years	Male > Female (3:1)	Elevated IgG4 levels, lymphoplasmacytic infiltration, systemic involvement, no IBD association	Up to 60%
Type II	~20%	40–50 years	Male ≈ Female	Normal IgG4 levels, granulocyte epithelial lesions, no systemic involvement, associated with IBD	<10%

exclude other conditions in the differential diagnosis [8]. For patients with contraindications to corticosteroid use, treatment with rituximab should be initiated or continued. Additionally, for certain AIP populations with co-existing IgG4-sclerosing cholangitis (IgG4-SC) as part of IgG4-RD, biliary stenting may be warranted depending on individual risk factors [9].

Follow-Up Care

After completing primary treatment, AIP patients should be monitored regularly by a physician. The suggested follow-up schedule includes clinical and serological evaluations every six months and radiological examinations initially at 4–6 weeks post-treatment, followed by a six-month interval thereafter [10].

Patients do not require maintenance therapy during follow-up, unless they experience a relapse or are considered at high risk for relapse. High-risk relapse characteristics include [8]:

1. Type I AIP with diffuse pancreatic enlargement.
2. Involvement of two or more extra-pancreatic organs or proximal IgG4-SC before treatment.
3. Delayed radiologic remission with corticosteroid therapy.
4. Persistently elevated serum IgG4 levels (>2 times the upper limit of normal) after treatment.

Maintenance Therapy

Patients experiencing relapse or identified as high risk for relapse require maintenance therapy during follow-up. The choice of maintenance therapy depends on the presence or absence of IgG4-RD:

- In AIP without IgG4-RD:
 - Azathioprine: 2 mg/kg per day (first-line therapy).
 - Alternative options:
 - Prednisone: 2.5–10 mg per day for one–three years or indefinitely.
 - Mycophenolate mofetil: 750 mg twice daily.
- In AIP with IgG4-RD:
 - Rituximab: 1 g IV on days 0 and 14 (maintenance therapy) [11].

It is important to recognize the increased risk of diabetes mellitus in AIP patients due to extensive pancreatic parenchymal atrophy. Proactive management of this risk should be considered during follow-up [12].

Potential New Treatment Options

Recent studies have explored the pathophysiology of AIP to identify new potential treatment options. Eotaxin,

a chemokine involved in the recruitment of inflammatory cells, has emerged as a promising therapeutic target, and efforts to develop targeted anti-eotaxin therapies are ongoing [13]. Regarding type II AIP, a case series has reported that colchicine may be an effective treatment option due to its ability to inhibit neutrophils and reduce the formation of pathognomonic granulocytic epithelial lesions [14]. Additionally, some patients with type II AIP and coexisting IBD have responded effectively to ustekinumab, a monoclonal antibody targeting interleukin-12 and interleukin-23 [15]. For type I AIP, several targeted therapies have been proposed, including inhibitors of B-cell activating factors such as ivalumab and inebilizumab (anti-CD19), as well as agents targeting other pathways, including simtuzumab (anti-LOX2), abatacept (anti-CD80/86), elotuzumab (anti-SLAMF7), and daratumumab (anti-CD38) [16]. More specifically, a recent study published in the *New England Journal of Medicine* (N.E.J.M.) with 135 IgG4-related disease (IgG4-RD) participants showed that inebilizumab reduced the risk of flares and increased the likelihood of flare-free complete remission at one year [17].

Furthermore, therapies like anifrolumab and sifalimumab (anti-IFN- γ), as well as etokimab (anti-IL-33), which have been successfully used in systemic lupus erythematosus, have been suggested for type I AIP. These therapies target interferon- γ and interleukin-33, which are known to drive chronic inflammation and fibrosis in this subtype [18]. The role of the gut microbiome in AIP pathogenesis has also been highlighted, particularly in type I AIP. Studies in mice have shown that gut sterilization reduces the accumulation of pathognomonic plasmacytoid dendritic cells in the pancreas, suggesting that modulation of the gut microbiome could be a potential therapeutic strategy. Future approaches may include probiotics, prebiotics, symbiotics, or fecal microbiota transplantation as prophylactic or adjunctive treatments to mitigate pancreatic inflammation and fibrosis [19].

CONCLUSION

Response to corticosteroids is a hallmark characteristic of AIP. Most patients achieve remission through induction or maintenance therapy, and relapse rates remain relatively low. While some studies are exploring and proposing novel therapies for different AIP subtypes, the majority of current clinical trials are focused on improving diagnostic methods. This emphasis reflects the greater challenge physicians face in accurately diagnos-

ing AIP in daily clinical practice. Among potential new treatment options, inebilizumab is a promising choice because the participants it tested included patients with AIP.

These trends suggest that research priorities in AIP are shifting toward a deeper understanding of its pathophysiology to facilitate the development of more precise and effective diagnostic tools. A better comprehension of the underlying mechanisms could not only improve diagnostic accuracy but also pave the way for identifying novel therapeutic approaches that enhance treatment outcomes for AIP patients.

Conflict of interest disclosure: None to declare.

Declaration of funding sources: None to declare.

Author contributions: PP was responsible for the literature review analysis; PP and IA were responsible for drafting the manuscript; CT was responsible for the revision of the manuscript for important intellectual content; all authors provided final approval for the version to be submitted.

REFERENCES

- Gallo C, Dispinzieri G, Zucchini N, Invernizzi P, Massironi S. Autoimmune pancreatitis: Cornerstones and future perspectives. *World J Gastroenterol*. 2024;30(8):817–32.
- Nishimori I, Tamakoshi A, Otsuki M. Research Committee on Intractable Diseases of the Pancreas, Ministry of Health, Labour, and Welfare of Japan. Prevalence of autoimmune pancreatitis in Japan from a nationwide survey in 2002. *J Gastroenterol*. 2007;42 Suppl 18:6–8.
- Kawa S, Kamisawa T, Notohara K, Fujinaga Y, Inoue D, Koyama T, et al. Japanese Clinical Diagnostic Criteria for Autoimmune Pancreatitis, 2018: Revision of Japanese Clinical Diagnostic Criteria for Autoimmune Pancreatitis, 2011. *Pancreas*. 2020;49(1):e13–4.
- Nagpal SJS, Sharma A, Chari ST. Autoimmune Pancreatitis. *Am J Gastroenterol*. 2018;113(9):1301.
- Shimosegawa T, Chari ST, Frulloni L, Kamisawa T, Kawa S, Mino-Kenudson M, et al. International consensus diagnostic criteria for autoimmune pancreatitis: guidelines of the International Association of Pancreatology. *Pancreas*. 2011;40(3):352–8.
- de Pretis N, Vieceli F, Brandolese A, Brozzi L, Amodio A, Frulloni L. Autoimmune pancreatitis not otherwise specified (NOS): Clinical features and outcomes of the forgotten type. *Hepatobiliary Pancreat Dis Int*. 2019;18(6):576–9.
- Sayed Ahmed A, Abreo M, Thomas A, Chari ST. Type 3 autoimmune pancreatitis (immune checkpoint inhibitor-induced pancreatitis). *Curr Opin Gastroenterol*. 2022;38(5):516–20.
- Okazaki K, Chari ST, Frulloni L, Lerch MM, Kamisawa T, Kawa S, et al. International consensus for the treatment of autoimmune pancreatitis. *Pancreatology*. 2017;17(1):1–6.
- Majumder S, Mohapatra S, Lennon RJ, Piovezani Ramos G, Postier N, Gleeson FC, et al. Rituximab Maintenance Therapy Reduces Rate of Relapse of Pancreaticobiliary Immunoglobulin G4-related Disease. *Clin Gastroenterol Hepatol*. 2018;16(12):1947–53.
- Soliman H, Vullierme MP, Maire F, Hentic O, Ruszniewski P, Lévy P, et al. Risk factors and treatment of relapses in autoimmune pancreatitis: Rituximab is safe and effective. *United Eur Gastroenterol J*. 2019;7(8):1073–83.
- Löhr JM, Beuers U, Vujasinovic M, Alvaro D, Frøkjær JB, Buttgerit F, et al. European Guideline on IgG4-related digestive disease - UEG and SGF evidence-based recommendations. *United Eur Gastroenterol* 2020;8(6):637–66.
- Majumder S, Takahashi N, Chari ST. Autoimmune Pancreatitis. *Dig Dis Sci*. 2017;62(7):1762–9.
- Mari A, Kadah A, Mahamid M, Sbeit W, Khoury T. IgG4 Related Autoimmune Pancreatitis: An Overview and the Emerging Role of Serum Eotaxin as a Potential Treatment Target. *Isr Med Assoc J*. 2019;21(9):620–3.
- Chiabrando F, Lanzillotta M, Palumbo D, Pedica F, Caruso M, Capurso G, et al. Treating Type 2 Autoimmune Pancreatitis With Colchicine: A Case Series. *Ann Intern Med*. 2021;174(12):1775–6.
- Lauri G, D'Amico F, Allocca M, Palumbo D, Della-Torre E, De Cobelli F, et al. Ustekinumab as Induction and Maintenance Therapy in Patients with Inflammatory Bowel Disease and Type II Autoimmune Pancreatitis: Report of Two Cases. *J Crohns Colitis*. 2023;17(9):1552–4.
- Okazaki K, Ikeura T, Uchida K. Recent progress on the treatment of type 1 autoimmune pancreatitis and IgG4-related disease. *Mod Rheumatol*. 2023;33(2):237–41.
- Stone H. J, Khosroshahi A, Zhang W, Torre D. E, Okazaki K, Tanaka Y, et al. Inebilizumab for Treatment of IgG4-Related Disease. *N Engl J Med*. 2024 Nov 14.
- Minaga K, Watanabe T, Hara A, Yoshikawa T, Kamata K, Kudo M. Plasmacytoid Dendritic Cells as a New Therapeutic Target for Autoimmune Pancreatitis and IgG4-Related Disease. *Front Immunol*. 2021;12:713779.
- Kamata K, Watanabe T, Minaga K, Hara A, Yoshikawa T, Okamoto A, et al. Intestinal dysbiosis mediates experimental autoimmune pancreatitis via activation of plasmacytoid dendritic cells. *Int Immunol*. 2019;31(12):795–809.

Corresponding author:

Christos Triantos
Associate Professor in Internal Medicine and Gastroenterology.
E-mail: chtriantos@upatras.gr

New Trends in the Management of Soft Tissue Sarcoma

Anastasios Kyriazoglou, Anna Boulouta

Abstract

Soft tissue sarcomas (STS) are a heterogeneous group of tumors originating from connective tissues. The management of STS has historically been challenging due to their rarity, heterogeneity, and complex anatomical locations. Surgical excision remains the standard treatment for localized STS, often supplemented by radiotherapy and chemotherapy to control recurrence and metastasis. Recent advancements in the era of immunotherapy and precision medicine have introduced novel therapeutic strategies, including neoadjuvant combination treatments and immune checkpoint inhibitors. Neoadjuvant therapies incorporating chemotherapy agents and tyrosine kinase inhibitors (TKIs), often combined with radiotherapy and regional hyperthermia, have improved surgical outcomes and survival rates. In metastatic STS, immune checkpoint inhibitors have shown promising efficacy, particularly in specific histologic subtypes. The identification of biomarkers such as tumor mutational burden (TMB) and tertiary lymphoid structures (TLS) has the potential to guide and personalize immunotherapy, improving prognostic accuracy and treatment efficacy. This review highlights these emerging trends, emphasizing the importance of integrating novel therapeutic approaches and precision medicine in the management of STS to optimize patient outcomes.

Key words: *Soft tissue sarcoma; treatment; chemotherapy; radiotherapy; immunotherapy; pathologic complete response.*

INTRODUCTION

Soft tissue sarcomas (STS) are a heterogeneous group of malignant tumors originating in connective tissues, including muscles, fat, blood vessels, nerves, and fibrous tissues, and accounting for approximately 1% of malignant tumors in adults [1]. Historically, the management of STS has been challenging due to their rarity, heterogeneity, and the complex anatomical locations they often involve. Surgical excision remains the standard treatment for early disease. However, due to high recurrence rates, the addition of perioperative systemic therapies and/or radiotherapy (RT) has been integrated [2,3]. RT has been a standard adjunct to surgery, particularly for high-grade and large tumors,

aiming to control local disease. Chemotherapy plays a well-established role in managing metastatic disease and serves as a perioperative treatment for certain high-risk patients [2]. With the advent of immunotherapy and precision medicine, significant advances have been made in the management of STS, such as neoadjuvant combination treatments and immune checkpoint inhibitors. New trends in STS therapeutics emphasize improving treatment efficacy and tailoring therapeutic strategies to individual patients.

MATERIALS AND METHODS

This review aims to highlight and evaluate the emerging trends and advancements in the management of STS, including an analysis of novel therapeutic approaches, the role of precision medicine, and the integration of multimodal treatment strategies. To achieve this, we conducted a bibliographic search of the PubMed database using the following terms: “Soft

tissue sarcoma" AND ("treatment") AND ("chemotherapy" OR "radiotherapy" OR "hyperthermia" OR "TKI") AND ("neoadjuvant" OR "adjuvant" OR "perioperative"). We included phase I, II or III clinical trials, reviews, as well as the ESMO–EURACAN–GENTURIS Clinical Practice Guidelines for diagnosis, treatment and follow-up of soft tissue and visceral sarcomas.

RESULTS

Early disease

Surgery is the standard treatment for all patients with localized STS, with en-bloc excision with R0 margins being the standard procedure [3]. For high-grade tumors, RT is the classic adjunct treatment. While it was historically preferred postoperatively, it is nowadays used as a preoperative treatment with or without chemotherapy [5]. Neoadjuvant treatment for STS can facilitate surgical resection and lower rates of local relapse and distant metastases, compared to adjuvant treatment. There is no standard neoadjuvant chemotherapy treatment for STS, however, among expert centers, a combination of doxorubicin and ifosfamide is preferred [3].

Radiotherapy and hyperthermia

For patients with localized high-risk STS, incorporating regional hyperthermia (RHT) into neoadjuvant therapy has led to improved survival rates and better local progression-free survival. Therefore, for those eligible for neoadjuvant therapy, adding RHT to the treatment regimen may be beneficial [6]. Another issue that is under investigation is the hypofractionation of RT, meaning an RT schedule that is given over a shorter period of time than standard RT. The standard-of-care preoperative RT dose regimen for STS remains 25×2 Gy [7]. Recent studies on moderate hypofractionation and ultra-hypofractionation for STS have demonstrated that these alternative schedules generally achieve comparable local control outcomes to the conventional RT regimen [8]. In addition, a phase II study of hypofractionated RT combined with chemotherapy for extremity STS resulted in a disease-free survival (DFS) of 72%, with a median follow-up of 29 months [9].

Chemotherapy and TKIs

A chemotherapeutic agent that has demonstrated promising activity in the neoadjuvant treatment of STS, especially in myxoid liposarcoma (MLS), is trabectedin. In a recent phase II clinical trial, 51% of patients treated with preoperative trabectedin combined with RT achieved over 90% necrosis in the tumor speci-

men [10]. Another agent showing promising results in combination with RT is pazopanib. In the ARST1321 randomized phase II trial, the addition of pazopanib to neoadjuvant chemoradiotherapy improved the rate of pathologic complete response, increasing to 58% compared to 22% [11]. A similar study of neoadjuvant pazopanib plus chemotherapy in patients with high-grade localized soft tissue sarcoma more than double the historical pathologic complete response (pCR) rates observed with neo-adjuvant RT alone [12].

Pathologic complete response

pCR is a marker that is used in daily clinical practice in other neoplasms, such as breast and urothelial cancers. Even though it is not yet integrated into the therapeutic approach of STS, there are recent data that indicate a correlation between pCR and clinical outcomes in these patients. A study of neoadjuvant RT for STS had shown a positive correlation between the rate of pCR and distant recurrence-free survival (DRFS) [13]. The RTOG 9514 and RTOG 0630 trials of neoadjuvant chemoradiotherapy proved that pCR after preoperative treatment is correlated with a superior five-year overall survival (OS) [14,16]. Additionally, a study evaluating neoadjuvant chemotherapy versus chemoradiotherapy for retroperitoneal STS, concluded in a superior five-year disease specific survival (DSS) in the group of patients that achieved pCR [15]. The above results show that pCR is a marker that needs to be investigated further and utilized in therapeutic decisions for localized STS.

Metastatic disease

Immunotherapy

Several immune checkpoint inhibitors have been, and are currently being, investigated, showing promising results in certain subtypes of STS.

The SARC028 phase II trial enrolled 84 patients with both bone sarcoma (BS) and soft tissue sarcoma (STS). The STS group included patients with undifferentiated pleomorphic sarcoma (UPS), liposarcoma (LPS), leiomyosarcoma (LMS), and synovial sarcoma (SS). All participants were administered pembrolizumab at a dose of 200 mg every 21 days. There was a notably high objective response rate (ORR) in the UPS subgroup, with one patient achieving a complete response and two others achieving partial responses. Additionally, 20% of patients with LPS exhibited partial responses. The median progression-free survival (mPFS) was 30 weeks for UPS and 25 weeks for LPS [17].

The AcSe trial, a phase II multicenter study, also evaluated the effectiveness of pembrolizumab monotherapy across different sarcoma subtypes. The updated results demonstrated an ORR of 6.2%, with a mPFS of 2.8 months and a median OS of 19.7 months. There was notable heterogeneity in responses among different histotypes, with alveolar soft part sarcoma (ASPS) and SMARCA4-deficient sarcoma or malignant rhabdoid tumor showing favorable outcomes, achieving partial response rates of 50% and 25%, respectively. One patient with ASPS (7%) and one patient with epithelioid sarcoma (17%) achieved complete responses [18].

A phase II study investigating the effect of atezolizumab in patients with ASPS reported an ORR of 37%, including one complete response and 18 partial responses. The mPFS was 20.8 months. Interestingly, seven patients discontinued treatment after two years and their responses were maintained [19].

In a Chinese phase I clinical trial, toripalimab was administered to patients with advanced or refractory tumors, including 12 with ASPS. Among the ASPS patients, the ORR was 25.0%, the mOS was 34.7 months and the mPFS was 11.1 months [20].

The randomized Alliance A091401 trial demonstrated that the combination of nivolumab and ipilimumab was more effective for treating metastatic, locally advanced, or unresectable sarcomas compared to nivolumab alone. The study's primary endpoint was met, with a response rate of 16%. Responses to the combination therapy were noted in patients with ASPS, UPS, LMS, myxofibrosarcoma (MFS), and angiosarcoma (AS), with five (12%) partial responses and two (5%) complete responses [21].

Immune checkpoint inhibitors and TKI combinations

In the phase Ib/II IMMUNOSARC trial, nivolumab plus sunitinib was tested in 16 patients with STS. The ORR was 21%, while the 18-month OS was 100% for patients who had an objective response and 75% for those who had stable disease. One patient with AS achieved a complete response, while two patients with ASPS, one with AS, one with SS, and one with extraskeletal myxoid CS experienced a partial response [22].

A phase II, single-arm clinical trial of pembrolizumab combined with axitinib for advanced soft tissue sarcoma, including ASPS, showed a three-month progression-free survival of 65.6% for all evaluable patients, with a three-month progression-free survival being 72.7%, especially for patients with ASPS [23].

Alliance A091902 is a multicenter phase II study of

cabozantinib plus nivolumab for patients with advanced AS previously treated with a taxane. Preliminary data from this trial show an ORR of 62%, with 11 partial responses and two complete responses. Median PFS was 9.6 months, and OS 20.5 months [24].

Recently, the combination of durvalumab plus pazopanib was tested in metastatic and/or recurrent STS. The ORR was 30.4%, meeting the prespecified endpoint and the median PFS was 7.7 months [25].

Biomarkers for immunotherapy response

PD-L1 is known to predict responses to immune checkpoint inhibitors in several solid tumors. However, its role as a prognostic and predictive biomarker in STS is controversial [26]. A marker that has shown a stronger correlation with ICI response is tumor mutational burden (TMB). High TMB values are observed in UPS, LMS, and SS [27].

Tertiary lymphoid structures (TLS), which include T cells, follicular dendritic cells, and B cells, are part of the tumor microenvironment and appear to be better predictors of prognosis and response to therapy. Recent findings from the PEMBROSARC trial showed that STS rich in TLS and intratumoral plasma cells exhibited a better response to pembrolizumab [28].

DISCUSSION

The management of soft tissue sarcomas (STS) has evolved significantly over the past decades, with advancements in surgical techniques, the incorporation of multimodal therapies, and the advent of precision medicine and immunotherapy. This review has highlighted several key trends and innovations in the treatment of STS, particularly focusing on the roles of neoadjuvant therapies, hyperthermia, and immune checkpoint inhibitors, as well as the exploration of biomarkers to guide therapy. These trends represent a move towards more personalized, precise, and effective management of soft tissue sarcomas, aimed at improving outcomes and quality of life for patients. Future research should focus on refining these therapeutic strategies and validating biomarkers to further improve clinical outcomes for patients with soft tissue sarcomas. The continued collaboration between clinical researchers, oncologists, and multidisciplinary teams will be essential in translating these advancements into standard practice, ultimately enhancing the prognosis and quality of life for patients with STS. Table 1 summarizes the new therapeutic options in the management of STS.

Table 1.

Therapeutic option	Sarcoma type	Disease stage	Outcome
Neoadjuvant RT +/- Chemotherapy	All STS	Locally advanced	Lower rates of local relapse and distant metastases [5]
Neoadjuvant Hypofractionated RT + Chemotherapy	All STS	Locally advanced	Increased DFS [8,9]
Neoadjuvant regional hyperthermia	All STS	Locally advanced	Increased survival rates and local PFS [6]
Neoadjuvant trabectedin + RT	MLS	Locally advanced	Increased tumor necrosis [10]
Neoadjuvant pazopanib + RT	All STS	Locally advanced	Increased tumor necrosis [11,12]
Pembrolizumab	All STS	Recurrent/Metastatic	High ORR, mPFS, mOS [18]
Atezolizumab	ASPS	Recurrent/Metastatic	High ORR, mPFS [19]
Toripalimab	All STS	Recurrent/Metastatic	High ORR, mOS, mPFS in ASPS patients [20]
Nivolumab/Ipilimumab	All STS	Locally advanced/Metastatic/Unresectable	High ORR [21]
Nivolumab/Sunitinib	All STS	Recurrent/Metastatic	High ORR [22]
Pembrolizumab/Axitinib	All STS	Recurrent/Metastatic	High 3-month PFS [23]
Nivolumab/Cabozantinib	AS	Recurrent/Metastatic previously treated with taxane	High ORR, mPFS, mOS [24]
Durvalumab/Pazopanib	All STS	Recurrent/Metastatic	High ORR [25]

CONCLUSION

The landscape of STS management is rapidly evolving, with significant advancements in both neoadjuvant and metastatic disease treatments. The integration of new radiotherapy regimens, hyperthermia, novel chemotherapeutic agents, TKIs, immune checkpoint inhibitors and combinations of the above, have enhanced the efficacy of traditional treatment regimens. Moreover, the identification of predictive biomarkers for therapy response represents a significant step towards personalized medicine in STS.

Conflict of interest disclosure: None to declare.

Declaration of funding sources: None to declare.

Author contributions: AB, conceptualization, methodology, writing – original draft, investigation, data curation, systematic review search; AK, conceptualization, methodology, writing – review and editing, supervision, investigation, data curation, systematic review search.

REFERENCES

1. Burningham Z, Hashibe M, Spector L, Schiffman JD. The epidemiology of sarcoma. Clin Sarcoma Res. 2012;2(1):14.
2. Gatta G, Capocaccia R, Botta L, Mallone S, De Angelis R, Ardanaz E, et al. Burden and centralised treatment in Europe of rare tumours: results of RARECAREnet-a population-based study. Lancet Oncol. 2017;18(8):1022-39.
3. Casali PG, Abecassis N, Bauer S, Biagini R, Bielack S, Bonvalot S, et al. Soft tissue and visceral sarcomas: ESMO-EURACAN Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann Oncol. 2018;29(Suppl 4):iv51-67.
4. Beane JD, Yang JC, White D, Steinberg SM, Rosenberg SA, Rudloff U. Efficacy of adjuvant radiation therapy in the treatment of soft tissue sarcoma of the extremity: 20-year follow-up of a randomized prospective trial. Ann Surg Oncol. 2014;21(8):2484-9.
5. Palassini E, Ferrari S, Verderio P, De Paoli A, Broto JM, Quagliuolo V, et al. Feasibility of preoperative chemotherapy with or without radiation therapy in localized soft tissue sarcomas of limbs and superficial trunk in the Italian Sarcoma Group/ Grupo Español de Investigación en Sarcomas randomized clinical trial: three versus five cycles of full-dose epirubicin plus ifosfamide. J Clin Oncol. 2015;33(31):3628-34.
6. Issels RD, Lindner LH, Verweij J, Wessalowski R, Reichardt

- P, Wust P, et al. Effect of neoadjuvant chemotherapy plus regional hyperthermia on long-term outcomes among patients with localized high-risk soft tissue sarcoma. *JAMA Oncol.* 2018;4(4):483-92.
7. Salerno KE, Alektiar KM, Baldini EH, Bedi M, Bishop AJ, Bradfield L, et al. Radiation therapy for treatment of soft tissue sarcoma in adults: executive summary of an ASTRO Clinical Practice Guideline. *Pract Radiat Oncol.* 2021;11(5):339-51.
 8. Guadagnolo BA, Baldini EH. Are we ready for life in the fast lane? A critical review of preoperative hypofractionated radiotherapy for localized soft tissue sarcoma. *Semin Radiat Oncol.* 2024;34(2):180-94.
 9. Gobo Silva ML, Lopes de Mello CA, Aguiar Junior S, D'Almeida Costa F, Stevanato Filho PR, Santoro Bezerra T, et al. Neoadjuvant hypofractionated radiotherapy and chemotherapy for extremity soft tissue sarcomas: safety, feasibility, and early oncologic outcomes of a phase 2 trial. *Radiother Oncol.* 2021;159:161-7.
 10. Sanfilippo R, Hindi N, Cruz Jurado J, Blay JY, Lopez-Pousa A, Italiano A, et al. Effectiveness and safety of trabectedin and radiotherapy for patients with myxoid liposarcoma: a nonrandomized clinical trial. *JAMA Oncol.* 2023;9(5):656-63.
 11. Weiss AR, Chen YL, Scharschmidt TJ, Chi YY, Tian J, Black JO, et al. Pathological response in children and adults with large unresected intermediate-grade or high-grade soft tissue sarcoma receiving preoperative chemoradiotherapy with or without pazopanib (ARST1321): a multicentre, randomised, open-label, phase 2 trial. *Lancet Oncol.* 2020;21(8):1110-22.
 12. Van Meekeren M, Bovee JVMG, van Coevorden F, van Houdt W, Schrage Y, Koenen AM, et al. A phase II study on the neoadjuvant combination of pazopanib and radiotherapy in patients with high-risk, localized soft tissue sarcoma. *Acta Oncol.* 2021;60(12):1557-64.
 13. Canter RJ, Martinez SR, Tamurian RM, Wilton M, Li C, Ryu J, et al. Radiographic and histologic response to neoadjuvant radiotherapy in patients with soft tissue sarcoma. *Ann Surg Oncol.* 2010;17(10):2578-84.
 14. Kraybill WG, Harris J, Spiro IJ, Ettinger DS, DeLaney TF, Blum RH, et al. Phase II study of neoadjuvant chemotherapy and radiation therapy in the management of high-risk, high-grade, soft tissue sarcomas of the extremities and body wall: Radiation Therapy Oncology Group Trial 9514. *J Clin Oncol.* 2006;24(4):619-25.
 15. Donahue TR, Kattan MW, Nelson SD, Tap WD, Eilber FR, Eilber FC. Evaluation of neoadjuvant therapy and histopathologic response in primary, high-grade retroperitoneal sarcomas using the sarcoma nomogram. *Cancer.* 2010;116(16):3883-91.
 16. Wang D, Harris J, Kraybill WG, Eisenberg B, Kirsch DG, Ettinger DS, et al. Pathologic complete response and clinical outcomes in patients with localized soft tissue sarcoma treated with neoadjuvant chemoradiotherapy or radiotherapy. *JAMA Oncol.* 2023;9(5):646-54.
 17. Tawbi HA, Burgess M, Bolejack V, Van Tine BA, Schuetze SM, Hu J, et al. Pembrolizumab in advanced soft-tissue sarcoma and bone sarcoma (SARC028): a multicentre, two-cohort, single-arm, open-label, phase 2 trial. *Lancet Oncol.* 2017;18(11):1493-501.
 18. Blay JY, Chevret S, Le Cesne A, Brahmi M, Penel N, Cousin S, et al. Pembrolizumab in patients with rare and ultra-rare sarcomas (AcSé Pembrolizumab): analysis of a subgroup from a non-randomised, open-label, phase 2, basket trial. *Lancet Oncol.* 2023;24(8):892-902.
 19. Chen AP, Sharon E, O'Sullivan-Coyne G, Moore N, Foster JC, Hu JS, et al. Atezolizumab for advanced alveolar soft part sarcoma. *N Engl J Med.* 2023;389(10):911-21.
 20. Yang J, Dong L, Yang S, Han X, Han Y, Jiang S, et al. Safety and clinical efficacy of toripalimab, a PD-1 mAb, in patients with advanced or recurrent malignancies in a phase I study. *Eur J Cancer.* 2020;130:182-92.
 21. D'Angelo SP, Mahoney MR, Van Tine BA, Atkins J, Milhem MM, Jahagirdar BN, et al. Nivolumab with or without ipilimumab treatment for metastatic sarcoma (Alliance A091401): two open-label, non-comparative, randomised, phase 2 trials. *Lancet Oncol.* 2018;19(3):416-26.
 22. Martin-Broto J, Hindi N, Grignani G, Martinez-Trufero J, Redondo A, Valverde C, et al. Nivolumab and sunitinib combination in advanced soft tissue sarcomas: a multicenter, single-arm, phase Ib/II trial. *J Immunother Cancer.* 2020;8(2):e001561.
 23. Wilky BA, Trucco MM, Subhawong TK, Florou V, Park W, Kwon D, et al. Axitinib plus pembrolizumab in patients with advanced sarcomas including alveolar soft-part sarcoma: a single-centre, single-arm, phase 2 trial. *Lancet Oncol.* 2019;20(6):837-48.
 24. Grilley-Olson JE, Allred JB, Schuetze S, Davis EJ, Wagner MJ, Poklepovic AS, et al. A multicenter phase II study of cabozantinib + nivolumab for patients with advanced angiosarcoma previously treated with a taxane (Alliance A091902). *J Clin Oncol.* 2023;41(16_suppl):11503.
 25. Cho HJ, Yun KH, Shin SJ, Lee YH, Kim SH, Baek W, et al. Durvalumab plus pazopanib combination in patients with advanced soft tissue sarcomas: a phase II trial. *Nat Commun.* 2024;15(1):685.
 26. Siozopoulou V, Domen A, Zwaenepoel K, Van Beeck A, Smits E, Pauwels P, et al. Immune checkpoint inhibitory therapy in sarcomas: is there light at the end of the tunnel? *Cancers (Basel).* 2021;13(2):360.
 27. Anastasiou M, Kyriazoglou A, Kotsantis I, Economopoulou P, Kyrkasiadou M, Giannopoulou A, et al. Immune checkpoint inhibitors in sarcomas: a systematic review. *Immunooncol Technol.* 2023;20:100407.
 28. Italiano A, Bessede A, Pulido M, Bompas E, Piperno-Neumann S, Chevreau C, et al. Pembrolizumab in soft-tissue sarcomas with tertiary lymphoid structures: a phase 2 PEMBROSARC trial cohort. *Nat Med.* 2022;28(6):1199-206.

Corresponding author:

Anna Boulouta
 Second Department of Internal Medicine, Oncology Unit,
 Attikon University Hospital, Athens, Greece
 E-mail: annitaboulouta@gmail.com

Difficulties in treating Rheumatoid Arthritis until nowadays

Alexandra Filippopoulou

Abstract

Rheumatoid arthritis (RA), mainly affecting women, is the most frequent rheumatic disease with different phenotypes. In recent years, despite the development of new therapies, clinical remission is not always achieved by all patients. Disease activity remains high or moderate and corticosteroids cannot be tapered, despite the use of conventional synthetic Disease-Modifying Antirheumatic Drugs (csDMARDs) and/ or at least two different biologic agents. The difficult to treat RA and the consequent decreased quality of life, remain an intractable problem in everyday clinical practice. Specific genetic factors are reported to be potentially involved, whereas comorbidities, including obesity, osteoarthritis and/or fibromyalgia may aggravate the uncontrolled disease. Moreover, atherosclerosis and malignancies are more frequently presented in cases of refractory RA, often leading to limited therapeutic options. Patients with difficult-to-treat arthritis are additionally prone to infections, occasionally pausing their treatment. Pregnancy and lactation may complicate the disease by imposing specific precautions, also involving the period before conception. Consequently, only allowable therapy is indicated in all cases. Furthermore, several environmental factors, including smoking, coffee consumption and lack of physical exercise, may negatively affect disease activity, despite treatment. Patients' quality of life may additionally deteriorate because of pharmacologically induced adverse events, such as osteoporosis, particularly following steroids administration. Finally, lower adherence to treatment, frequently related to patients' lower educational and financial situation, combined with the absence of only one standardized tool for assessing patient's perception of the severity of their condition, may also lead to difficult-to-treat disease. It is worth noting that the RA population is not treated uniformly worldwide, since drug prescription and administration vary across different areas, according to local economic regulations. For all the above reasons, early recognition of poor prognostic factors is essential to prevent disease flares and hence, therapy escalation, with controversial outcomes.

Key words: *Rheumatoid arthritis; difficult-to-treat; high disease activity; complications*

INTRODUCTION

Rheumatoid arthritis (RA) is one of the most common autoimmune inflammatory diseases, mainly affecting the joints of upper or/and lower extremities in a symmetrical pattern. Extra-articular manifestations, including pleuritis, pericarditis and formation of rheumatoid nodules may also occur. Different phenotypes are not uncommon in everyday clinical practice, since

Abbreviations: RA = Rheumatoid Arthritis, EULAR = European League Against Rheumatism, DMARD = Disease-Modifying Antirheumatic Drugs, csDMARDs = conventional synthetic Disease-Modifying Antirheumatic Drugs, bDMARDs = biologic Disease-Modifying Antirheumatic Drugs, ACPA = Anti-Citrullinated Protein Antibody, DAS-28 = Disease Activity Score-28, TNFi = Tumor Necrosis Factor Inhibitor, BMI = Body Mass Index, tsDMARDs = synthetic targeted Disease-Modifying Antirheumatic Drugs, jakinh = Jak inhibitor, RCT = Randomized Control Trial, TNFa = Tumor Necrosis Factor α , IL 6 = Interleukin 6

Private Medical Practice, 12 Eleftheriou Venizelou St.,
Amaliada Ileia, Greece

Received: 30 May 2024; Accepted: 07 Oct 2024

several patients may present with early joint destruction, whereas others exhibit milder symptoms at baseline [5]. Epidemiologically, RA is estimated to affect 1 % of the worldwide population [1,2] with females being twice as likely to be affected, according to most studies [3].

The last decades, the evolution of therapy has been proven to be beneficial for the majority of patients. Clinical remission or low disease activity is achieved through new therapeutic approaches, according to the European League Against Rheumatism (EULAR) recommendations. However, 5- 20% of patients remain difficult to treat, failing to reach treatment targets [4]. Unachievable corticosteroid tapering, usually accompanied by the failure of two or more conventional synthetic Disease-Modifying Antirheumatic Drugs csDMARDs and/or biologic Disease-Modifying Antirheumatic Drugs (bDMARDs) or targeted synthetic DMARDs (tsDMARDs), also known as Jak inhibitors (Jakinh), is often considered as difficult-to-treat RA. Therefore, despite the treat-to-target strategy, refractory disease is frequently identified [5,6]. Under these circumstances, the concurrent presence of comorbidities, fatigue and /or extra-articular manifestations, is frequent [4]. Additionally, the variant response to treatment, commonly observed in everyday clinical practice, is considered multifactorial.

Main text

Genetic risk factors may lead to the presence of heterogeneous clinical phenotypes; however, no specific genetic factor has been strongly associated with a better response to treatment [4]. Positivity for various alleles, such as HLA DR1 and HLA DR4, is prone to the development of RA [6]. Anti-citrullinated protein antibodies (ACPAs), the most specific autoantibodies for the disease [7,6], are more frequently present in HLA DR1 positive patients. Therefore, radiological damage, accompanied by higher inflammatory markers in sera and more swollen joints in clinical examination, is often observed [7-9]. According to De Rooy et al, the presence of SNPs located near TNF receptor-associated factor 1 (TRAF1) is highly associated with aggravation of radiographic damage, indicating that SNPs may play an important role in difficult to treat RA cases [10]. Conigliaro et al report that exacerbation of RA is associated with several SNPs, such as PSORS1C (psoriasis susceptibility 1 candidate 1, PTPN2 (protein tyrosine phosphatase non-receptor type 2, FOXO3A (Forkhead Box O3A) [4]. The methylation of DNA may also affect the synovium cells of the joint and is frequently associated with the development of a more

severe and aggressive RA phenotype and consequently more difficult to treat [4,11]. The last decades the use of DMARDs, and more specifically of bDMARDs, aiming at better management and treatment of RA, is proven to be promising [15]. On the other hand, bDMARDs are not always successful, despite their increased use. According to Schipper et al, positive predictors for rapid reach of low disease activity include male gender, younger age and low Disease Activity Score-28 (DAS28) at baseline [12]. These predictors often require early diagnosis. Unfortunately, delays in identifying RA are not uncommon. It is reported that the time lag between onset of symptoms and diagnosis ranges from one month to ten years [58], increasing the risk for a more difficult to treat disease. However, refractory RA is frequently associated, not only with delayed diagnosis, but also with the presence of comorbidities. Despite the use of biologic agents and more specifically Tumor Necrosis Factor inhibitors (TNFi), disease remission may not occur, as revealed by an Italian cohort of 308 patients [13,14]. Generally, clinical outcomes in RA are often influenced by the presence of comorbidities; hence it is highly recommended that patients should be monitored for co- or pre- existing comorbidities [26]. Among them, obesity plays an important role in difficult to treat patients. Concurrent osteoarthritis, in conjunction with depression, is often associated with exacerbation of fibromyalgia and consequently chronic pain, have a negative impact on clinical remission [16]. The assessment of DAS28 score, widely used for the estimation of disease activity in RA, may not be accurate in cases of obesity, since obese patients are frequently considered to have more swollen joints during clinical examination. For that reason, the use of ultrasound (not available in all rheumatology outpatient clinics) may be beneficial in patients with a higher BMI (Body Mass Index) [17]. Furthermore, obesity is also associated with a worse clinical response to treatment with TNFi, when compared to patients with normal BMI [18,19]. Therefore, non-TNFi administration, including abatacept or tocilizumab, is considered to be more beneficial in overweighted RA subjects, whose sera have higher amounts of IL 6, frequently resulting in exacerbation of inflammation [20-22]. The pro-inflammatory cytokine IL 6, especially in high levels, is also associated with cardiovascular diseases and atherosclerosis. Many studies report that RA patients have an increased risk for cardiovascular events, especially in high levels of inflammation [23-25].

According to an observational cohort-study with

combinatorial data from Norway and Sweden, the increased RA disease activity is correlated with higher risk for acute coronary syndrome. These results indicate the imperative need for close monitoring of these patients, who are prone to remain in a continuous non-remission state, despite treatment [25]. Additionally, Oral Surveillance survey revealed an increased risk for major adverse cardiovascular events (MACE), also concerning malignancies, in RA patients under treatment with tofacitinib (the first approved jak inhibitor in RA), compared to subjects under TNFi administration [58]. The survey results raise suspicions for the safe use of Jak inhibitors (jakinhs) in RA, and inflammatory diseases in general. Consequently, the use of jakinh is recommended after risk assessment [59] and as a result, several patients may be excluded from this kind of therapeutic options by their rheumatologists. Nonetheless post hoc analysis did not reveal significant danger for these events after jakinh administration [60], indicating that there is no real contraindication for their use. However, it is worth noting that comorbidities, including cardiovascular disease, may lead to contraindications concerning the use of several approved for RA drugs, resulting in a sustainable active disease.

Malignancies are another major problem in RA patients. An increased rate of lymphoma and lung cancer is observed among them [62–64], due to immune dysregulation and/or chronic inflammation, that often result in cell proliferation, mutagenesis, oncogene activation and angiogenesis [65]. Although it was initially believed that bDMARDs, and mainly TNFi, were potentially involved in carcinogenesis, meta-analyses, observational studies and post marketing surveillance, indicated that early use of these drugs is safe, not increasing the overall risk for malignancies [66]. Data remain controversial, reporting that patients receiving treatment for RA, especially rituximab, have a greater risk for all cancers, compared to general population [67]. The recent ORAL surveillance study also detected an increased risk for carcinogenesis after tofacitinib (jakinhs) administration compared to TNFi [58]. Burmester et al, however, did not report a higher risk of malignancy, excluding NMSC, in RA patients after a 5.45 year administration of another jak inhibitor, upadacitinib [69].

Nonetheless, managing patients with RA and concurrent cancer is a non-rare complex problem for both rheumatologists and oncologists, often resulting in a difficult to treat disease. In patients with a history of malignancy, bDMARDs, and specifically TNFi, are not

commonly preferred as therapeutic tools, while an increased use of rituximab is noticed in most cases [68]. Nowadays, treatment with TNFi in patients with prior cancer (except in cases of lymphoma or melanoma), is considered to be safe after a mean follow-up of five years, since no significant cancer recurrence is reported, under these circumstances [70]. However, since no robust data from randomized clinical trials (RCTs) exist, the therapeutic management of RA patients during the first five years after malignancy diagnosis still remains an ongoing question.

An increased rate of infections (both outpatient and those requiring hospitalization), is observed in patients with refractory disease [28]. Serious infections, especially under treatment with bDMARDs or jakinh, are common in RA population with high disease activity, especially during the first year after treatment initiation [27,72]. Bacterial infections are more commonly observed in these patients. Interestingly, in case of sepsis, a lower risk of death is noted in subjects treated with bDMARDs, compared to subjects, who are on therapy with conventional synthetic DMARDs (csDMARDs) [71]. CsDMARDs, initially administered after the diagnosis of RA and frequently in combination with low daily doses of glucocorticosteroids (5–7.5 mg), are generally considered safe [71–74]. Risk factors for severe infections among RA patients include older age (> 65 years), high disease activity and disability score (as defined by DAS-28 and HAQ score, respectively), presence of comorbidities (chronic lung or kidney disease), higher doses of glucocorticoids (> 7.5 mg/day), history of previous serious infections and previous DMARDs failures and, finally, current immunosuppressive therapy [71]. Infectious complications should be taken into consideration, because in case of infection, according to general recommendations in everyday clinical practice, patients with autoimmune diseases, including RA, are advised to temporarily suspend their immunosuppressive therapy. This practice, however, often results in a non remission state, especially when pausing is frequent.

Pregnancy is another factor that may cause difficulties in treating RA. Since women are mainly affected by the disease, the periods of pregnancy and puerperium, especially with concomitant lactation, are critical. Although 60 % of pregnant RA women (particularly without positive RF and/or ACPAs in the serum) are reported to be in remission during pregnancy [29–30], treatment in this period remains challenging. Contraindications for the use of several drugs are the

major problem for rheumatologists. Methotrexate, the basic and most widely used DMARD in rheumatology, is known for its teratogenic effects on the fetus. The drug is also combined with an increased rate of abortion [31]. Generally, for safety reasons, the discontinuation of methotrexate is recommended, three months prior to scheduled conception [34,35].

Taking into consideration that 40% of pregnancies are unintended worldwide [32], women diagnosed with RA in their reproductive age, should be advised for contraception measures, and if they still express the desire to conceive in near future, only conventional medications can be prescribed, including: glucocorticoids (prednisolone is particularly selected, since, due to its lower transplacental transfer, earlier fetal lung maturation is avoided), hydroxychloroquine, azathioprine, cyclosporine and sulfasalazine [4,33]. Those drugs are also compatible with breastfeeding, except from sulfasalazine that is not recommended in case of premature or ill infants [34]. However, several of them are occasionally proven to be inadequate in controlling disease activity and therefore bDMARDs should be used. During pregnancy the most widely used bDMARD is certolizumab pegol, a PEGylated, Fc-free anti-TNF agent with a high safety profile. In 1,137 prospectively reported pregnancies with maternal exposure to the drug, neither teratogenesis, nor increased rate of fetal death was found [35]. Data from gastroenterology indicate that adalimumab may also be a safe choice for pregnant patients with autoimmune diseases [37]. Additionally, there are no reports linking etanercept to unfavorable outcomes during pregnancy and lactation, indicating that etanercept may be administered, if necessary [39].

Congenital abnormalities are not referred to in cases of compatible TNFi administration during the first half of pregnancy [4]. However, there are several concerns about the potential affection of infants' immune system after birth, and consequently live vaccines are contraindicated for the first 12 months of life [38]. As far as the newer therapies with targeted synthetic therapies (ts-DMARDs or jakinh) are concerned, no clear evidence to this date exist regarding their safety during pregnancy. Due to their small size, these molecules are considered capable of crossing the placenta; therefore, their use is not indicated in pregnant women [40].

Environmental factors may also play an important role in unsuccessful treatment in RA. Smoking increases the risk of ACPA-positive RA [41]. Seropositive RA (as defined by positive ACPA and/ or RF in patients' serum)

is known to be more severe than seronegative disease and therefore more difficult to treat. The administration of the most widely used bDMARDs, such as TNFi, is negatively correlated with smoking [46]. Smoking is also associated with radiographic progression in RA. According to Rydell et al, other risk factors for rapid joint destruction are seropositivity, presence of erosions, and high disease activity at baseline; maintenance of active disease at one year after diagnosis is additionally considered dangerous for the development of novel erosions [47]. Therefore, cigarette smoking may be related to a more severe and often refractory disease, with limited response to treatment options.

Coffee consumption is also associated with higher risk for RA development and mainly for a seropositive, and consequently more difficult to treat, disease [42]. However, inflammatory responses are influenced by caffeine and its metabolite paraxanthine. Caffeine has been shown to suppress chemotaxis of monocytes and neutrophils, as well as the production of pro-inflammatory cytokines, including TNF α [49], indicating the existence of contradictory data concerning caffeine consumption in RA patients [4]. The lack of physical exercise may also exacerbate disease activity. Due to inflammatory mechanisms, lower bone mineral density and aggravation of joint mobility may occur [43]. More specifically, Th1 cell production is reported to be decreased after physical activity. IL 6 is also secreted inducing the production of anti-inflammatory cytokines, including IL 1 α , IL 10 and TNF receptor (TNF-R) [48]. Furthermore, lack of physical activity may also increase the cardiovascular risk. Patients with RA, due to uncontrolled systemic inflammation, are prone to atherosclerotic cardiovascular diseases [50], as already mentioned. Consequently, they are advised for a healthier way of living, including aerobic exercise, as well as strength and resistance training, as a main part of their therapy, according to EULAR recommendations published in 2018, concomitantly aiming at a better quality of life [44,45,51]. However, nowadays, the maintenance of good physical condition is limited by the new way of urban living. The concurrent long-term use of steroid agents may also easily lead to muscle weakness and osteoporosis [4]. Systemic inflammation, as observed in RA, is often associated with bone loss. Osteoclastic activity may also be activated, resulting in a reduced bone mineral density (BMD) [52]. Osteoporosis is estimated to be found in 30–50% of patients with RA, mainly in cases of high disease activity [53]. However, the prolonged use of corticosteroids, a major cornerstone

in the treatment of RA patients, may increase the risk for bone loss and, consequently, fractures. As a result, anti-osteoporotic treatment is also recommended in RA patients, contributing to polypharmacy and frequently poor treatment compliance [53,54].

Considering all the above, the use of several commonly preferred drugs for RA treatment, may be present challenges and, therefore, inevitably not prescribed for several patients. Comorbidities and adverse drug reactions may not be compatible with the administration of DMARDs, leading to refractory disease, difficult to treat [55]. Furthermore, lower adherence to treatment in RA is mainly observed in patients with higher disease activity. In addition, factors such as lower education level and worse financial situation of several patients further result in non-compliance and, therefore, poor disease outcomes [54]. In order to calculate disease activity, several tools are often used, taking into consideration the Patient Global Assessment (PGA), too. PGA is a widely used self-report measure reflecting patients' own assessment of the severity of their condition. However, since many versions of PGA world widely exist, its standardization remains a major problem, indicating the necessity for validity and reliability [56]. Since patients' lives are undoubtedly affected by the disease in every aspect of their everyday activities, the development for new patient – centered tools, capable of incorporating the symptoms and consequences is imposed.

Finally, the high cost of several treatment options (especially bDMARDs) may influence the physician's prescription. Rheumatologists should lean on robust data for better disease control, but financial obstacles may influence the therapy that is administered in their patients [61], especially in different regions with different economic laws.

CONCLUSION

Despite the treat-to-target strategy in combination with novel drugs in RA, some patients may still fail to respond. Difficult-to-treat RA remains a major problem in everyday clinical life of rheumatologists, indicating the need for early recognition of poor prognostic factors in order to avoid disease flares and hence therapy escalation, resulting in controversial outcomes. Refractory disease may require better management of comorbidities and environmental factors, which, combined with the different pathways, molecules, and cells involved in the pathogenesis of RA, may in part justify the lack of response of some patients.

Conflict of interest disclosure: None to declare.

Declaration of funding sources: None to declare.

Author contributions: AF was responsible for the conception, research, writing and the final draft of this review.

REFERENCES

- Huang J, Fu X, Chen X, Li Z, Huang Y, Liang C. Promising Therapeutic Targets for Treatment of Rheumatoid Arthritis. *Front Immunol*. 2021;12:686155.
- Prasad P, Verma S, Surbhi, Ganguly NK, Chaturvedi V, Mittal SA. Rheumatoid arthritis: advances in treatment strategies. *Mol Cell Biochem*. 2023;478(1):69-88.
- Alamanos Y, Drosos AA. Epidemiology of adult rheumatoid arthritis. *Autoimmun Rev*. 2005;4(3):130-6.
- Colignaro P, Triggianese P, de Martino E, Fonti GL, Chimenti MS, Sunzini F, et al. Challenges in the treatment of Rheumatoid Arthritis. *Autoimmun Rev*. 2019;18(7):706-13.
- Wijbrandts CA, Tak PP. Prediction of Response to Targeted Treatment in Rheumatoid Arthritis. *Mayo Clin Proc*. 2017;92(7):1129-43.
- Raychaudhuri S, Remmers EF, Lee AT, Hackett R, Guiducci C, Burt NP, et al. Genetic variants at CD28, PRDM1 and CD2/CD58 are associated with rheumatoid arthritis risk. *Nat Genet*. 2009;41(12):1313-8.
- Viatte S, Barton A. The role of rheumatoid arthritis genetic susceptibility markers in the prediction of erosive disease. *Eur Musculoskelet Rev*. 2012;7(2):102-7.
- Viatte S, Plant D, Han B, Fu B, Yarwood A, Thomson W, et al. Association of HLA-DRB1 haplotypes with rheumatoid arthritis severity, mortality, and treatment response. *JAMA*. 2015;313(16):1645-56.
- Ling SF, Viatte S, Lunt M, Van Staa TP, Fu B, Deighton C, et al. HLA-DRB1 amino acid positions 11/13, 71, and 74 are associated with inflammation level, disease activity, and the health assessment questionnaire score in patients with inflammatory polyarthritis. *Arthritis Rheumatol*. 2016;68(11):2618-28.
- de Rooy DP, Tsonaka R, Andersson ML, Forslind K, Svensson B, van der Helm-van Mil AH. Genetic factors for the severity of ACPA-negative rheumatoid arthritis in 2 cohorts of early disease: a genome-wide study. *J Rheumatol*. 2015;42(8):1383-91.
- Ciccacci C, Conigliaro P, Perricone C, Rufini S, Triggianese P, Politi C, et al. Polymorphisms in STAT-4, IL-10, PSORS1C1, PTPN2 and MIR146A genes are associated differently with prognostic factors in Italian patients affected by rheumatoid arthritis. *Clin Exp Immunol*. 2016;186(2):157-63.
- Schipper LG, Fransen J, den Broeder AA, Van Riel PL. Time to achieve remission determines time to be in remission. *Arthritis Res Ther*. 2010;12(3):R97.
- Conigliaro P, Triggianese P, Chimenti MS, Tonelli M, Perricone R. Factors predicting 2 years of remission and low disease activity in rheumatoid arthritis patients treated with TNF-

- inhibitors. *Isr Med Assoc J*. 2017;19(8):467-72.
14. Bazzichi L, Rossi P, Giacomelli C, Bombardieri S, De Feo F. A proposal of simple calculation (ERI calculator) to predict the early response to TNF- α blockers therapy in rheumatoid arthritis. *Rheumatol Int*. 2012;32(2):349-56.
 15. Larid G, Vix J, Garlantezec R, Loppin E, Gervais E. Increased remission with fewer corticosteroids and more biologics in rheumatoid arthritis at 7-year follow-up in real-life conditions. *Sci Rep*. 2022;12(1):2563.
 16. Arranz LI, Rafecas M, Alegre C. Effects of obesity on function and quality of life in chronic pain conditions. *Curr Rheumatol Rep*. 2014;16(1):390.
 17. Goossens J, Coustet B, Palazzo E, Dieudé P, Allanore Y, Ottaviani S. Overweight and obesity affect clinical assessment of synovitis in rheumatoid arthritis: comparison of ultrasonography and clinical exam. *Clin Exp Rheumatol*. 2019;37:49-54.
 18. Daïen CI, Sellam J. Obesity and inflammatory arthritis: impact on occurrence, disease characteristics and therapeutic response. *RMD Open*. 2015;1:e000012.
 19. Gremese E, Carletto A, Padovan M, Atzeni F, Raffeiner B, Giardina AR, et al. Obesity and reduction of the response rate to anti-tumor necrosis factor α in rheumatoid arthritis: an approach to a personalized medicine. *Arthritis Care Res (Hoboken)*. 2013;65(1):94-100.
 20. D'Agostino MA, Alten R, Mysler E, Bessette L, Alecock E, Woodworth T, et al. Body mass index and clinical response to intravenous or subcutaneous abatacept in patients with rheumatoid arthritis. *Clin Rheumatol*. 2017;36(12):2655-65.
 21. Iannone F, Courvoisier DS, Gottenberg JE, Hernández MV, Lie E, Canhão H, et al. Body mass does not impact the clinical response to intravenous abatacept in patients with rheumatoid arthritis: Analysis from the pan-European registry collaboration for abatacept (PANABA). *Clin Rheumatol*. 2017;36(4):773-9.
 22. Gardette A, Ottaviani S, Sellam J, Meyer O, Dieudé P. Body mass index and response to tocilizumab in rheumatoid arthritis: a real life study. *Clin Rheumatol*. 2016;35(4):857-61.
 23. Kanda T, Takahashi T. Interleukin-6 and Cardiovascular Disease. *Jpn Heart J*. 2004;45(2):183-93.
 24. Cavagna L, Boffini N, Cagnotto G, Inverardi F, Grosso V, Caporali R. Atherosclerosis and rheumatoid arthritis: more than a simple association. *Mediators Inflamm*. 2012;2012:147354.
 25. Delcoigne B, Provan SA, Kristianslund EK, Askling J, Ljung L. How does current disease activity in rheumatoid arthritis affect the short-term risk of acute coronary syndrome? A clinical register based study from Sweden and Norway. *Eur J Intern Med*. 2023;115(1):55-61.
 26. Galloway JB, Hyrich KL, Mercer LK, Dixon WG, Fu B, Usitanowski AP, et al. Anti-TNF therapy is associated with an increased risk of serious infections in patients with rheumatoid arthritis especially in the first 6 months of treatment: updated results from the British Society for Rheumatology biologics register with special emphasis on risks in the elderly. *Rheumatology (Oxford)*. 2011;50(1):124-31.
 27. Ballanti E, Conigliaro P, Chimenti MS, Kroegler B, Di Muzio G, Guarino MD, et al. Use of anti-tumor necrosis factor alpha therapy in patients with concurrent rheumatoid arthritis and hepatitis B or hepatitis C: a retrospective analysis of 32 patients. *Drug Dev Res*. 2014;75(Suppl 1):S42-5.
 28. Au K, Reed G, Curtis JR, Kremer JM, Greenberg JD, Strand V, et al. High disease activity is associated with an increased risk of infection in patients with rheumatoid arthritis. *Ann Rheum Dis*. 2011;70(5):785-91.
 29. Jethwa H, Lam S, Smith C, Giles I. Does rheumatoid arthritis really improve during pregnancy? A systematic review and meta-analysis. *J Rheumatol*. 2018;45(2):180-226.
 30. de Man YA, Dolhain RJ, Van de Geijn FE, Willemsen SP, Hazes JM. Disease activity of rheumatoid arthritis during pregnancy: results from a nationwide prospective study. *Arthritis Rheum*. 2008;59(9):1241-8.
 31. Weber-Schoendorfer C, Diav-Citrin O. Safety of mycophenolate during pregnancy. *RMD Open*. 2023;9:e002899.
 32. Singh S, Sedgh G, Hussain R. Unintended pregnancy: worldwide levels, trends, and outcomes. *Stud Fam Plann*. 2010;41(4):241-50.
 33. Gotestam Skorpen C, Hoeltzenbein M, Tincani A, Fischer-Betz R, Elefant E, Chambers C, et al. The EULAR points to consider for use of antirheumatic drugs before pregnancy, and during pregnancy and lactation. *Ann Rheum Dis*. 2016;75(5):795-810.
 34. Østensen M, Khamashta M, Lockshin M, Parke A, Brucato A, Carp H, et al. Anti-inflammatory and immunosuppressive drugs and reproduction. *Arthritis Res Ther*. 2006;8(3):209.
 35. Weber-Schoendorfer C, Chambers C, Wacker E, Beghin D, Bernard N, Shechtman S, et al. Pregnancy outcome after methotrexate treatment for rheumatic disease prior to or during early pregnancy: a prospective multicenter cohort study. *Arthritis Rheumatol*. 2014;66(5):1101-10.
 36. Clowse MEB, Scheuerle AE, Chambers C, Afzali A, Kimball AB, Cush JJ, et al. Pregnancy Outcomes After Exposure to Certolizumab Pegol: Updated Results From a Pharmacovigilance Safety Database. *Arthritis Rheumatol*. 2018;70(9):1399-1407.
 37. Chambers CD, Johnson DL, Xu R, Luo Y, Lopez-Jimenez J, Adam MP, et al. Birth outcomes in women who have taken adalimumab in pregnancy: A prospective cohort study. *PLoS One*. 2019;14(10):e0223603.
 38. De Felice KM, Kane S. Safety of anti-TNF agents in pregnancy. *J Allergy Clin Immunol*. 2021;148(3):661-7.
 39. Nishide M, Yagita M, Kumanogoh A. Continuous Use of Etanercept During Pregnancy Does Not Affect TNF-Alpha Levels in Umbilical Cord Blood. *Biologics*. 2022;16:17-9.
 40. Wieringa JW, van der Woude CJ. Effect of biologicals and JAK inhibitors during pregnancy on health-related outcomes in children of women with inflammatory bowel disease. *Best Pract Res Clin Gastroenterol*. 2020;44-45:101665.
 41. Klareskog L, Stolt P, Lundberg K, Källberg H, Bengtsson C, Grunewald J, et al. A new model for an etiology of rheumatoid arthritis: smoking may trigger HLA-DR (shared epitope)-restricted immune reactions to autoantigens modified by citrullination. *Arthritis Rheum*. 2006;54(1):38-46.
 42. Lee YH, Bae SC, Song GG. Coffee or tea consumption and

- the risk of rheumatoid arthritis: a meta-analysis. *Clin Rheumatol*. 2014;33(11):1575-83.
43. Sharif K, Watad A, Bragazzi NL, Lichtbroun M, Amital H, Shoenfeld Y. Physical activity and autoimmune diseases: get moving and manage the disease. *Autoimmun Rev*. 2018;17(1):53-72.
 44. Curtis GL, Chughtai M, Khlopas A, Newman JM, Khan R, Shaffiy S, et al. Impact of physical activity in cardiovascular and musculoskeletal health: can motion be medicine? *J Clin Med Res*. 2017;9(5):375-81.
 45. Rausch Osthoff AK, Niedermann K, Braun J, Adams J, Brodin N, Dagfinrud H, et al. 2018 EULAR recommendations for physical activity in people with inflammatory arthritis and osteoarthritis. *Ann Rheum Dis*. 2018;77(9):1251-60.
 46. Abhishek A, Butt S, Gadsby K, Deighton C. Anti-TNF-alpha agents are less effective for the treatment of rheumatoid arthritis in current smokers. *J Clin Rheumatol*. 2010;16(1):15-8.
 47. Rydell E, Forslind K, Nilsson JA, Jacobsson LT, Turesson C. Smoking, body mass index, disease activity, and the risk of rapid radiographic progression in patients with early rheumatoid arthritis. *Arthritis Res Ther*. 2018;20(1):82.
 48. Rochette E, Duché P, Merlin E. Juvenile idiopathic arthritis and physical activity: possible inflammatory and immune modulation and tracks for interventions in young populations. *Autoimmun Rev*. 2015;14(8):726-34.
 49. Horrigan LA, Kelly JP, Connor TJ. Immunomodulatory effects of caffeine: friend or foe? *Pharmacol Ther*. 2006;111(3):877-92.
 50. Hansildaar R, Vedder D, Baniaamam M, Tausche AK, Gerritsen M, Nurmohamed MT. Cardiovascular risk in inflammatory arthritis: rheumatoid arthritis and gout. *Lancet Rheumatol*. 2021;3(1):e58-70.
 51. Rausch Osthoff AK, Niedermann K, Braun J, Adams J, Brodin N, Dagfinrud H, et al. 2018 EULAR recommendations for physical activity in people with inflammatory arthritis and osteoarthritis. *Ann Rheum Dis*. 2018;77(9):1251-60.
 52. Shimizu T, Takahata M, Kimura-Suda H, Kameda Y, Endo T, Hamano H, et al. Autoimmune arthritis deteriorates bone quantity and quality of periarticular bone in a mouse model of rheumatoid arthritis. *Osteoporos Int*. 2016;27(2):709-18.
 53. Ashai S, Harvey NC. Rheumatoid arthritis and bone health. *Clin Med (Lond)*. 2020;20(6):565-7.
 54. Alhefny AE, Abd El-Rahman MA, Abd El-Moteleb S, Shedid NH, Sakr HM, Hassan RM. Evaluation of Adherence to Drug Treatment in Patients with Rheumatoid Arthritis. *Egypt J Rheumatol Clin Immunol*. 2016;3(1):68-80.
 55. Nagy G, Roodenrijs NM, Welsing PM, Kedves M, Hamar A, van der Goes MC, et al. EULAR definition of difficult-to-treat rheumatoid arthritis. *Ann Rheum Dis*. 2021;80(1):31-5.
 56. Ferreira RJ, de Wit M, Henriques M, Pinto AF, Duarte C, Mateus E, et al. 'It can't be zero!' Difficulties in completing patient global assessment in rheumatoid arthritis: a mixed methods study. *Rheumatology (Oxford)*. 2020;59(5):1137-47.
 57. Rosa JE, García MV, Luissi A, Pierini F, Sabelli M, Mollerach F, et al. Rheumatoid Arthritis Patient's Journey: Delay in Diagnosis and Treatment. *J Clin Rheumatol*. 2020;26(7S):S148-52.
 58. Ytterberg SR, Bhatt DL, Mikuls TR, Koch GG, Fleischmann R, Rivas JL, et al. Cardiovascular and Cancer Risk with Tofacitinib in Rheumatoid Arthritis. *N Engl J Med*. 2022;386(4):316-26.
 59. Smolen JS, Landewé RBM, Bergstra SA, Kerschbaumer A, Sepriano A, Aletaha D, et al. EULAR recommendations for the management of rheumatoid arthritis with synthetic and biological disease-modifying antirheumatic drugs: 2022 update. *Ann Rheum Dis*. 2023;82(1):3-18.
 60. Karpouzas GA, Szekanecz Z, Baecklund E, Mikuls TR, Bhatt DL, Wang C, et al. Rheumatoid arthritis disease activity and adverse events in patients receiving tofacitinib or tumor necrosis factor inhibitors: a post hoc analysis of ORAL Surveillance. *Ther Adv Musculoskelet Dis*. 2023;15:1759720X231201047.
 61. Hresko A, Lin TC, Solomon DH. Medical Care Costs Associated With Rheumatoid Arthritis in the US: A Systematic Literature Review and Meta-Analysis. *Arthritis Care Res (Hoboken)*. 2018;70(10):1431-8.
 62. Abasolo L, Judez E, Descalzo MA, Gonzalez-Alvaro I, Jover JA, Carmona L, et al. Cancer in rheumatoid arthritis: occurrence, mortality, and associated factors in a South European population. *Semin Arthritis Rheum*. 2008;37(6):388-97.
 63. Chen YJ, Chang YT, Wang CB, Wu CY. The risk of cancer in patients with rheumatoid arthritis: a nationwide cohort study in Taiwan. *Arthritis Rheum*. 2011;63(2):352-8.
 64. Simon TA, Thompson A, Gandhi KK, Hochberg MC, Suissa S. Incidence of malignancy in adult patients with rheumatoid arthritis: a meta-analysis. *Arthritis Res Ther*. 2015;17(1):212.
 65. Shacter E, Weitzman SA. Chronic inflammation and cancer. *Oncology (Williston Park)*. 2002;16(2):217-26.
 66. Cho SK, Lee J, Han M, Bae SC, Sung YK. The risk of malignancy and its incidence in early rheumatoid arthritis patients treated with biologic DMARDs. *Arthritis Res Ther*. 2017;19(1):277.
 67. Beydon M, Pinto S, De Rycke Y, Fautrel B, Mariette X, Seror R, et al. Risk of cancer for patients with rheumatoid arthritis versus general population: a national claims database cohort study. *Lancet Reg Health Eur*. 2023;35:100768.
 68. Chatzidionysiou K, Delcoigne B, Frisell T, Hetland ML, Hauge EM, Nordström D, et al. How do we use biologics in rheumatoid arthritis patients with a history of malignancy? An assessment of treatment patterns using Scandinavian registers. *RMD Open*. 2020;6(1):e001363.
 69. Burmester GR, Cohen SB, Winthrop KL, Nash P, Irvine S, De Leonardi F, et al. Safety profile of upadacitinib over 15 000 patient-years across rheumatoid arthritis, psoriatic arthritis, ankylosing spondylitis and atopic dermatitis. *RMD Open*. 2023;9(1):e002735.
 70. Raaschou P, Simard JF, Holmqvist M, Askling J. Tumor necrosis factor inhibitors and cancer recurrence in Swedish patients with rheumatoid arthritis: A nationwide population-based cohort study. *Ann Intern Med*. 2018;169(5):291-9.
 71. Thomas K, Vassilopoulos D. Infections in Patients with Rheumatoid Arthritis in the Era of Targeted Synthetic Therapies. *Mediterr J Rheumatol*. 2020;31(Suppl 1):129-36.
 72. Wolfe F, Caplan L, Michaud K. Treatment for rheumatoid arthritis and the risk of hospitalization for pneumonia: asso-

- ciations with prednisone, disease-modifying antirheumatic drugs, and anti-tumor necrosis factor therapy. *Arthritis Rheum.* 2006;54(2):628-34.
73. Ozen G, Pedro S, England BR, Mehta B, Wolfe F, Michaud K. Risk of Serious Infection in Patients With Rheumatoid Arthritis Treated With Biologic Versus Nonbiologic Disease-Modifying Antirheumatic Drugs. *ACR Open Rheumatol.* 2019;1(7):424-32.
74. Zink A, Manger B, Kaufmann J, Eisterhues C, Krause A, Listing J, et al. Evaluation of the RABBIT Risk Score for serious infections. *Ann Rheum Dis.* 2014;73(9):1673-6.
75. Vander Cruyssen B, Peene I, Cantaert T, Hoffman IEA, De Rycke L, Veys EM, et al. Anti-citrullinated protein/peptide antibodies (ACPA) in rheumatoid arthritis: Specificity and relation with rheumatoid factor. *Autoimmun Rev.* 2005;4(7):468-74.

Corresponding author:

Alexandra Filippopoulou MD, PhD
Rheumatologist. 12, Eleftheriou Venizelou St.,
Amaliada Ileia, Greece, E-mail: alexfilippop@yahoo.com

Immunotherapy - Induced Colitis

Gerasimia-Marina Chardalia, Angelos Koutras

Abstract

The introduction of Immune Checkpoint Inhibitors (ICIs) entirely altered the landscape of the oncologic therapeutic approach. The immunotherapy agents administered to date have been proven to be efficient and improve overall survival even in advanced malignancies, whose prognosis was poor before the emergence of ICIs. Nevertheless, the immune system's overresponsive activity may lead to toxic consequences, gastrointestinal adverse events being the most frequently reported. Their clinical manifestations range from mild, self-restricted diarrhea to fatal complications such as intestinal perforation. Few studies have been conducted to detect the susceptibility factors leading to the development of immune-checkpoint inhibitor-mediated colitis (IMC) and clinicians remain incapable of identifying high-risk patients in time. Management of IMC principally includes corticosteroids, showing favorable outcomes and remission in a significant percentage of patients. However, refractory disease cases still require biologic agents such as Infliximab or Vedolizumab. This review aims to highlight the clinical importance of diagnosing IMC promptly and treating it according to the latest evidence-based guidelines, while discussing promising management approaches.

Key words: *Immune checkpoint inhibitors; diarrhea; colitis; immune-related adverse events*

INTRODUCTION

Immunity checkpoint inhibitors (ICIs) have dramatically transformed the current treatment approach of malignancies. Since their incorporation into oncology, there has been a revolution in the management of cancers of all grades. The improvement of both overall survival and quality of life for patients represents a significant scientific breakthrough of the 21st century [1]. It is well acknowledged that tumor cells employ mechanisms to escape the immunosurveillance system. Hence, scientists developed the ICIs, monoclonal antibodies targeting three critical molecules of this process: the cytotoxic T-lymphocyte-associated antigen-4 (CTLA-4), the programmed cell death protein-1 (PD-1) and its ligand PD-L1 (PD-L1). These proteins consist of inhibitory receptors of the immune system, suppressing the T-cell-mediated activity against the tumor. Consequently, the ICIs, by blocking these receptors, lead to enhanced T cell activity

in order to eliminate the tumor cells [2].

The introduction of immunotherapy as an additional tool against cancer has not only resulted in favorable outcomes, but has also posed challenges. Unfortunately, the overresponse of T cells has a potentially harmful effect on almost every healthy tissue, imitating the entities of autoimmune diseases, known as immune-related adverse events (irAEs) [3]. One of the most common irAEs implicates the gastrointestinal tract, with clinical presentation ranging from diarrhea to severe colitis leading even to fatal perforation [4]. Its prevalence in the population receiving ICIs ranges from 1 to 25%, depending on several parameters [5]. Wang et al. [6], in their meta-analysis, reported that the incidence of colitis across all grades varies depending on the type of immunotherapy administered. More specifically, the percentage of patients who presented ICIs-induced colitis was 1.4% for anti-PD1 agents, 1% for anti-PDL1 agents, and 9.1% for anti-CTLA4 agents. It is noteworthy that the combination of two types of agents, Nivolumab and Ipilimumab, is linked to even higher incidence [7].

This review aims to highlight the importance of

promptly diagnosing Immune-checkpoint inhibitor-mediated colitis (IMC), differentiating it from other forms of colitis and effectively managing it according to the latest guidelines.

Pathophysiology

The precise mechanism underlying IMC remains obscure. Nevertheless, three events play a pivotal role in its pathology: the excessive activation of T effector cells (Teff), the lymphocyte infiltration of the intestinal mucosa, and the increased circulation of T memory cells [8-10].

CTLA-4 is a critical checkpoint inducing the suppressive function of CD4+ regulatory T cells (Tregs) [11]. Hence, the inhibition of CTLA-4 may contribute to the development of autoimmune diseases such as IMC. In addition, Luoma et al. [12] proposed that one of the initiating events of IMC is the conversion of resident CD8+ T memory cells in the intestinal mucosa into cytotoxic effector T cells (CTLs), overproducing inflammatory cytokines such as IL-17, IFN- γ , and TNF- α . Furthermore, the upregulation of chemokine receptors CXCR3, 6, 9/10, 16, and integrin receptors $\alpha 4\beta 7/\alpha E\beta 7$ appears to enhance the T cell activity [13].

The role of gut microbiota is also in the spotlight of researchers of irAEs as an imbalance in the intestinal mucosa's flora can affect both the patient's response to immunotherapy and severity of IMC [14]. In most patients receiving ICIs, species such as *Bacteroides fragilis*, *Burkholderiales*, and *Lactobacillus reuteri* are decreased, while bacteria from the *Faecalibacterium prausnitzii* genus are enriched [13]. This dysbiosis in gut microbiota may play a predictive role in patients' antitumor response and their likelihood of developing IMC.

Risk factors

Since the emergence and recognition of the IMC entity as an irAE, researchers have sought to identify the main parameters defining the level of patient's susceptibility in developing IMC. As mentioned above, the type of immunotherapy agents administered must be considered as a risk factor. CTLA4 inhibitors are associated with a higher incidence of colitis compared to both anti-PD1 and anti-PDL1 agents [6]. Notably, Ascierto et al. [15], in their randomized, double-blind, multicenter study, concluded that patients with advanced melanoma who received higher doses of Ipilimumab were more susceptible to developing colitis in comparison with the patient group receiving lower Ipilimumab

doses. Hence, this finding establishes Ipilimumab, an anti-CTLA4 factor, as the only demonstrating dose-dependent predisposition to colitis.

In the largest study examining the epidemiology of IMC, Farha et al. [16] were the first to identify the characteristics associated with increased susceptibility to developing IMC. Accordingly, it was discussed that female gender, Caucasian ancestry, alcoholism, obesity, and age over 65 years are the primary underlying risk factors to consider when administering immune checkpoint inhibitors. Furthermore, the same cohort study demonstrated that patients with a background of autoimmune disease have an elevated risk of developing colitis. The presence of preexisting autoimmune disease in patients with IMC was found to be 24.6%, with rheumatoid arthritis being the most prevalent, affecting 9.8% of the population studied. In addition, on a genetic basis, HLA-DQB1*03:01 was found to be associated with gastrointestinal adverse events [17].

Patients with a history of Inflammatory Bowel Disease (IBD) have a significantly higher median risk of developing IMC compared to those without an IBD background (41% vs 11%, respectively; $P < .001$) [18]. Therefore, clinicians should maintain a high level of suspicion in such cases but always consider the differential diagnosis with an exacerbation of IBD, as the clinical manifestations may overlap [19].

Clinical presentation and differential diagnosis

Based on clinical manifestations, the National Cancer Institute's Common Terminology Criteria for Adverse Events (CTCAE) provides classification systems for both diarrhea and colitis in order to assess disease severity and guide proper management (Table 1) [20]. The classification of diarrhea is based on the number of daily bowel movements above the patient's baseline. Diarrhea is often the first alarming sign of IMC and requires thorough examination [21]. Specifically, Grade 1 diarrhea is defined as fewer than four stools above baseline per day, while Grade 2 is characterized by four to six additional stools per day above baseline. Severe cases where the number of stools exceeds seven above baseline are classified as Grade 3. Grade 4 indicates life-threatening conditions requiring urgent intervention, such as hemodynamic collapse. Finally, both diarrhea and colitis classifications designate Grade 5 as death.

Regarding the classification of colitis, Grade 1 corresponds to asymptomatic patients, while Grade 2 refers to patients with a recent history of or currently receiving

Table 1. The CTCAE classification of diarrhea and colitis [20].

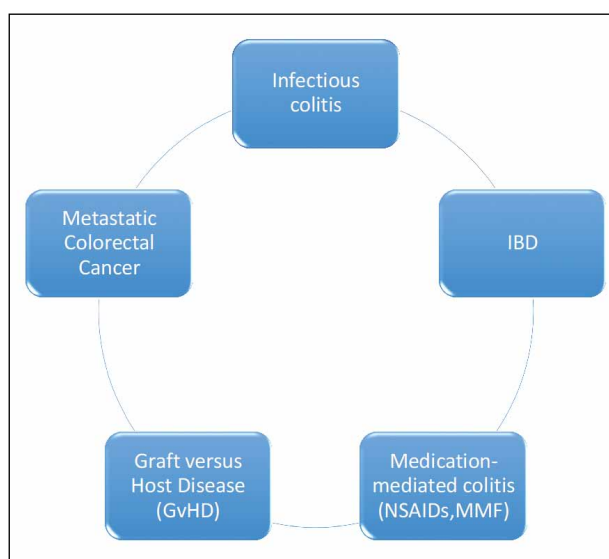
CLASSIFICATION	DIARRHEA	COLITIS
Grade 1	Increase of <4 stools per day over baseline; mild increase in ostomy output compared to baseline	Asymptomatic, pathologic or radiographic findings only
Grade 2	Increase of 4 – 6 stools per day over baseline; moderate increase in ostomy output compared to baseline	Abdominal pain; mucus or blood in stool
Grade 3	Increase of ≥7 stools per day over baseline; severe increase in ostomy output compared to baseline	Abdominal pain, fever, change in bowel habits with ileus; peritoneal signs
Grade 4	Life-threatening consequences (e.g., hemodynamic collapse)	Life-threatening consequences (e.g., perforation, bleeding, ischemia, necrosis, toxic megacolon)
Grade 5	Death	Death

immunotherapy who present with abdominal pain, and bloody or mucousy diarrhea. Concomitant presentations that raise suspicion of IMC include fever, severe pain, signs of peritoneal irritation (Grade 3), or even life-threatening conditions (Grade 4) such as perforation, ischemia, bleeding, necrosis, and toxic megacolon [19]. Additionally, common clinical manifestations include hematochezia, vomiting, and nausea, while patients may also report less typical symptoms such as weight loss or loss of appetite [13]. Furthermore, IMC may also involve upper gastrointestinal symptoms, including dyspepsia, heartburn, and regurgitation. Clinicians must be mindful of the potential coexistence of immunotherapy-induced gastritis alongside IMC [22].

The diagnosis of IMC is based on exclusion, as other forms of colitis or clinical entities (Figure 1) may present with the same manifestations and may even share comparable laboratory, imaging, or histological findings [23].

Diagnosis

Clinical manifestations suggestive of IMC should prompt clinicians to undertake a thorough investigation to establish a timely diagnosis. A detailed patient history and physical examination play a crucial role in the initial assessment and in determining the grade of colitis. Moreover, a complete blood count and biochemical panel should be performed, as along with a measurement of C-reactive protein (CRP) and Erythrocyte Sedimentation Rate (ESR) [24]. Infectious causes of colitis should be ruled out from the beginning, and therefore, it is recommended to obtain stool cultures for detecting pathogens such as *Clostridioides difficile*. It may also be prudent to conduct a supplemental viral

**Figure 1.** Differential Diagnosis of Immune-checkpoint inhibitor-mediated colitis (IMC) [23].

check and parasite test in patients residing in regions with a high prevalence of viruses and parasites known to cause colitis. Further investigation could include stool inflammatory markers such as lactoferrin and calprotectin. The American Gastroenterological Association (AGA) recommends the detection of these markers as they identify patients with Grade 1 colitis as being at high-risk for further endoscopic assessment. Regarding Grade 2 or higher colitis, their detection is strongly recommended, though not strictly required, before proceeding to endoscopy [25]. Furthermore, exocrine pancreatic insufficiency is a rare complication of ICIs, but it must still be excluded from the differential

diagnosis by measuring fecal pancreatic elastase [26]. Additionally, the possibility of new-onset celiac disease following immunotherapy should be considered, and tissue transglutaminase immunoglobulin A (IgA) and total IgA tests should be ordered [27].

Endoscopic assessment through colonoscopy or sigmoidoscopy is the gold standard for diagnosing IMC. In patients receiving Ipilimumab, sigmoidoscopy could be considered a first-choice diagnostic procedure because the rectum and the sigmoid colon are the principal colon segments affected [10]. Random biopsies should always be performed, as 33% of patients with IMC undergoing colonoscopy show normal-appearing intestinal mucosa [28]. This subgroup of patients tends to have better clinical outcomes and a lower risk of developing refractory disease or biologic therapies [29]. Abnormal endoscopic findings may include loss of vascular patterns, erythema, ulcerations, edema, necrosis and friable mucosa, which are associated with a worse prognosis [30]. Furthermore, the biopsies obtained can be used to rule out microscopic colitis or infectious colitis caused by *Clostridioides difficile* or cytomegalovirus (CMV) [31]. Notably, endoscopy plays a multi-dimensional role in the management of IMC. Clinicians should be aware of indications for repeat endoscopy applied in Grade 2 and above, such as lack of response to immunosuppressive treatment, deterioration of symptoms despite initial improvement, and evaluation of the intestinal mucosa before resuming immunotherapy agents following temporary discontinuation [5].

Computed tomography (CT) has also proven useful in assessing probable IMC cases, especially if there is a high clinical suspicion index of life-threatening events such as toxic megacolon or perforation, which necessitate immediate surgical intervention [32]. Findings such as diffuse or segmental colitis patterns on CT cannot establish a definite diagnosis of IMC as CT is characterized by low sensitivity and a high rate of false negatives [33]. Consequently, in early-stage patients, combining sigmoidoscopy with CT may be beneficial [32].

Histopathologic findings from the biopsies obtained are suggestive of the diagnosis of IMC. Remarkably, histological differences have been noted depending on the ICI administered. For instance, CTLA-4-associated colitis is linked to lamina propria expansion and lymphocytic intraepithelial infiltration, along with cryptic apoptosis or occasional prevalence of eosinophils in the lamina propria [34]. Concurrently, the lamina propria expansion by lymphoplasmacytic infiltration may be a histologic

feature of anti-PD1/anti-PDL1-associated colitis. In addition, neutrophils are the prominent inflammatory cells found, while they can also form cryptic abscesses [35,36]. Ischemic or collagenous colitis have also been described [37]. Most of the patients' biopsies, in both CTLA-4 and anti-PD1/anti-PDL1-induced colitis, show acute inflammatory changes that are indicative of IMC, while chronic inflammatory alterations of the intestinal submucosal tissue are found in approximately 50% of patients [33,37]. However, it is noteworthy that these histologic findings are not pathognomonic for IMC and have been described in other clinical entities that mimic IMC, such as infectious colitis, IBD, and GvHD [38]. Consequently, the final diagnosis must be made by integrating the patient's history, physical examination, laboratory work-up, and both endoscopic and histopathologic findings [32].

Management

Early-stage colitis, assessed as Grade 1, is typically managed conservatively. Specifically, liquid supplementation is imperative in case of dehydration and electrolyte imbalances, which should be corrected. Anti-diarrheal agents such as loperamide may be used, but they are only approved for patients with Grade-1 colitis [39]. Foods containing a high percentage of fat and lactose should be avoided in order to prevent secondary lactose intolerance or deterioration of the disease [40]. Moreover, the immunotherapy agent may be temporarily ceased or, depending on the patient's tolerance, continued. At this stage of IMC, discontinuing ICIs is not usually considered. However, this decision should be made after a multidisciplinary discussion involving oncologists and gastroenterologists, who will assess the patient's performance status, comorbidities, and clinical presentation [41]. Persisting Grade-1 colitis for more than 14 days may lead to the addition of oral budesonide [42], which has been shown to improve colitis symptoms and prevent the immunotherapy discontinuation [43]. Despite its efficacy, budesonide was tested in two randomized trials as possible prophylactic treatment for patients receiving Ipilimumab, but no statistically significant benefit was demonstrated [44,45]. Systemic corticosteroids could be an alternative option in persisting colitis of Grade 1, but clinicians should assess their effectiveness within the first few days of administration and, in case of a positive reciprocation, taper them throughout four to six weeks [46]. Furthermore, patients should be evaluated for stool

inflammatory markers in order to determine whether they are low- or high-risk, with the latter requiring assessment with an endoscopy [25].

The therapeutic management of Grade-2 IMC differs as systemic corticosteroids intravenously (IV) should be initiated from the outset (preferably prednisone or methylprednisolone in doses of 1-2 mg/kg/day). If improvement is noted after introducing IV corticosteroids, the patient may transition to oral corticosteroids [47]. Abdominal CT and endoscopy should always be performed for further evaluation [13]. In addition, patients receiving CTLA4 inhibitors should likely discontinue treatment permanently, while anti-PD1 and anti-PDL1 agents may be temporarily withheld and reintroduced when clinical improvement is achieved [48]. Biologic agents such as Infliximab, an anti-TNF α antibody or Vedolizumab, an integrin blocker, can be administered in case of corticosteroid refractory disease, whose incidence may reach 40% [49]. Early initiation of such biologic agents typically leads to better results, including fewer hospitalizations, earlier disease remission and reduced corticosteroid requirements [50].

Grade-3 colitis necessitates hospitalization of all patients, with close monitoring of hydration level and electrolytes. Therapeutic management does not differ significantly from Grade 2. Discontinuation of CTLA-4 inhibitors is mandatory, while anti-PD1/PDL-1 agents can be temporarily withheld. Systemic corticosteroids should be administered intravenously at high doses but if there is no immediate clinical improvement (after three to five days), escalation therapy with the biologic agents discussed should be initiated [39]. The recommended dosages are the same as those used in IBD patients: 5 mg/kg for Infliximab and 300mg for Vedolizumab [51]. The responsiveness after the biologic agent's infusion is typically rapid within the first week [52]. Notably, a single infusion may be adequate for complete remission of clinical manifestations with either Infliximab or Vedolizumab used, though there is a higher risk of relapse compared to 3 infusions [50]. Furthermore, there is currently no evidence to suggest whether either of the two biologic factors predominate. Clinicians should decide based on the availability or any contraindications that may arise [53].

Grade-4 colitis is managed similarly to Grade 3; however, in this case, immunotherapy agents are permanently discontinued regardless of the factor administered and reintroduction is not reconsidered (Table 2). CT imaging must be performed as life-threatening

events such as intestinal necrosis, perforation or toxic megacolon may have occurred, potentially requiring further intervention or surgery [19].

According to the treatment algorithm of IMC (Figure 2), biologic agents, particularly Infliximab and Vedolizumab, play an indispensable role in the therapeutic arsenal considering their efficacy [54]. Besides their role as an escalation therapy, the multidisciplinary modified Delphi consensus [55] recommended their introduction as a first-line treatment in patients with high-risk endoscopic findings, including large ulcerations and extensive colitis [50]. Furthermore, in case of no significant clinical improvement after administering one of these two commonly used biologic agents, the introduction of the other is strongly recommended [24]. Nonetheless, in case of Infliximab and Vedolizumab refractory disease the American Society of Clinical Oncology (ASCO) suggests the introduction of Tofacitinib, a Janus kinase (JAK) inhibitor or Ustekinumab, an antibody blocking interleukin-12 (IL-12) [56].

Management of IMC can be particularly challenging for clinicians, as refractory disease is common, even with the use of biologic agents [57]. In response, researchers are conducting trials to discover alternative and more effective treatments. Specifically, the correlation between gut microbiome composition and the IMC, as well as the influence of gut microbiota on the efficacy of ICIs against tumor cells, has led scientists to explore the potential of fecal microbiota transplantation (FMT) [58-62]. The first two patients receiving FMT due to refractory IMC were reported by Wang et al. in 2018 [62]. Notably, the second patient underwent two cycles of FMT to achieve complete remission of colitis. Since these

Table 2. ASCO and NCCN Guidelines on discontinuation rules of ICIs [46,56].

Grade 1	No consideration of discontinuation.
Grade 2	Discontinuation of ICIs. A resume may be considered after the improvement of symptoms to Grade 1 or less, completion of steroid tapering, review of risks and benefits and achievement of endoscopic/histologic remission.
Grade 3	Same principles as in Grade 2. If CTLA-4 was the ICI agent previously administered, permanent discontinuation is strongly recommended. A resume with an anti-PD-1/PD-L1 agent after the resolution of toxicity may be considered.
Grade 4	Permanent discontinuation of ICIs.

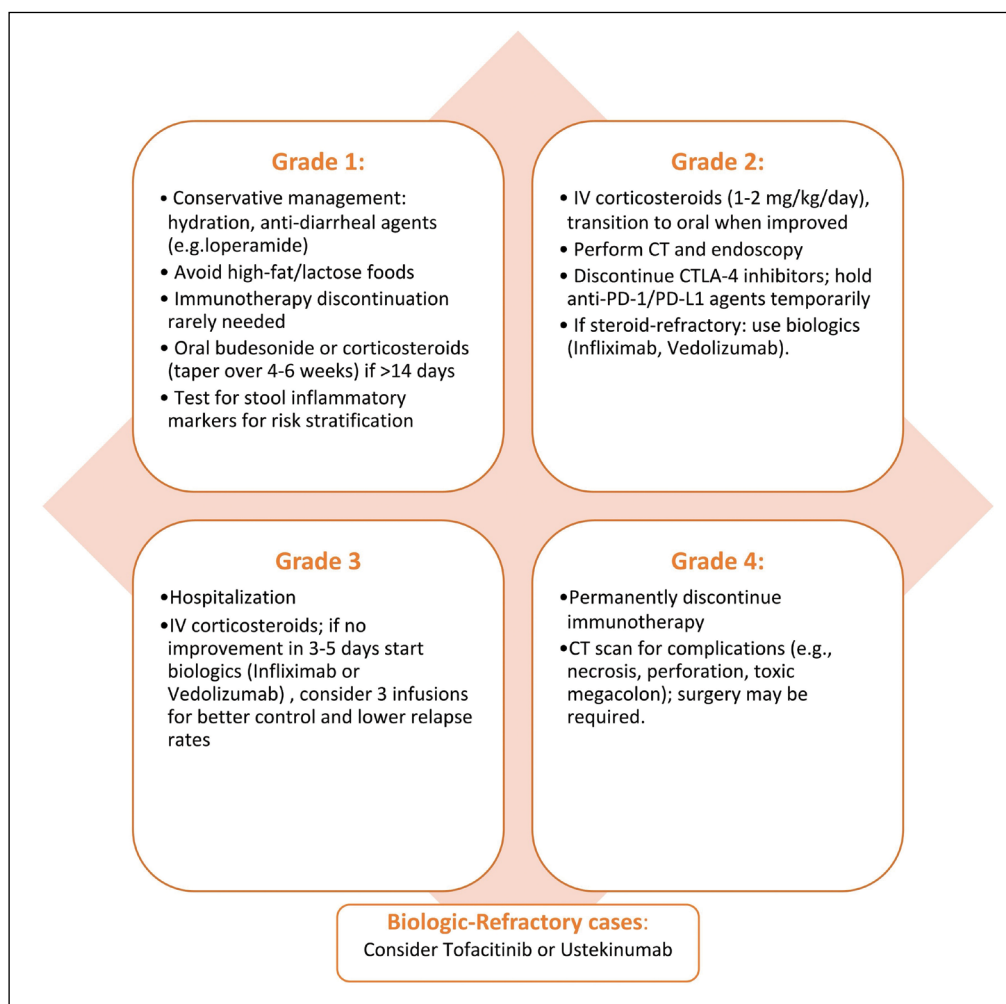


Figure 2. Practical Guide for Immune-checkpoint inhibitor-mediated colitis (IMC) management [46,56].

initial case reports, other researchers have applied FMT in refractory cases with favorable outcomes, rendering it an extremely promising treatment option [63-65].

CONCLUSION

Checkpoint inhibitors are among the most revolutionary oncologic treatments to date. However, they can result in various forms of toxicity, with IMC being one of the most frequently encountered adverse events. Clinicians must be aware that this side effect can arise even months after immunotherapy has been discontinued. Evaluating patients' risk factors is essential for assessing susceptibility to developing IMC. Clinical manifestations are nonspecific, as many pathologies mimic IMC, necessitating a thorough investigation, including laboratory workup, imaging and endoscopic procedures. The management of these patients must be meticulous, with

collaboration between oncologists and gastroenterologists being essential. Stratifying patients and defining the colitis grade is the first step in managing this irAE. Corticosteroids remain the cornerstone of treatment, but unfortunately, the rate of steroid-refractory disease is high. Hence, the use of biologic agents plays a crucial role and is associated with favorable outcomes. However, despite their efficacy, some patients fail to respond to this approach, prompting researchers to introduce new treatments aimed at achieving complete disease remission. Notably, FMT is one of the most promising alternatives, but further trials are needed to embed it as a safe and effective treatment in the management guidelines algorithm. Consequently, IMC is an adverse event that significantly affects a patient's quality of life, requiring prompt diagnosis and evidence-based management to ensure optimal remission.

Conflict of interest disclosure: None to declare.

Declaration of funding sources: None to declare

Author contributions: Conception and Design, AK, GMC; Analysis and Interpretation of the data, GMC; Drafting of the article, GMC; Critical revision of the article for important intellectual content, AK; Final approval, AK.

REFERENCES

- Gonzalez BD, Eisel SL, Bowles KE, Hoogland AI, James BR, Small BJ, et al. Meta-Analysis of Quality of Life in Cancer Patients Treated With Immune Checkpoint Inhibitors. *J Natl Cancer Inst.* 2020;112(6):808–18.
- Hargadon KM, Johnson CE, Williams CJ. Immune checkpoint blockade therapy for cancer: An overview of FDA-approved immune checkpoint inhibitors. *Int Immunopharmacol.* 2018;62:29–39.
- Naidoo J, Murphy C, Atkins MB, Brahmer JR, Champiat S, Feltquate D, et al. Society for Immunotherapy of Cancer (SITC) consensus definitions for immune checkpoint inhibitor-associated immune-related adverse events (irAEs) terminology. *J Immunother Cancer.* 2023;11(3):e006398.
- Marin-Acevedo JA, Harris DM, Burton MC. Immunotherapy-Induced Colitis: An Emerging Problem for the Hospitalist. *J Hosp Med.* 2018;13(6):390–5.
- Som A, Mandalia R, Alsaadi D, Farshidpour M, Charabaty A, Malhotra N, et al. Immune checkpoint inhibitor-induced colitis: A comprehensive review. *World J Clin Cases.* 2019;7(4):405–18.
- Wang DY, Ye F, Zhao S, Johnson DB. Incidence of immune checkpoint inhibitor-related colitis in solid tumor patients: A systematic review and meta-analysis. *Oncoimmunology.* 2017;6(10):e1344805.
- Nielsen DL, Juhl CB, Chen IM, Kellermann L, Nielsen OH. Immune checkpoint Inhibitor-Induced diarrhea and colitis: Incidence and management. A systematic review and meta-analysis. *Cancer Treat Rev.* 2022;109:102440.
- Liu YH, Zang XY, Wang JC, Huang SS, Xu J, Zhang P. Diagnosis and management of immune-related adverse events (irAEs) in cancer immunotherapy. *Biomed Pharmacother.* 2019;120:109437.
- Barnes MJ, Griseri T, Johnson AMF, Young W, Powrie F, Izcue A. CTLA-4 promotes Foxp3 induction and regulatory T cell accumulation in the intestinal lamina propria. *Mucosal Immunol.* 2013;6(2):324–34.
- Marthey L, Mateus C, Mussini C, Nachury M, Nancey S, Grange F, et al. Cancer immunotherapy with anti-CTLA-4 monoclonal antibodies induces an inflammatory bowel disease. *J Crohns Colitis.* 2016;10(4):395–401.
- Wing K, Onishi Y, Prieto-Martin P, Yamaguchi T, Miyara M, Fehervari Z, et al. CTLA-4 control over Foxp3+ regulatory T cell function. *Science.* 2008;322(5899):271–5.
- Luoma AM, Suo S, Williams HL, Sharova T, Sullivan K, Manos M, et al. Molecular pathways of colon inflammation induced by cancer immunotherapy. *Cell.* 2020;182(3):655–71.
- Tang L, Wang J, Lin N, Zhou Y, He W, Liu J, et al. Immune checkpoint inhibitor-associated colitis: From mechanism to management. *Front Immunol.* 2021;12:651377.
- Miller PL, Carson TL. Mechanisms and microbial influences on CTLA-4 and PD-1-based immunotherapy in the treatment of cancer: a narrative review. *Gut Pathog.* 2020;12(1):44.
- Ascierto PA, Del Vecchio M, Robert C, Mackiewicz A, Chiarion-Sileni V, Arance A, et al. Ipilimumab 10 mg/kg versus ipilimumab 3 mg/kg in patients with unresectable or metastatic melanoma: a randomised, double-blind, multicentre, phase 3 trial. *Lancet Oncol.* 2017;18(5):611–22.
- Farha N, Alkhayat M, Lindsey A, Mansoor E, Abou Saleh M. Immune checkpoint inhibitor-induced colitis: A nationwide population-based study. *Clin Res Hepatol Gastroenterol.* 2022;46(1):101778.
- Hasan Ali O, Berner F, Bomze D, Fässler M, Diem S, Cozzio A, et al. Human leukocyte antigen variation is associated with adverse events of checkpoint inhibitors. *Eur J Cancer.* 2019;107:8–14.
- Abu-Sbeih H, Faleck DM, Ricciuti B, Mendelsohn RB, Naqash AR, Cohen JV, et al. Immune checkpoint inhibitor therapy in patients with preexisting inflammatory bowel disease. *J Clin Oncol.* 2020;38(6):576–83.
- Vaziri H, Turshudzhyan A, Vecchio E. Immunotherapy-induced colitis. *J Clin Gastroenterol.* 2022;56(4):283–9.
- Portenkirchner C, Kienle P, Horisberger K. Checkpoint inhibitor-induced colitis—A clinical overview of incidence, prognostic implications and extension of current treatment options. *Pharmaceuticals.* 2021;14(4):367.
- Gong Z, Wang Y. Immune checkpoint inhibitor-mediated diarrhea and colitis: A clinical review. *JCO Oncol Pract.* 2020;16(8):453–61.
- Rajha E, Chaftari P, Kamal M, Maamari J, Chaftari C, Yeung S-CJ. Gastrointestinal adverse events associated with immune checkpoint inhibitor therapy. *Gastroenterol Rep (Oxf).* 2020;8(1):25–30.
- Li H, Fu ZY, Arslan ME, Cho D, Lee H. Differential diagnosis and management of immune checkpoint inhibitor-induced colitis: A comprehensive review. *World J Exp Med.* 2021;11(6):79–92.
- Hashash JG, Francis FF, Farraye FA. Diagnosis and management of immune checkpoint inhibitor colitis. *Gastroenterol Hepatol (N Y).* 2021;17(8):1–12.
- Dougan M, Wang Y, Rubio-Tapia A, Lim JK. AGA clinical practice update on diagnosis and management of immune checkpoint inhibitor (ICI) colitis and hepatitis: Expert review. *Gastroenterology.* 2021;160(4):1384–93.
- Eshet Y, Baruch EN, Shapira-Frommer R, Steinberg-Silman Y, Kuznetsov T, Ben-Betzalel G, et al. Clinical significance of pancreatic atrophy induced by immune-checkpoint inhibitors: A case-control study. *Cancer Immunol Res.* 2018;6(12):1453–8.
- Badran Y, Shih A, Leet D, Mooradian MJ, Coromilas A, Chen

- J, et al. Immune checkpoint inhibitor-associated celiac disease. *J Immunother Cancer*. 2020;8(1):e000958.
28. Abu-Sbeih H, Ali FS, Luo W, Qiao W, Raju GS, Wang Y. Importance of endoscopic and histological evaluation in the management of immune checkpoint inhibitor-induced colitis. *J Immunother Cancer*. 2018;6(1):95.
 29. Abu-Sbeih H, Ali FS, Wang Y. Immune-checkpoint inhibitors induced diarrhea and colitis. *Curr Opin Gastroenterol*. 2020;36(1):25–32.
 30. Geukes Foppen MH, Rozeman EA, van Wilpe S, Postma C, Snaebjornsson P, van Thienen JV, et al. Immune checkpoint inhibition-related colitis: symptoms, endoscopic features, histology and response to management. *ESMO Open*. 2018;3(1):e000278.
 31. Beck KE, Blansfield JA, Tran KQ, Feldman AL, Hughes MS, Royal RE, et al. Enterocolitis in patients with cancer after antibody blockade of cytotoxic T-lymphocyte-associated antigen 4. *J Clin Oncol*. 2006;24(15):2283–9.
 32. Nishida T, Iijima H, Adachi S. Immune checkpoint inhibitor-induced diarrhea/colitis: Endoscopic and pathologic findings. *World J Gastrointest Pathophysiol*. 2019;10(2):17–28.
 33. Wang Y, Abu-Sbeih H, Mao E, Ali N, Qiao W, Trinh VA, et al. Endoscopic and histologic features of immune checkpoint inhibitor-related colitis. *Inflamm Bowel Dis*. 2018;24(8):1695–705.
 34. Oble DA, Mino-Kenudson M, Goldsmith J, Hodi FS, Seliem RM, Dranoff G, et al. α -CTLA-4 mAb-associated panenteritis. *Am J Surg Pathol*. 2008;32(8):1130–7.
 35. Chen J, Pezhohu MK, Lauwers GY, Masia R. Histopathologic features of colitis due to immunotherapy with anti-PD-1 antibodies. *Am J Surg Pathol*. 2017;41(5):643–54.
 36. Gonzalez RS, Salaria SN, Bohannon CD, Huber AR, Feely MM, Shi C. PD-1 inhibitor gastroenterocolitis: Case series and appraisal of “immunomodulatory gastroenterocolitis.” *Histopathology*. 2017;70(4):558–67.
 37. Baroudjian B, Lourenco N, Pagès C, Chami I, Maillet M, Bertheau P, et al. Anti-PD1-induced collagenous colitis in a melanoma patient. *Melanoma Res*. 2016;26(3):308–11.
 38. Verschuren EC, van den Eertwegh AJ, Wonders J, Slangen RM, van Delft F, van Bodegraven AA, et al. Clinical, endoscopic, and histologic characteristics of ipilimumab-associated colitis. *Clin Gastroenterol Hepatol*. 2016;14(6):836–42.
 39. Grover S, Rahma OE, Hashemi N, Lim RM. Gastrointestinal and hepatic toxicities of checkpoint inhibitors: Algorithms for management. *Am Soc Clin Oncol Educ Book*. 2018;38:13–9.
 40. Haanen JB, Carbone F, Robert C, Kerr KM, Peters S, Larkin J, et al. Management of toxicities from immunotherapy: ESMO clinical practice guidelines for diagnosis, treatment and follow-up. *Ann Oncol*. 2017;28(suppl_4):iv119–42.
 41. Brahmer JR, Lacchetti C, Schneider BJ, Atkins MB, Brassil KJ, Caterino JM, et al. Management of immune-related adverse events in patients treated with immune checkpoint inhibitor therapy: American Society of Clinical Oncology clinical practice guideline. *J Clin Oncol*. 2018;36(17):1714–68.
 42. Puzanov I, Diab A, Abdallah K, Bingham CO, Brogdon C, Dadu R, et al. Managing toxicities associated with immune checkpoint inhibitors: Consensus recommendations from the Society for Immunotherapy of Cancer (SITC) Toxicity Management Working Group. *J Immunother Cancer*. 2017;5(1):95.
 43. Hughes MS, Molina GE, Chen ST, Zheng H, Deshpande V, Fadden R, et al. Budesonide treatment for microscopic colitis from immune checkpoint inhibitors. *J Immunother Cancer*. 2019;7(1):292.
 44. Berman D, Parker SM, Siegel J, Chasalow SD, Weber J, Galbraith S, et al. Blockade of cytotoxic T-lymphocyte antigen-4 by ipilimumab results in dysregulation of gastrointestinal immunity in patients with advanced melanoma. *Cancer Immun*. 2010;10:11.
 45. Weber J, Thompson JA, Hamid O, Minor D, Amin A, Ron I, et al. A randomized, double-blind, placebo-controlled, phase II study comparing the tolerability and efficacy of ipilimumab administered with or without prophylactic budesonide in patients with unresectable stage III or IV melanoma. *Clin Cancer Res*. 2009;15(17):5591–8.
 46. Thompson JA, Schneider BJ, Brahmer J, Andrews S, Armand P, Bhatia S, et al. Management of immunotherapy-related toxicities, version 1.2019. *J Natl Compr Canc Netw*. 2019;17(3):255–89.
 47. Gupta A, De Felice KM, Loftus EV, Khanna S. Systematic review: Colitis associated with anti-CTLA-4 therapy. *Aliment Pharmacol Ther*. 2015;42(4):406–17.
 48. Desmedt V, Jauregui-Amezaga A, Fierens L, Aspeslagh S, Dekervel J, Wauters E, et al. Position statement on the management of the immune checkpoint inhibitor-induced colitis via multidisciplinary modified Delphi consensus. *Eur J Cancer*. 2023;187:36–57.
 49. Powell N, Ibraheim H, Raine T, Speight RA, Papa S, Brain O, et al. British Society of Gastroenterology endorsed guidance for the management of immune checkpoint inhibitor-induced enterocolitis. *Lancet Gastroenterol Hepatol*. 2020;5(7):679–97.
 50. Abu-Sbeih H, Ali FS, Wang X, Mallepally N, Chen E, Altan M, et al. Early introduction of selective immunosuppressive therapy associated with favorable clinical outcomes in patients with immune checkpoint inhibitor-induced colitis. *J Immunother Cancer*. 2019;7(1):93.
 51. Dougan M. Checkpoint blockade toxicity and immune homeostasis in the gastrointestinal tract. *Front Immunol*. 2017;8:1547.
 52. Johnston RL, Lutzky J, Chodhry A, Barkin JS. Cytotoxic T-lymphocyte-associated antigen 4 antibody-induced colitis and its management with infliximab. *Dig Dis Sci*. 2009;54(11):2538–40.
 53. Dougan M. Gastrointestinal and hepatic complications of immunotherapy: Current management and future perspectives. *Curr Gastroenterol Rep*. 2020;22(4):11.
 54. Ibraheim H, Baillie S, Samaan MA, Abu-Sbeih H, Wang Y, Talley NJ, et al. Systematic review with meta-analysis: effectiveness of anti-inflammatory therapy in immune checkpoint inhibitor-induced enterocolitis. *Aliment Pharmacol Ther*.

- 2020;52(9):1432–52.
55. Desmedt V, Jauregui-Amezaga A, Fierens L, Aspeslagh S, Dekervel J, Wauters E, et al. Position statement on the management of the immune checkpoint inhibitor-induced colitis via multidisciplinary modified Delphi consensus. *Eur J Cancer*. 2023;187:36–57.
 56. Schneider BJ, Naidoo J, Santomaso BD, Lacchetti C, Adkins S, Anadkat M, et al. Management of immune-related adverse events in patients treated with immune checkpoint inhibitor therapy: ASCO guideline update. *J Clin Oncol*. 2021;39(36):4073–126.
 57. Harvey C, Nahar KJ, McKeown J, Lo SN, Farag S, Yousaf N, et al. Management of infliximab refractory immune checkpoint inhibitor gastrointestinal toxicity: A multicenter case series. *J Immunother Cancer*. 2024;12(1):e008232.
 58. Soularue E, Lepage P, Colombel JF, Coutzac C, Faleck D, Marthey L, et al. Enterocolitis due to immune checkpoint inhibitors: A systematic review. *Gut*. 2018;67(11):2056–67.
 59. Routy B, Le Chatelier E, Derosa L, Duong CPM, Alou MT, Daillère R, et al. Gut microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors. *Science*. 2018;359(6371):91–7.
 60. Gopalakrishnan V, Spencer CN, Nezi L, Reuben A, Andrews MC, Karpnits TV, et al. Gut microbiome modulates response to anti-PD-1 immunotherapy in melanoma patients. *Science*. 2018;359(6371):97–103.
 61. Matson V, Fessler J, Bao R, Chongsuwat T, Zha Y, Alegre ML, et al. The commensal microbiome is associated with anti-PD-1 efficacy in metastatic melanoma patients. *Science*. 2018;359(6371):104–8.
 62. Wang Y, Wiesenloski DH, Helmink BA, Gopalakrishnan V, Choi K, DuPont HL, et al. Fecal microbiota transplantation for refractory immune checkpoint inhibitor-associated colitis. *Nat Med*. 2018;24(12):1804–8.
 63. Fasanella MK, Robillard KT, Boland PM, Bain AJ, Kanehira K. Use of fecal microbial transplantation for immune checkpoint inhibitor colitis. *ACG Case Rep J*. 2020;7(4):e00360.
 64. Groenewegen B, Terveer EM, Joosse A, Barnhoorn MC, Zwitterink RD. Fecal microbiota transplantation for immune checkpoint inhibitor-induced colitis is safe and contributes to recovery: Two case reports. *J Immunother*. 2023;46(6):216–20.
 65. Elkrif A, Waters NR, Smith N, Dai A, Slingerland J, Aleynick N, et al. Immune-related colitis is associated with fecal microbial dysbiosis and can be mitigated by fecal microbiota transplantation. *Cancer Immunol Res*. 2023;12(3):308–21.

Corresponding author:

Angelos Koutras
Associate Professor, Division of Oncology, Department
of Internal Medicine, Medical School, University of Patras,
Patras, Greece, PC 26504
Tel.: +30 2610999535, E-mail: angkoutr@otenet.gr

Immunotherapy-induced thyroid dysfunction

George C. Kyriakopoulos

Abstract

Immune checkpoint inhibitors (ICIs) are a groundbreaking class of drugs that significantly advance cancer treatment by leveraging the immune system against cancer cells. However, their efficacy as anti-cancer agents is accompanied by a broad range of immune-related adverse effects (irAEs), including endocrinopathies. Thyroid dysfunction is among the most common endocrinopathies induced by ICI therapy, making it a major concern. The pathophysiology typically involves destructive thyroiditis, leading to the common patterns of thyrotoxicosis followed by hypothyroidism or isolated hypothyroidism. Diagnosis relies mostly on clinical presentation and laboratory tests. Treatment varies from levothyroxine substitution to the use of beta blockers, based on the severity of thyroid dysfunction. As immunotherapy evolves, recognizing and managing ICI-induced thyroid dysfunction is essential for improving patient safety and outcomes. This review aims to explore the significance of ICI-induced thyroid dysfunction, detailing the patterns, mechanisms, diagnostic approaches, and treatment strategies.

Key words: *Immunotherapy; adverse effects; thyroid dysfunction; hypothyroidism; hyperthyroidism*

INTRODUCTION

In recent years, immune checkpoint inhibitor (ICI) therapy has become a promising advancement in cancer treatment. This innovative approach utilizes the body's immune system to target cancer cells, offering patients new therapeutic options and improved survival rates. Initially used for treating malignant melanoma and lung cancer, this therapy involves administering monoclonal antibodies that target specific cell proteins, such as cytotoxic T lymphocyte-associated protein 4 (CTLA-4), programmed cell death protein 1 (PD-1), and its ligand (PD-L1), thereby inducing T cell activation against neoplasms [1].

However, despite its benefits, ICI therapy is associated with various adverse effects, with thyroid disorders being among the most common endocrine

complications. Thyroid dysfunction can appear as either primary hypothyroidism or thyrotoxicosis. The most common type of thyroid abnormality is destructive thyroiditis, which initially causes a phase of thyrotoxicosis followed by permanent hypothyroidism. Rarely, Graves' disease could also occur. The pathophysiology is primarily linked to destructive thyroiditis caused by a T cell-mediated acute autoimmune response [2]. Additionally, research consistently points to the role of autoantibodies against thyroglobulin (Tg), thyroid peroxidase (TPO), and thyroid-stimulating hormone (TSH) receptor, as well as the role of cytokines in the disease's pathogenesis [3]. Consequently, laboratory tests measuring TSH, free thyroxine (fT4), and antibodies are crucial for accurate diagnosis and for monitoring before and during ICI therapy. ICI-induced thyroidopathy can range from asymptomatic cases to severe conditions, including fatalities. Addressing these complications requires prompt diagnosis and therapeutic strategies customized to the clinical manifestations and their severity. Close monitoring and collaboration between

oncologists and endocrinologists are essential to manage these effects effectively [4,5].

Possible mechanisms of immunotherapy-induced adverse effects

The immune activation responsible for most immune-related adverse events (irAEs) may be linked to the mechanisms driving antitumor immune responses. This tumor-specific hypothesis is supported by the consistent positive correlation between therapeutic responses and the incidence of irAEs [6–8]. Correlative studies offer further evidence of this association, showing shared T cell receptor sequences and upregulated organ-specific transcripts between tumors and non-malignant tissues affected by toxicities [9,10]. Additionally, the development of vitiligo, an autoimmune response targeting melanocytes, serves as a reliable indicator of ICI antitumor activity, particularly in melanoma patients. This suggests a mechanistic link between irAEs and antitumor immunity [11]. If both the beneficial and adverse outcomes stem from the same processes, long-term responders might face a higher risk of chronic toxicities compared to those who do not benefit from the therapy.

Evidence also suggests that certain irAEs may emerge from mechanisms unrelated to antitumor activity, including factors such as the microbiome and viral or tissue-specific elements [12–15]. The diversity of irAEs likely reflects distinct and varied mechanisms for each type of event. Different cell types are implicated as dominant drivers in various preclinical models and biopsy samples from affected tissues. For instance, tissue-resident memory CD8+ T cells were predominant in colon biopsy samples from patients with ICI-induced colitis, while cytotoxic activated memory CD4+ T cells were most prevalent in the brain of a patient with fatal encephalitis [13,16–18]. Targeted inhibition of specific cytokines, such as IL-6, could potentially separate anti-tumor from antihost immune responses in preclinical models [19,20]. Ultimately, a single mechanistic explanation for irAEs is unlikely; they probably result from both tumor-related and tumor-unrelated factors. Additionally, the mechanisms distinguishing acute versus chronic irAEs remain poorly understood.

The mechanism underlying immune checkpoint inhibitor (ICI)-induced thyroid disorders also remains unclear. In a study conducted by Osario et al., among ten patients who developed thyroid dysfunction following pembrolizumab administration, eight patients were found to have antithyroglobulin or antimicroso-

mal antibodies [21]. Another study reported ten cases of thyroiditis following anti-PD-1 treatment. Among these cases, six patients experienced transient thyrotoxicosis followed by hypothyroidism, while four patients developed hypothyroidism without a preceding thyrotoxic phase. All patients tested positive for antithyroid antibodies [22]. Iyer et al. also observed that thyroid peroxidase and thyroglobulin antibodies were present in 44.7% and 33% of patients, respectively, after ICI therapy [23]. However, other immune pathways, involving T-cells, natural killer cells, and monocytes might also contribute to thyroid dysfunction during anti-PD-1 therapy, independent of antibody presence [24]. Interestingly, positron emission tomography (PET) scans of patients affected by thyroid irAEs frequently show a diffuse increase in 18-fludeoxyglucose (18-FDG) uptake, suggesting it may serve as a better biomarker for thyroid irAEs. Thyroid antibodies, on the other hand, might indicate the severity and increased risk of needing hormone replacement [25]. Finally, certain cytokines are associated with thyroid irAEs. Elevated levels of cytokines, such as IL-1 β , IL-2, and GM-CSF, and lower levels of IL-8, G-CSF, and MCP-1, indicating a shift in the Th1/Th2 balance, may be linked to the development of these immune-related thyroid disorders [26].

Risk factors for immunotherapy-induced thyroid dysfunction

Patients undergoing ICI therapy are exposed to several risk factors that may predispose them to thyroid dysfunction and other endocrine-related adverse effects (Table 1). One significant risk factor is gender, as women are more likely to develop thyroid dysfunction during ICI therapy compared to men. Age is another critical factor, with younger patients being at a higher risk for

Table 1. Risk factors for immunotherapy-induced thyroid dysfunction.

Female
Younger age
Elevated anti-TPO or anti-TG antibodies before starting ICI
Higher baseline TSH levels
High number of immunotherapy cycles
Obesity
Race: Caucasians and Latinos (hypothyroidism)
African Americans (thyrotoxicosis)

thyroid-related issues [27,28]. The presence of elevated anti-TPO or anti-TG antibodies before starting ICI therapy is another important risk factor. Patients with higher levels of these antibodies have an increased likelihood of developing thyroid dysfunction. Additionally, higher baseline TSH levels at the commencement of therapy are associated with a greater risk of experiencing thyroid problems during treatment [29–31]. The number of immunotherapy cycles a patient undergoes also influences the risk, with a higher number of cycles correlating with an increased incidence of thyroid and other endocrine issues. Obesity further compounds this risk, as it has been identified as a significant factor in the development of thyroid dysfunction during ICI therapy [30,32]. Finally, racial background impacts the type of thyroid dysfunction experienced. Caucasians and Latinos are more prone to developing hypothyroidism as a result of ICI therapy, whereas African Americans are more likely to experience thyrotoxicosis [33]. Recognizing these risk factors can aid healthcare providers in monitoring and managing potential endocrine side effects more effectively in patients undergoing ICI therapy.

Thyroid disorders induced by immunotherapy

Thyroid dysfunction is one of the most common endocrinopathies following therapy with immune checkpoint inhibitors (ICIs). The clinical spectrum ranges from overt hypothyroidism to overt thyrotoxicosis, with destructive thyroiditis being the common underlying pathophysiological mechanism. Thyroid dysfunction is more frequently observed in patients treated with anti-PD-1 agents or a combination of ipilimumab and nivolumab, and less commonly in those treated with anti-CTLA-4 or anti-PD-L1 monotherapies [34–37].

More specifically, primary hypothyroidism affects 6%–9% of patients undergoing anti-PD-1 and/or anti-PD-L1 therapy, 4% of those on anti-CTLA-4 therapy, and approximately 16% of patients receiving a combination of anti-PD(L)1 and anti-CTLA-4 therapies (Figure 1) [38]. It can be preceded by a hyperthyroid state, which may be subclinical. Most cases arise within the first three months of treatment initiation, but onset can occur at any time during the therapy [39].

On the other hand, Immune-related (IR) hyperthyroidism occurs less frequently. It is reported in approximately 2%–5% of patients treated with immune checkpoint inhibitor (ICI) monotherapy and in about 10% of those receiving a combination of anti-PD(L)1 and anti-CTLA-4 therapies (Figure 1) [38]. Transient

thyroiditis remains the most common cause of IR hyperthyroidism. Among these cases, around 40% are present with symptomatic thyrotoxicosis, while 60% manifest as subclinical thyrotoxicosis, which is usually followed by hypothyroidism [40]. Primary hyperthyroidism due to Graves-like disease is rarely reported. When it occurs, persistent hyperthyroidism, diffuse goiter, and ophthalmopathy may indicate this diagnosis (41). Thyroid dysfunction typically emerges within a few weeks of ICI initiation and can occur after a single dose (42). Median onset time ranges from 18 to 123 days, but cases have been reported as early as seven days or as late as three years after initiation [34–37]. The dysfunction often starts as painless thyroiditis, beginning with a transient thyrotoxic phase that is usually mild or asymptomatic, lasting a few weeks before transitioning to euthyroidism or hypothyroidism. Symptoms are generally nonspecific, such as fatigue and weight loss, but can include tremor, anxiety and heat intolerance [43,44]. Physical examination may reveal tachycardia or warm, smooth skin, although severe thyrotoxicosis (thyroid storm) is rare [45,46].

The median time for progression to euthyroidism or hypothyroidism is four–seven weeks [24,42]. In some cases, hypothyroidism, whether subclinical or clinical, is the initial presentation and may be transient or permanent. Symptoms are usually mild and nonspecific, including fatigue and weight gain, but may also involve bradycardia, cold skin, constipation, and cold intolerance [43,44]. Myxedema coma, a complication resulting from untreated hypothyroidism, is very rare [47].

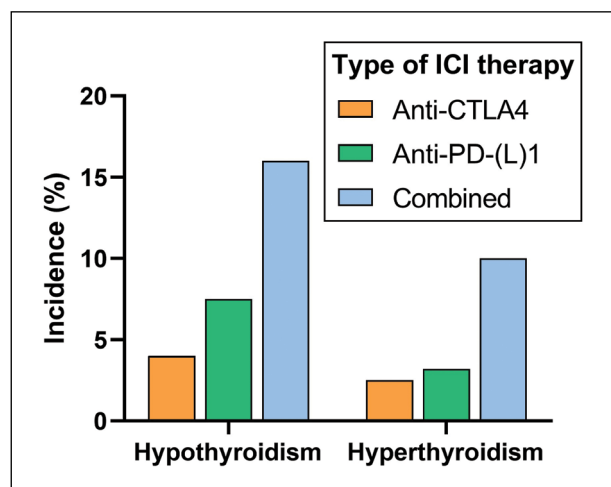


Figure 1. Incidence of thyroid adverse events in patients treated with ICIs.

Elevated autoantibodies against thyroid peroxidase (anti-TPO) and thyroglobulin (anti-Tg) are found in some patients developing thyroid dysfunction after ICI therapy [34–37]. High titers of these antibodies prior to therapy appear to be related to ICI-induced thyroid dysfunction, though they are not necessary but represent a risk factor [26]. Stimulating autoantibodies for the TSH receptor (TRAb or TSI) were negative in most patients, though in rare cases, TRAb positivity suggests co-existing Graves’ disease [34–37,48,49].

Measurement of TSH, FT4, and occasionally T3 levels is typically sufficient for diagnosis. In thyrotoxicosis due to thyroiditis, FT4 levels are more elevated compared to T3 levels due to the release of stored thyroxine (T4) into the bloodstream. In hyperthyroidism, T3 levels are higher due to stimulation of the thyroid gland. Differentiating between primary and central hypothyroidism is crucial, as hypophysitis should be considered in these patients [43,44,50,51]. More detailed information can be found in Table 2.

Monitoring and management of immune-related hypothyroidism

The management of endocrine adverse events related to immunotherapy, specifically thyroid dysfunction, follows a structured approach based on the assessment of thyroid function tests (TFTs). For patients with asymptomatic or subclinical hypothyroidism, it is recommended to monitor TSH and free T4 every four-six weeks. If TSH is elevated, the next steps depend on the

levels of TSH and free T4. If TSH levels range between 4 and less than 10 mIU/ml, the patient is asymptomatic with normal free T4 levels, the patient should continue immunotherapy and regular monitoring of TFTs. If TSH exceeds 10 mIU/ml but free T4 is normal, immunotherapy can continue, and the consideration of levothyroxine treatment is advised. In cases where TSH is normal or low and free T4 is low, it is important to investigate for central hypothyroidism or hypophysitis and exclude recovery from thyrotoxicosis [50,51].

For patients with clinical (overt) primary hypothyroidism, TSH should be monitored every four-six weeks. Management includes continuing immunotherapy and considering endocrine consultation. Thyroid hormone supplementation with levothyroxine should be initiated, and TSH levels should be reassessed in four-six weeks to guide dosing adjustments. Additionally, it is essential to exclude concomitant adrenal insufficiency by checking the morning cortisol level. This approach ensures that thyroid dysfunctions are identified and managed effectively without discontinuing necessary immunotherapy, thus maintaining the therapeutic benefits while mitigating adverse endocrine effects (Figure 2) [50,51].

Monitoring and management of immune-related hyperthyroidism

For patients experiencing thyrotoxicosis induced by immunotherapy, the management process is detailed and depends on specific thyroid function test

Table 2. Evaluation and monitoring of patients on ICIs for possible immune-related thyroid dysfunction.

Before ICI administration (Baseline)	Clinical evaluation for symptoms: Extreme weakness, unusual headache patterns, increased sweating, rapid heartbeat, weight loss or weight gain, mood changes, constipation or diarrhea, nausea or vomiting, abdominal pain Laboratory evaluation: Morning TSH, FT4, cortisol
Every 4-6 weeks (incl. 4-6 weeks after the last cycle)	Clinical evaluation for symptoms Laboratory evaluation: Morning TSH, FT4, cortisol
Additional tests	
<i>If hypothyroidism suspected</i>	anti-TPO, anti-TG
<i>If hyperthyroidism suspected</i>	T3 TSH, FT4, T3 every 2-3 weeks to diagnose persistent hyperthyroidism or hypothyroidism (due to destructive thyroiditis)

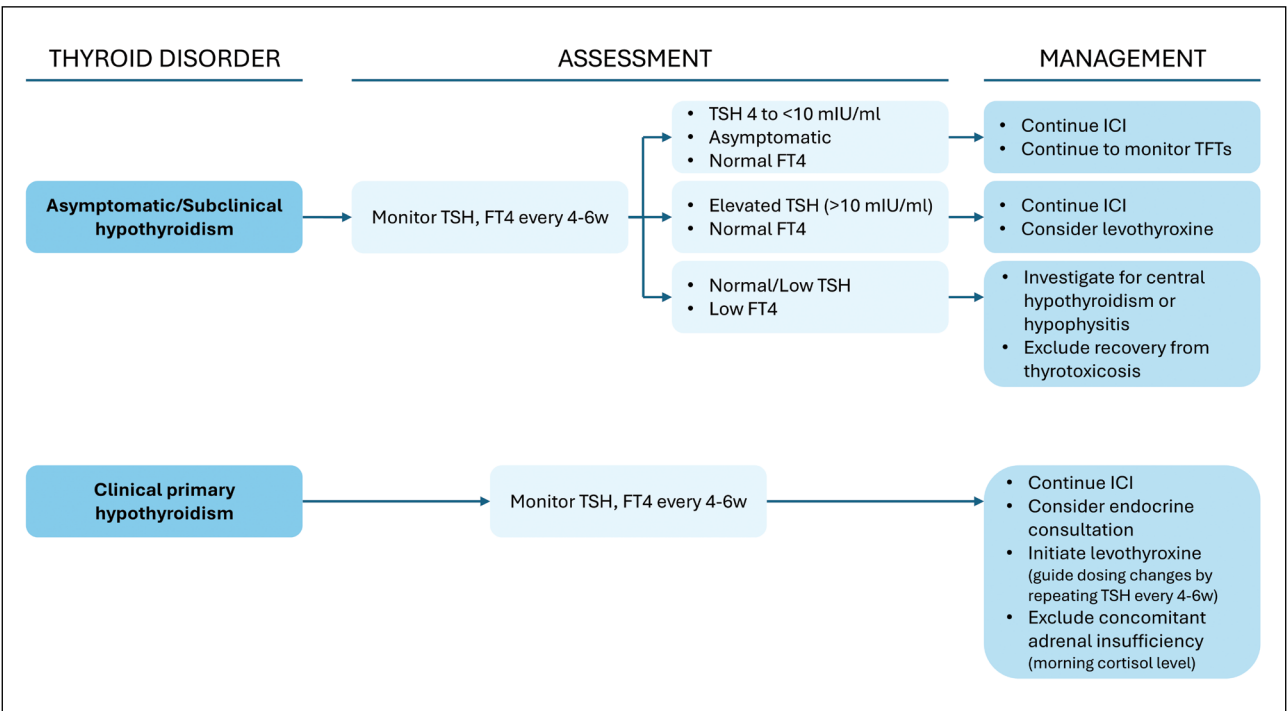


Figure 2. Monitoring and management of immune-related hypothyroidism.

(TFT) results. Immunotherapy-induced thyrotoxicosis is typically caused by transient or evolving painless thyroiditis. Antithyroid medications such as methimazole or propylthiouracil are not recommended for this condition. When thyrotoxicosis is identified, indicated by low or suppressed TSH levels and high free T4/total T3, an endocrine consultation should be considered if the patient is symptomatic. If the patient is asymptomatic, immunotherapy can continue. Symptomatic patients may be treated with propranolol (10–20 mg every 4–6 hours as needed) or atenolol or metoprolol until thyrotoxicosis resolves. TFTs should be repeated every 4–6

weeks. If thyrotoxicosis resolves, no further therapy for thyrotoxicosis is necessary. However, it often evolves into hypothyroidism in 50%–90% of cases, requiring treatment with thyroid hormone replacement, as mentioned above. Rarely, painful thyroiditis can occur, and in such cases, prednisolone 0.5 mg/kg should be prescribed [50]. If thyrotoxicosis persists, an evaluation for Graves' disease should be considered (Figure 3) [50,51].

Role of glucocorticoids in immunotherapy-induced thyroid dysfunction

Even though thyroid disorders have emerged as

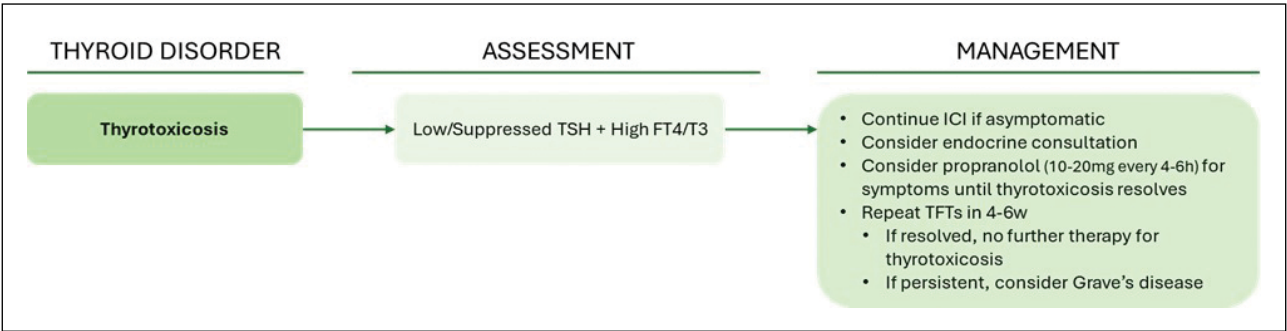


Figure 3. Monitoring and management of immune-related hyperthyroidism.

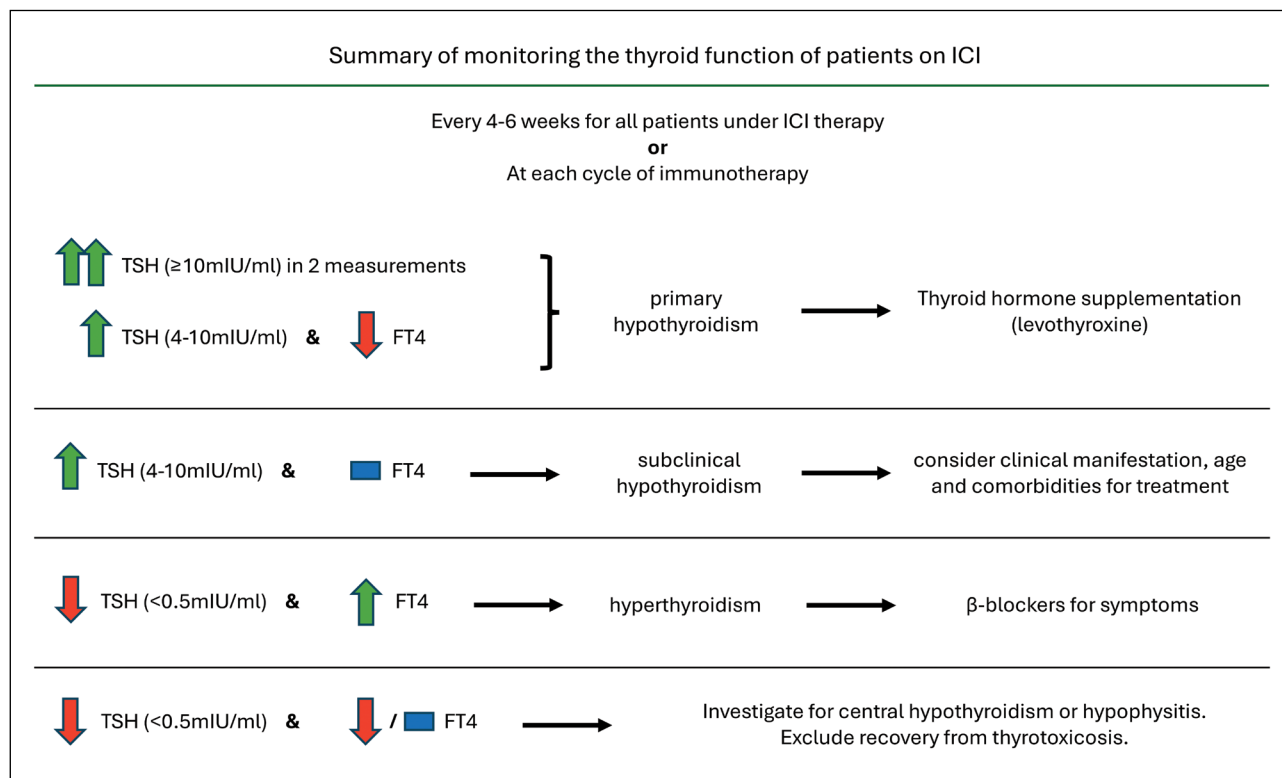


Figure 4. Summary of monitoring the thyroid function of patients on ICI therapy.

one of the most common immune-related adverse events (irAE) associated with immunotherapy, optimum management strategies and predictive biomarkers for vulnerable individuals remain to be fully explored. High-dose glucocorticoid (HDG) therapy is routinely recommended for irAEs [52,53]. However, a systematic analysis of the impact of glucocorticoid therapy on the outcome of immune-checkpoint inhibitor (ICI)-induced thyroid disorders is lacking. In a study analyzing 151 patients with or without ICI-related thyroid disorders, the patients with ICI-related thyroid disorders were divided into two subgroups: those receiving HDG treatment and those not. The results showed no significant differences between the HDG and no HDG groups in terms of the median duration of thyrotoxicosis (28 vs 42 days), the median time to conversion from thyrotoxicosis to hypothyroidism (39 vs 42 days), the median time to onset of hypothyroidism (both 63 days), and the median maintenance dose of levothyroxine (1.5 vs 1.3 mg/kg/day). The median pretreatment TSH was higher in patients with ICI-related thyroid disorders compared to those without (2.3 vs 1.7 mIU/L). Baseline TSH was significantly higher in patients who developed ICI-related thyroid disorders.

Subgroup analysis revealed significantly higher baseline TSH in male patients with ICI-induced thyroid dysfunction, but not in female patients. This study concluded that HDG treatment did not improve the outcome of ICI-related thyroid disorders [54].

CONCLUSIONS

Immune checkpoints targeted by ICIs are crucial for maintaining immunological self-tolerance, making these therapies capable of triggering autoimmune adverse effects. Among the most common are thyroid disorders, as well as other endocrine complications like hypophysitis, diabetes mellitus, and primary adrenal insufficiency. Physicians managing such patients need to be well-informed about ICI-related endocrine adverse events, including their clinical presentation, laboratory findings, frequency, and severity. Effective screening and management require close collaboration between oncologists and endocrinologists. As survival rates improve, the monitoring and management of long-term consequences have become increasingly crucial, highlighting the necessity for joint clinics to provide comprehensive care.

To summarize, the management of thyroid-related adverse events from immunotherapy involves structured monitoring and intervention based on thyroid function tests (TFTs). For asymptomatic or subclinical hypothyroidism, TSH and free T4 levels should be monitored every four-six weeks. If TSH is elevated (ranging from 4 and less than 10 mIU/ml with normal free T4), immunotherapy may proceed with regular TFT monitoring. If TSH exceeds 10 mIU/ml, consideration of levothyroxine alongside ongoing immunotherapy is advised. In overt primary hypothyroidism, TSH is monitored similarly, with initiation of levothyroxine and adrenal insufficiency exclusion. Immune-related thyrotoxicosis management involves symptom-driven use of beta-blockers, ongoing immunotherapy if asymptomatic, and periodic TFT checks, anticipating potential transition to hypothyroidism. Persistent thyrotoxicosis may prompt evaluation for Graves' disease. This approach ensures thyroid dysfunctions are effectively managed without interrupting essential immunotherapy, balancing therapeutic benefits with managing adverse endocrine effects [50,51]. A summary can be found in Figure 4.

Acknowledgements: *The author would like to thank Prof. A. Koutras for the invitation and for all the support. G.C.K. is a recipient of a Fulbright Doctoral Dissertation Visiting Research Student Fellowship (2023–2024).*

Conflict of interest disclosure: *None to declare.*

Declaration of funding sources: *None to declare.*

Author contributions: *GK was responsible for the conception, research, writing and the final draft of this review.*

REFERENCES

1. Johnson DB, Nebhan CA, Moslehi JJ, Balko JM. Immune-checkpoint inhibitors: long-term implications of toxicity. *Nat Rev Clin Oncol.* 2022;19(4):254–67.
2. Martins F, Sofiya L, Sykietis GP, Lamine F, Maillard M, Fraga M, et al. Adverse effects of immune-checkpoint inhibitors: epidemiology, management and surveillance. *Nat Rev Clin Oncol.* 2019;16(9):563–80.
3. Basek A, Jakubiak GK, Cieślak G, Stanek A. Life-Threatening Endocrinological Immune-Related Adverse Events of Immune Checkpoint Inhibitor Therapy. *Cancers (Basel).* 2023;15(24):5786.
4. Deligiorgi M V., Sagredou S, Vakkas L, Trafalis DT. The Continuum of Thyroid Disorders Related to Immune Checkpoint Inhibitors: Still Many Pending Queries. *Cancers (Basel).* 2021;13(21):5277.
5. Paschou SA, Stefanaki K, Psaltopoulou T, Liontos M, Koutsoukos K, Zagouri F, et al. How we treat endocrine complications of immune checkpoint inhibitors. *ESMO Open.* 2021;6(1):100011.
6. Das S, Johnson DB. Immune-related adverse events and anti-tumor efficacy of immune checkpoint inhibitors. *J Immunother Cancer.* 2019;7(1):306.
7. Quach HT, Dewan AK, Davis EJ, Ancell KK, Fan R, Ye F, et al. Association of Anti-Programmed Cell Death 1 Cutaneous Toxic Effects With Outcomes in Patients With Advanced Melanoma. *JAMA Oncol.* 2019;5(6):906.
8. Eggermont AMM, Kicinski M, Blank CU, Mandala M, Long G V., Atkinson V, et al. Association Between Immune-Related Adverse Events and Recurrence-Free Survival Among Patients With Stage III Melanoma Randomized to Receive Pembrolizumab or Placebo. *JAMA Oncol.* 2020;6(4):519.
9. Berner F, Bomze D, Diem S, Ali OH, Fässler M, Ring S, et al. Association of Checkpoint Inhibitor-Induced Toxic Effects With Shared Cancer and Tissue Antigens in Non-Small Cell Lung Cancer. *JAMA Oncol.* 2019;5(7):1043.
10. Johnson DB, Balko JM, Compton ML, Chalkias S, Gorham J, Xu Y, et al. Fulminant Myocarditis with Combination Immune Checkpoint Blockade. *New England Journal of Medicine.* 2016;375(18):1749–55.
11. Guida M, Strippoli S, Maule M, Quagliano P, Ramondetta A, Chiaron Sileni V, et al. Immune checkpoint inhibitor associated vitiligo and its impact on survival in patients with metastatic melanoma: an Italian Melanoma Intergroup study. *ESMO Open.* 2021;6(2):100064.
12. Dubin K, Callahan MK, Ren B, Khanin R, Viale A, Ling L, et al. Intestinal microbiome analyses identify melanoma patients at risk for checkpoint-blockade-induced colitis. *Nat Commun.* 2016;7(1):10391.
13. Luoma AM, Suo S, Williams HL, Sharova T, Sullivan K, Manos M, et al. Molecular Pathways of Colon Inflammation Induced by Cancer Immunotherapy. *Cell.* 2020;182(3):655–671.e22.
14. Andrews MC, Duong CPM, Gopalakrishnan V, Iebba V, Chen WS, Derosa L, et al. Gut microbiota signatures are associated with toxicity to combined CTLA-4 and PD-1 blockade. *Nat Med.* 2021;27(8):1432–41.
15. Lam KC, Goldszmid RS. Can gut microbes predict efficacy and toxicity of combined immune checkpoint blockade? *Cancer Cell.* 2021;39(10):1314–6.
16. Wei SC, Meijers WC, Axelrod ML, Anang NAAS, Screever EM, Wescott EC, et al. A Genetic Mouse Model Recapitulates Immune Checkpoint Inhibitor-Associated Myocarditis and Supports a Mechanism-Based Therapeutic Intervention. *Cancer Discov.* 2021;11(3):614–25.
17. Das R, Bar N, Ferreira M, Newman AM, Zhang L, Bailur JK, et al. Early B cell changes predict autoimmunity following combination immune checkpoint blockade. *Journal of Clinical Investigation.* 2018;128(2):715–20.
18. Yasuda Y, Iwama S, Sugiyama D, Okuji T, Kobayashi T, Ito M, et al. CD4+ T cells are essential for the development of destructive thyroiditis induced by anti-PD-1 antibody in thyroglobulin-immunized mice. *Sci Transl Med.* 2021;13(593).
19. Johnson DH, Hailemichael Y, Foo WC, Hess KR, Haymaker

- CL, Wani KM, et al. Interleukin-6 is potential target to decouple checkpoint inhibitor-induced colitis from antitumor immunity. *Journal of Clinical Oncology*. 2019;37(15_suppl):2616–2616.
20. Stroud CR, Hegde A, Cherry C, Naqash AR, Sharma N, Addepalli S, et al. Tocilizumab for the management of immune mediated adverse events secondary to PD-1 blockade. *Journal of Oncology Pharmacy Practice*. 2019;25(3):551–7.
 21. Osorio JC, Ni A, Chaft JE, Pollina R, Kasler MK, Stephens D, et al. Antibody-mediated thyroid dysfunction during T-cell checkpoint blockade in patients with non-small-cell lung cancer. *Annals of Oncology*. 2017;28(3):583–9.
 22. Orlov S, Salari F, Kashat L, Walfish PG. Induction of Painless Thyroiditis in Patients Receiving Programmed Death 1 Receptor Immunotherapy for Metastatic Malignancies. *J Clin Endocrinol Metab*. 2015;100(5):1738–41.
 23. Iyer PC, Cabanillas ME, Waguespack SG, Hu MI, Thosani S, Lavis VR, et al. Immune-Related Thyroiditis with Immune Checkpoint Inhibitors. *Thyroid*. 2018;28(10):1243–51.
 24. Delivanis DA, Gustafson MP, Bornschlegl S, Merten MM, Kottschade L, Withers S, et al. Pembrolizumab-Induced Thyroiditis: Comprehensive Clinical Review and Insights Into Underlying Involved Mechanisms. *J Clin Endocrinol Metab*. 2017;102(8):2770–80.
 25. Kotwal A, Kottschade L, Ryder M. PD-L1 Inhibitor-Induced Thyroiditis Is Associated with Better Overall Survival in Cancer Patients. *Thyroid*. 2020;30(2):177–84.
 26. Kurimoto C, Inaba H, Ariyasu H, Iwakura H, Ueda Y, Uraki S, et al. Predictive and sensitive biomarkers for thyroid dysfunctions during treatment with immune-checkpoint inhibitors. *Cancer Sci*. 2020;111(5):1468–77.
 27. Stelmachowska-Banaś M, Czajka-Oraniec I. Management of endocrine immune-related adverse events of immune checkpoint inhibitors: an updated review. *Endocr Connect*. 2020;9(10):R207–28.
 28. Campredon P, Mouly C, Lusque A, Bigay-Game L, Bousquet E, Mazières J, et al. Incidence of thyroid dysfunctions during treatment with nivolumab for non-small cell lung cancer: Retrospective study of 105 patients. *Presse Med*. 2019;48(4):e199–207.
 29. Kobayashi T, Iwama S, Yasuda Y, Okada N, Tsunekawa T, Onoue T, et al. Patients With Antithyroid Antibodies Are Prone To Develop Destructive Thyroiditis by Nivolumab: A Prospective Study. *J Endocr Soc*. 2018;2(3):241–51.
 30. Inaba H, Ariyasu H, Takeshima K, Iwakura H, Akamizu T. Comprehensive research on thyroid diseases associated with autoimmunity: autoimmune thyroid diseases, thyroid diseases during immune-checkpoint inhibitors therapy, and immunoglobulin-G4-associated thyroid diseases. *Endocr J*. 2019;66(10):843–52.
 31. Pollack RM, Kagan M, Lotem M, Dresner-Pollak R. Baseline TSH Level is Associated with Risk of Anti-PD-1-Induced Thyroid Dysfunction. *Endocr Pract*. 2019;25(8):824–9.
 32. Pollack R, Ashash A, Cahn A, Rottenberg Y, Stern H, Dresner-Pollak R. Immune Checkpoint Inhibitor-induced Thyroid Dysfunction Is Associated with Higher Body Mass Index. *J Clin Endocrinol Metab*. 2020;105(10):e3620–7.
 33. D'Aiello A, Lin J, Gucalp R, Tabatabaie V, Cheng H, Bloomgarden NA, et al. Thyroid Dysfunction in Lung Cancer Patients Treated with Immune Checkpoint Inhibitors (ICIs): Outcomes in a Multiethnic Urban Cohort. *Cancers (Basel)*. 2021;13(6):1464.
 34. de Filette J, Jansen Y, Schreuer M, Everaert H, Velkeniers B, Neyns B, et al. Incidence of Thyroid-Related Adverse Events in Melanoma Patients Treated With Pembrolizumab. *J Clin Endocrinol Metab*. 2016;101(11):4431–9.
 35. Guaraldi F, La Selva R, Samà MT, D'Angelo V, Gori D, Fava P, et al. Characterization and implications of thyroid dysfunction induced by immune checkpoint inhibitors in real-life clinical practice: a long-term prospective study from a referral institution. *J Endocrinol Invest*. 2018;41(5):549–56.
 36. Osorio JC, Ni A, Chaft JE, Pollina R, Kasler MK, Stephens D, et al. Antibody-mediated thyroid dysfunction during T-cell checkpoint blockade in patients with non-small-cell lung cancer. *Annals of Oncology*. 2017;28(3):583–9.
 37. Orlov S, Salari F, Kashat L, Walfish PG. Induction of Painless Thyroiditis in Patients Receiving Programmed Death 1 Receptor Immunotherapy for Metastatic Malignancies. *J Clin Endocrinol Metab*. 2015;100(5):1738–41.
 38. de Filette J, Andreescu C, Cools F, Bravenboer B, Velkeniers B. A Systematic Review and Meta-Analysis of Endocrine-Related Adverse Events Associated with Immune Checkpoint Inhibitors. *Hormone and Metabolic Research*. 2019;51(03):145–56.
 39. Wright JJ, Powers AC, Johnson DB. Endocrine toxicities of immune checkpoint inhibitors. *Nat Rev Endocrinol*. 2021;17(7):389–99.
 40. Muir CA, Clifton-Bligh RJ, Long G V, Scolyer RA, Lo SN, Carlino MS, et al. Thyroid Immune-related Adverse Events Following Immune Checkpoint Inhibitor Treatment. *J Clin Endocrinol Metab*. 2021;106(9):e3704–13.
 41. Brancatella A, Viola N, Brogioni S, Montanelli L, Sardella C, Vitti P, et al. Graves' Disease Induced by Immune Checkpoint Inhibitors: A Case Report and Review of the Literature. *Eur Thyroid J*. 2019;8(4):192–5.
 42. Lee H, Hodi FS, Giobbie-Hurder A, Ott PA, Buchbinder EI, Haq R, et al. Characterization of Thyroid Disorders in Patients Receiving Immune Checkpoint Inhibition Therapy. *Cancer Immunol Res*. 2017;5(12):1133–40.
 43. Chang LS, Barroso-Sousa R, Tolaney SM, Hodi FS, Kaiser UB, Min L. Endocrine Toxicity of Cancer Immunotherapy Targeting Immune Checkpoints. *Endocr Rev*. 2019;40(1):17–65.
 44. Okura N, Asano M, Uchino J, Morimoto Y, Iwasaku M, Kaneko Y, et al. Endocrinopathies Associated with Immune Checkpoint Inhibitor Cancer Treatment: A Review. *J Clin Med*. 2020;9(7):2033.
 45. Yu C, Chopra IJ, Ha E. A novel melanoma therapy stirs up a storm: ipilimumab-induced thyrotoxicosis. *Endocrinol Diabetes Metab Case Rep*. 2015;2015:140092.
 46. McMillen B, Dhillon MS, Yong-Yow S. A rare case of thyroid storm. *BMJ Case Rep*. 2016; bcr2016214603.
 47. Khan U, Rizvi H, Sano D, Chiu J, Hadid T. Nivolumab induced

- myxedema crisis. *J Immunother Cancer*. 2017;5(1):13.
48. Azmat U, Liebner D, Joehlin-Price A, Agrawal A, Nabhan F. Treatment of Ipilimumab Induced Graves' Disease in a Patient with Metastatic Melanoma. *Case Rep Endocrinol*. 2016;2016:2087525.
49. Gan EH, Mitchell AL, Plummer R, Pearce S, Perros P. Tremelimumab-Induced Graves Hyperthyroidism . *Eur Thyroid J*. 2017;6(3):167–70.
50. Haanen J, Obeid M, Spain L, Carbonnel F, Wang Y, Robert C, et al. Management of toxicities from immunotherapy: ESMO Clinical Practice Guideline for diagnosis, treatment and follow-up. *Ann Oncol*. 2022;33(12):1217–38.
51. Brahmer JR, Lacchetti C, Schneider BJ, Atkins MB, Brassil KJ, Caterino JM, et al. Management of Immune-Related Adverse Events in Patients Treated With Immune Checkpoint Inhibitor Therapy: American Society of Clinical Oncology Clinical Practice Guideline. *J Clin Oncol*. 2018;36(17):1714–68.
52. Bruyère CL de la, Souquet PJ, Dalle S, Corbaux P, Boespflug A, Duruisseaux M, et al. Investigating the Impact of Immune-Related Adverse Events, Glucocorticoid Use and Immunotherapy Interruption on Long-Term Survival Outcomes. *Cancers (Basel)*. 2021;13(10):2365.
53. Bruera S, Suarez-Almazor ME. The effects of glucocorticoids and immunosuppressants on cancer outcomes in checkpoint inhibitor therapy. *Front Oncol*. 2022 23;12.
54. Ma C, Hodi FS, Giobbie-Hurder A, Wang X, Zhou J, Zhang A, et al. The Impact of High-Dose Glucocorticoids on the Outcome of Immune-Checkpoint Inhibitor-Related Thyroid Disorders. *Cancer Immunol Res*. 2019;7(7):1214–20.

Corresponding author:

George C. Kyriakopoulos, MPharm, MSc, MD, PhD
Tel.: +30 2610 642 141, E-mail: gkyriakop.123@gmail.com

Childhood vitiligo: Report of a case associated with psychiatric morbidity in the patient's mother and an approach to differential diagnosis

Daniel A. Ndidiamaka Onyiriuka

Abstract

This report describes the case of an adolescent 13-year-old Nigerian girl, diagnosed with the non-segmental variant of vitiligo, specifically the acrofacial subtype. She exhibited some clinical features of psychological disturbance as a result of her current depigmentation skin disorder. She developed the tendency to avoid strangers due to a fear of being talked about, which could potentially lead to social isolation. Although the patient's mother did not have vitiligo herself, she expressed anxiety regarding her daughter's skin disorder. In this context, her major concern was how the disorder might affect her daughter's eligibility for marriage as she approached the age of marriage, as well as whether the disease was curable. Thus, childhood vitiligo is a common skin disorder with significant cosmetic concern on the patient and psychosocial impact on both the patient and primary caregiver. Attention should be given to parents of children with vitiligo, because they may suffer quality of life impairment, requiring psychosocial support to promote better treatment outcomes. In addition, this report highlights the recommended guidelines for managing childhood vitiligo and points out some drawbacks regarding their application.

Key words: *Childhood vitiligo; depigmentation; differential diagnosis; parents; psychosocial impact*

INTRODUCTION

Vitiligo is an acquired, chronic and non-contagious disease characterized by depigmentation, resulting from selective immunologic destruction of epidermal melanocytes [1], causing achromatic macules or spots. Pigment cells of the skin, hair follicles and mucous membranes are commonly involved in the destructive process. Reports of recent translational research has linked key mechanisms of the disease to cellular stress,

innate immune activation, T-cell mediated elimination of melanocytes from the skin, resulting in clinically obvious white spots, as well as stem cell regeneration that reverses established lesions [2]. Vitiligo can appear at sites of trauma or sunburn (Koebner's phenomenon). In older children and adolescents, vitiligo may occur as a component of certain hereditary syndromes such as Vogt-Koyanagi-Harada and Alezzandrini syndromes [3]. In both syndromes, vitiligo appears later and is usually persistent, despite therapy.

Worldwide, the prevalence of vitiligo ranges from 0.5% to 2.0%, with approximately 50% appearing before the age of 20 and 25% of cases starting before the age of 10 [1,4]. A hospital-based study of childhood dermatoses in Ile Ife and Ilesha, Nigeria, reported a vitiligo preva-

Endocrinology and Metabolism Unit, Department of Child Health, University of Benin Teaching Hospital, PMB 1111, Benin City, Nigeria

Received: 31 Dec 2024; Accepted: 14 Apr 2025

lence of 5.3% [5]. The median onset age of childhood vitiligo is 5-10 years [6]. Vitiligo affects people of all ages, genders and races and may be presented anytime from neonatal period to adolescence. In one-third of cases, there is either a positive family history of vitiligo or a halo nevi or premature graying of the hair [7]. It presents as depigmented, well-defined, milk-white macules and patches of different configurations and variable sizes with an unpredictable disease course. With regard to its evolution, vitiligo can change and exhibit three distinct behaviors: stability, progression or regression. Vitiligo is described as stable or inactive if no new lesions have developed over the last 12 months, or an existing lesion shows no sign of progression. It is considered progressive or unstable when new lesions form or develop within 12 months, or when a pre-existing lesion grows larger. Vitiligo is referred to as regressive when spontaneous re-pigmentation of an existing lesion occurs [6]. Childhood vitiligo differs in its clinical characteristics and response to treatment, when compared to adults. The differences in clinical characteristics include higher rates of instability, re-pigmentation, female preponderance and a rarer frequency of association of other autoimmune and endocrine disorders in children compared to adults [4,8]. The frequency of relapses is lower in children than adults.

Childhood vitiligo is classified into two major clinical forms: segmental (SV) and non-segmental (NSV) [4,8]. The NSV is further sub-divided into generalized, acrofacial, universal, mucosal and mixed types. In the USA, Patel et al [9] reported that out of 9,118 eligible children and adolescents with vitiligo, two-thirds had the NSV subtype based on clinician-adjudicated prevalence. Similarly, a systematic review and meta-analysis study, involving 17 studies from different countries to evaluate the clinicoepidemiological features of childhood vitiligo conducted by Faradjzadeh et al. [10], found that non-segmental variant was the most common pattern of presentation. On the other hand, two separate studies in Brazil and China found that the SV is a more frequent pattern of presentation of childhood vitiligo [11,12]. Also in Nigeria, Anaba et al [13] found that the segmental variant was more common than the non-segmental variant. These conflicting findings suggest that there may be differences in the patterns of presentation across geographic regions. This view is reinforced by Faradjzadeh et al [10], who stated that the clinicoepidemiological pattern of childhood vitiligo varies from one geographical region to another.

The diagnosis of childhood vitiligo is primarily based on clinical examination, as the lesions have a typical appearance [8]. Wood's light examination enhances the contrast between pigmented and non-pigmented skin, particularly in light-skinned individuals. Based on severity/extent of body surface area involved, vitiligo can be categorized into four as follows: limited (< 10% involvement), moderate (10-25% involvement), moderately severe (26-50% involvement) and severe (> 50% depigmentation) [14].

Although childhood vitiligo is not typically associated with physical discomfort, it causes great cosmetic concern, not only for the affected person, but also for the primary caregivers because of the associated stigma [15,16]. This concern is more pronounced in darker-skinned individuals, as the contrast between affected and unaffected areas is more noticeable. It has been shown that the psychological effect may persist in adulthood with a negative impact on social development [17]. Vitiligo has been reported to be associated with autoimmune disorders, such as thyroid conditions, type 1 diabetes mellitus, pernicious anaemia, Addison's disease and alopecia areata [18]. In India, Handa and Dogra [19] found that 1.3% of vitiligo cases in children were linked to an associated autoimmune disorder.

The differential diagnosis includes other causes of widespread acquired leukoderma. The three leading differential diagnoses of childhood vitiligo are: tinea versicolor, post inflammatory hypopigmentation and pityriasis alba [17,19]. The purpose of this case report is to raise awareness among physicians that a patient's primary caregiver, such as parents, may be affected psychologically and may need screening for adverse mental health impact.

Case Report

A 13-year-old Nigerian post-menarcheal girl presented with a history of whitish skin lesions. They were first noticed five years earlier in the scalp near the forehead, accompanied by a depigmentation of the hair (poliosis). One year later, similar lesions appeared on her face. Over the next two years, the lesions spread to both hands and feet. Parents have applied some cream, but there has been no improvement. In addition, fluid extracted from certain herbs was also applied to the lesions. The lesion was not associated with itching, pain, or a burning sensation and there was no loss of superficial sensation. The patient expressed anxiety as a result of her clinical condition, and she tends to avoid

strangers to prevent discussions about her appearance. Her mother's major concern was her child's eligibility for marriage and the possibility of a cure. In addition, her mother sometimes feels embarrassed despite the small size of the lesion. Historically, there was no identifiable precipitating factor and there is no history of seizures or intellectual disability.

Physical examination revealed macular depigmented patches on the face, hands and feet. The lesions were milk-white in color, with distinct margins. The lesion in the scalp showed poliosis (leucotrichia) (Figure 1). She had vitiligo lesions on her face, hands and feet (Figures 1 and 2). The extent of skin involvement was less than 5% of body surface area. On palpation of the lesion,



Figure 1. Vitiligo lesions on the forehead and face.



Figure 2. Vitiligo lesion on the feet and hands.

there was no change in texture between unaffected skin and depigmented patches. No underlying atrophy or induration was observed, and the overlying surface was not scaly. The patient did not have halo nevi or goitre. Based on clinical presentation and physical examination, a diagnosis of non-segmental childhood vitiligo with the acrofacial subtype was made.

DISCUSSION

The index patient presented with non-segmental vitiligo (NSV) of the acrofacial subtype. This form of vitiligo is characterized by sparse lesions that generally appear bilaterally over the face and distal extremities. As in the index patient, truncal lesions are usually not seen in the initial stages but may develop over time as acrofacial variants evolve into generalized vitiligo [20]. In line with the findings of several studies [4-7,21,22], the diagnosis of childhood vitiligo in the index patient was based entirely on clinical evaluation, as the lesions have a typical appearance. With regard to severity, the index patient had a limited form of vitiligo as less than 10% of her body surface area was involved [14]. The patient's skin lesion was progressive, starting from the forehead, and spreading to the face, followed by the limbs (Figures 1 and 2). The progressive pattern demonstrated in the index patient aligns with non-segmental vitiligo (NSV), whereas segmental vitiligo (SV) is known to usually stabilize within six months to two years [8,23]. However, some SV may progress to generalized vitiligo over time. Considering that Mazereeuw-Hautier et al [24] reported that vitiligo-associated-autoimmune disorders in childhood occur largely in children with NSV, we determined the patient's fasting blood glucose, thyroid function tests and full blood count. All the results were within normal limits, supporting the reported low incidence of vitiligo-associated-autoimmune disorders in childhood [6,23]. In this context, Handa and Dogra in India found a prevalence of only 1.3% [19].

Regarding the differential diagnosis of childhood vitiligo, clinicians must recognize conditions with similar presentation to avoid misdiagnosis. The common acquired clinical conditions in the differential diagnoses of childhood vitiligo are tinea versicolor, post-inflammatory hypopigmentation and pityriasis alba [25]. Vitiligo is probably the most common disorder of skin pigmentation encountered by primary care physicians. Therefore, a simplified approach to differential diagnosis may be helpful. The common differential diagnoses of childhood vitiligo are presented in tabular form (Table 1). Other

Table 1. Differential diagnosis of childhood vitiligo.

Disorder	Clinical differentiating point from childhood vitiligo
Childhood vitiligo	Acquired skin depigmentation. The lesion is completely amelanotic, non-scaly, chalky-white macule with well-defined margin. Absence of induration, atrophy, bruising and itch/pain in the lesion is characteristic. On Wood's light examination, it emits bright blue-white fluorescence. May be associated with autoimmune and endocrine comorbidities.
Tinea versicolor	Asymptomatic skin infection caused by yeast <i>Malassezia furfur</i> . Characterized by hypopigmented scaly macules mainly over the face in children but the upper trunk in adults. On Wood's light examination, it emits orange fluorescence. Light microscopy of skin scrapings prepared with 10% potassium hydroxide (KOH) may reveal round yeast forms (spores) and short hyphae (referred to as "apples and bananas"). The most appropriate therapy is topical azole or terbinafine. Whitefield's ointment applied twice daily for 3 weeks is also effective.
Postinflammatory Hypopigmentation	History of active skin lesions in the past. Hypopigmented lesion. May occur as sequelae of impetigo, chicken pox infection, onchocerciasis, insect bites or physical trauma. Often retains the shape and size of the original skin lesion. Self-resolution is common
Pityriasis alba	Common in prepubertal children with personal or family history of atopy. Characterized by ill-defined hypopigmented macules with fine scaling; mostly over the face and upper arm. History of pruritus and irritation, especially at the borders may be present. It does not progress and is self-limited. On Wood's light examination, it emits off-white glow with non-circumscribed edges. Low-potency topical corticosteroids may help decrease the inflammatory component and lead to faster return of normal pigmentation. Healthy skin barrier can be achieved with the application of emollients. No associated comorbidities.

significant differential diagnoses include the ash-leaf macule of tuberous sclerosis (lesions typically present at birth or early in infancy) and morphea (the skin of the lesion is usually smooth, thickened and has shiny appearance), whereas vitiligo lesions maintain the same texture as the surrounding skin.

The index patient was an adolescent girl aged 13 years and manifested some clinical features of psychological disturbance as a result of her current skin disorder. As previously reported, in India [26], the index patient developed the tendency to avoid strangers to prevent discussions about her. This behavior often drives patients into social isolation and introversion, particularly when the condition affects the exposed parts of the body. It is known that adolescence is a critical period for developing self-identity and self-esteem [15]. Developmentally, the patient belongs to an age group where self-image is being formed, and social acceptance is of great importance. Thus, the patient's skin disorder has negatively affected her self-confidence and social interactions, leading to diminished quality of life. Despite the fact that the patient's mother did not have vitiligo, she expressed anxiety regarding her daughter's skin disorder. In this context, the mother's major concern was its potential in affecting her daughter's eligibility

for marriage, as she approaches the age of marriage. As highlighted in previous reports [16,26,27], parents of children with vitiligo are often considered hidden victims of this relatively common childhood skin disorder. The mother who represents the closest and most caring relationships to her affected daughter becomes a hidden victim. In consideration of the psychological impact of the patient's skin disorder on the mother, it is important to recognize parental quality of life impairment and provide psychological support to promote treatment adherence and decrease the family burden of chronic skin disorder. The patient was accompanied only by her mother during clinic visits, which indicates higher maternal concern regarding the condition. This observation suggests that mothers may be more distressed than the father with regard to the child's skin disorder. A similar observation has been alluded to in the report by Ameer et al [16] in China. The report of some previous studies suggested that because of the cosmetic concern in vitiligo, patients/parents tend to seek medical attention early [28,29]. In contrast, the mother of the index patient sought medical attention five years after the first lesion appeared, despite its progressive nature and involvement of the face in the index case, suggesting poor health-seeking behavior,

is probably due to a lack of symptoms in the patient.

The natural history of vitiligo involves periods of remission and exacerbation, with complete spontaneous re-pigmentation being unusual. Partial or temporary re-pigmentation has been reported in children, particularly for lesions of less than two years' duration and during summer. The re-pigmentation process is slow, with the face and trunk typically responding better than the dorsa of hands and feet [30]. In the case of the index patient, the duration of the lesion was five years (exceeding the two-year threshold associated with re-pigmentation), potentially making spontaneous re-pigmentation less probable.

The treatment of vitiligo in children is very challenging, as many of available modalities of therapy cannot be applied to them. The two main goals of therapy of childhood vitiligo are (i) stabilization of active disease and (ii) promotion of re-pigmentation [31]. Factors influencing the choice of treatment plan include the patient's age, disease duration, extent and severity of the condition, presence of Koebner phenomenon and the association of other autoimmune conditions [31]. Other factors include the socio-economic status of the patient's family and availability of treatment options. According to the 2021 guidelines of the British Association of Dermatologist, the first-line option for managing vitiligo is topical non-steroid therapy [32].

In this regard, the use of tacrolimus is considered the first-line option in the treatment of childhood vitiligo because of the delicate nature of children's skin, which exposes them to a higher risk of skin atrophy from the use of topical corticosteroids (TCs). Additionally, the surface area-to-volume ratio in children increases the risk of the occurrence of adverse effects of steroids. Oral corticosteroids can interfere with growth in children; therefore, their use requires caution. Regarding therapy for childhood vitiligo, Raju et al [33] suggested that moderately potent topical corticosteroids (TCs) are considered as first-line therapy in children with localized vitiligo. Topical calcineurin inhibitors (TCIs) such as tacrolimus/pimecrolimus are good alternative to TCs and are recommended localized forms of vitiligo, as well as for areas with thin skin like eyelids where potential side effects of corticosteroids are high. TCIs cannot be used in children less than two years old because of the risk of skin cancer and lymphoma. In addition, TCIs are expensive and are not readily available in the environment where we practice. A synthetic vitamin D3 analogue (calcipotriol) though less effective than topical corticosteroids can stimulate melanogenesis and inhibit the destruction of melanocytes by T-cells. The combination of phototherapy (NB-UVB), psoralen and UAV (PUAV) is considered a second-line treatment option. Phototherapy acts as an immunosuppressant

Table 2. Various treatment modalities of childhood vitiligo [31].

Topical medical therapy	Systemic medical therapy	Surgical therapy
Corticosteroids	1. Corticosteroids (OMP with betamethasone/ethylprednisolone)	1. Conventional
Tacrolimus/pimecrolimus		Mini-punch graft
Calcipotriol	2. Phototherapy	Suction blister epidermal Graft
Pseudocatalase	Topical PUVA	Thin Thiersch graft
Combination (corticosteroids and tacrolimus)	NB-UVB	
	Systemic PUVA (> 12 years)	3. Newer cellular transplantation techniques
	Phenylalanine + PUVA	Epidermal cell suspension
	Excimer laser (308nm)/targeted NB-UVB phototherapy	Cultured melanocyte Suspension
		Cultured epidermis

NB-UVB = Narrow Band Ultraviolet B.

and stimulates melanocyte activity [34]. The third-line treatment option is surgical therapy. Though not commonly used in children, suction blister epidermal grafting is preferred. Surgical procedures are not performed in very young children because segmental or stable focal lesions in younger children extend proportionate to their body growth. In addition, success of many surgical procedures depends on post-operative immobility of the operated part which is more difficult to achieve in young children [31]. Other potential treatment modalities of childhood vitiligo are displayed in Table 2.

Other modalities of treatment include (i) cosmetic camouflage (ii) Total depigmentation, using 20% Monobenzyl ether of hydroquinone (MBEH) [31].

In summary, a practical conclusion from this report is that increasing the use of psychiatric screening questionnaires might enhance recognition of psychiatric morbidity and the adverse mental health impact on mothers of children with vitiligo, ultimately leading to early intervention.

Conflict of interest disclosure: None to declare.

Declaration of funding sources: None to declare.

Author Contributions: DNO confirms sole responsibility for all aspects of the manuscript, including conceptualization, methodology, data collection, analysis, writing, and final approval of the submitted version.

REFERENCES

- Kruger C, Schallreuter KU. A review of the worldwide prevalence of vitiligo in children/adolescents and adults. *Int J Dermatol*. 2012;51(10):1206-12.
- Katz EL, Harris JE. Translational research in vitiligo. *Front Immunol*. 2021;12:624517.
- Paller AS, Mancini AJ. Disorders of pigmentation. In: Paller AS, Mancini AJ, editors. *Hurwitz Clinical Pediatric Dermatology*. 3rd ed. Philadelphia: Elsevier Saunders; 2006. p. 265-305.
- Palit A, Inamadar AC. Childhood vitiligo. *Indian J Dermatol Venereol Leprol*. 2012;78(1):30-41.
- Oninla AO, Oninla SO, Onayemi O, Olasode AO. Pattern of paediatric dermatoses at dermatology clinics in Ile-Ife and Ilesha, Nigeria. *Paediatr Int Child Health*. 2016;36(1):1-7.
- Ezzedine K, Silverberg N. A practical approach to the diagnosis and treatment of vitiligo in children. *Pediatrics*. 2016;138(1):e20154126.
- Bolognia JL, Pawelck JM. Biology of hypopigmentation. *J Am Acad Dermatol*. 1988;19(2):217-55.
- Petrovic-Stern A, Ahmad N, Wiley A, Levy RM, Kundu RV, Mancini AJ, et al. The relationship between family medical history and childhood vitiligo. *J Am Acad Dermatol*. 2006;55(2):238-44.
- Patel R, Pandya AG, Sikirica V, Ghandi K, Daniel SR, Anastassopoulos KP, et al. Prevalence of vitiligo among children and adolescents in the United States. *Dermatology*. 2023;239(3):227-34.
- Faradjzadeh S, Khalili M, Mirmohammadkhani M, Paknazar F, Rastegarnasab F, Abtahi-Naeini B. Global clinicoepidemiological pattern of childhood vitiligo: a systematic review and meta-analysis. *BMJ Paediatr Open*. 2023;7(1):e001839.
- Lin X, Tang LY, Fu WW, Kang KF. Childhood vitiligo in China: clinical profiles and immunological findings in 620 cases. *Am J Clin Dermatol*. 2011;12(4):277-81.
- de Barros JC, Machado Filho CD, Abreu LC, de Barros JA, Paschoal FM, Nomura MT, et al. A study of clinical profiles of vitiligo in different ages: analysis of 669 outpatients. *Int J Dermatol*. 2014;53(7):842-8.
- Anaba EL, George AO, Ogunbiyi AO. Vitiligo: any differences in adult and childhood clinical characteristics? *Niger Health J*. 2018;18(3):90-6.
- Grimes PE. Therapies for vitiligo. In: Millikan LE, editor. *Drug Therapy in Dermatology*. New York: Marcel Dekker; 2000. p. 339-56.
- Silverberg JI, Silverberg NB. Quality of life impairment in children and adolescents with vitiligo. *Pediatr Dermatol*. 2014;31(3):309-18.
- Amer AAA, McHepange UO, Gao X-H, Hong Y, Qi R, Wu Y, et al. Hidden victims of childhood vitiligo: impact on parents' mental health and quality of life. *Acta Derm Venereol*. 2015;95(3):322-5.
- Linthorst Homan MW, de Korte J, Grootenhuys MA, Bos JD, Sprangers MA, van der Veen JP. Impact of childhood vitiligo on adult life. *Br J Dermatol*. 2008;159(4):915-20.
- Pride HB. Pigmentary disorders: white spots, brown spots, and other dyschromias. In: Pride HB, Yan AC, Zaenglein AL, editors. *Requisites in Dermatology: Pediatric Dermatology*. Philadelphia: Saunders Elsevier; 2008. p. 105-24.
- Handa S, Dogra S. Epidemiology of childhood vitiligo: a study of 625 patients from north India. *Pediatr Dermatol*. 2003;20(3):207-10.
- Delgado P, Crumpton K. A practical guide to vitiligo differential diagnoses in primary care. *Nurse Pract*. 2021;46(11):29-36.
- Habib A. Vitiligo in children: a distinct subset. *J Coll Physicians Surg Pak*. 2016;26(3):173-6.
- Davda B, Sivasubramaniam V. Childhood vitiligo: a hospital-based retrospective study in coastal south India. *IP Indian J Clin Exp Dermatol*. 2020;6(3):227-30.
- Taieb A, Picardo M. Clinical practice: vitiligo. *N Engl J Med*. 2009;360(2):160-9.
- Mazereeuw-Hautier J, Bezio S, Mahe E, Bodemer C, Eschard C, Viseux V, et al. Segmental and nonsegmental childhood vitiligo has distinct clinical characteristics: a prospective observational study. *J Am Acad Dermatol*. 2010;62(6):945-9.
- Morelli JG. Hypopigmented lesions. In: Kliegman RM, Behr-

- man RE, Jenson HB, Stanton BF, editors. *Nelson Textbook of Pediatrics*. 18th ed. Philadelphia: Saunders Elsevier; 2007. p. 2682-5.
26. Mattoo SK, Handa S, Kaur I, Gupta N, Malhotra R. Psychiatric morbidity in vitiligo and psoriasis: a comparative study from India. *J Dermatol*. 2001;28(8):424-32.
 27. Andrade G, Rangu S, Povlin L, Putterman E, Gauthier A, Castelo-Soccio L. Childhood vitiligo impacts emotional health of parents: a prospective, cross-sectional study of quality of life for primary caregivers. *J Patient Rep Outcomes*. 2020;4(1):20.
 28. Zahra FT, Amin SS, Adil M, et al. Clinico-epidemiological study of childhood vitiligo and its associations: a hospital-based cross-sectional study. *Indian J Paediatr Dermatol*. 2022;23(2):116-21.
 29. El-Husseiny R, Abd-Elhaleem A, Salah El-Din W, et al. Childhood vitiligo in Egypt: clinic-epidemiologic profile of 483 patients. *J Cosmet Dermatol*. 2021;20(1):237-42.
 30. Cline DJ, Nordlund JJ. Vitiligo. In: Greek KE, editor. *Common Problems in Dermatology*. Chicago: Year Book Medical Publishers; 1988. p. 321-39.
 31. Tamesis ME, Morelli JG. Vitiligo treatment in childhood: a state of the art review. *Pediatr Dermatol*. 2010;27(5):437-45.
 32. Eleftheriadou V, Atkar R, Batchelor J, et al. British Association of Dermatologists guidelines for the management of people with vitiligo. *Br J Dermatol*. 2021;186(1):18-29.
 33. Raju SP, Kaur S, Loganathan E. Management of childhood vitiligo—a brief review. *Pigment Int*. 2022;9(1):14-24.
 34. Gianfaldoni S, Zarrab Z, Lotti T. Phototherapy and vitiligo repigmentation: from PUVA to micro-focused phototherapy. *J Pigment Disord*. 2014;1:102.

Corresponding author:

Daniel A. Ndidiamaka Onyiriuka
E-mail: alpondiony@yahoo.com

INSTRUCTIONS FOR AUTHORS

The journal "Achaiki Iatriki" publishes original papers on clinical and basic research from all areas of the health sciences including healthcare. The journal is published exclusively in English. Manuscripts should conform to the guidelines set out in "Uniform Requirements for Manuscripts Submitted to Biomedical Journals" by the International Committee of Medical Journal Editors (<http://www.icmje.org>).

COVER LETTER

A submission letter to the Editor should accompany the manuscript and contain the following:

- The manuscript has not been published previously, and is not under consideration for publication elsewhere.
- Acknowledgment of grants or financial support.
- The manuscript has been approved by all authors.

INFORMATION ABOUT ARTICLE TYPES

The Editors will consider and publish the following:

1. Original research articles
2. Narrative Reviews
3. Systematic Reviews and Meta-analyses
4. Editorials
5. Letters to the Editor
6. Case Reports

Original research articles

The maximum length of the main text is 3,500 words excluding the abstract, references, tables, and figure legends. A maximum of 6 tables and/or figures is allowed. References should not exceed a maximum of 100.

Narrative Reviews / Systematic Reviews / Meta-analyses

These manuscripts are solicited and unsolicited manuscripts that feature an organized and detailed review of the scientific literature about a particular topic. This section is peer-reviewed and acceptance for publication is not guaranteed. The maximum length of the main text is 5,000 words excluding the abstract, references, tables, and figure legends. A maximum of 6 tables and/or figures to summarize critical points is highly desirable. References should not exceed a maximum of 150.

Editorials

Editorials are usually solicited by the Editor. The maximum length is 1500 words excluding the references, tables, and figure legends. One table or 1 figure is allowed. References should not exceed a maximum of 20. Editorials may have a maximum of three (3) authors.

Letters to the Editor

Letters to the Editor will be considered for publication if they are related to articles published in recent issues of the Achaiki Iatriki Journal. The maximum length is 800 words (excluding references, table, and figure legend). A total number of 1 table or figure is allowed and up to 10 references. Such letters will be passed to the authors of the original paper, who will be offered an opportunity to reply. Letters to the Editor may have a maximum of two (2) authors.

Case Reports

Case reports should ideally include a short introduction, the case presentation and a brief discussion. The maximum length is 1500 words (excluding references, tables, and figure legend). A total number of 2 tables or figures is allowed. References should not exceed a maximum of 15.

Formatting guide

The articles must be typewritten and double spaced. They should include the following sections, each starting on a separate page:

- Title Page
- Abstract and Key Words
- Main Text
- Acknowledgements
- References
- Tables
- Figures

Margins should be not less than 2.5 cm. Pages should be numbered consecutively.

Abbreviations

Do not use non-standard abbreviations. The use of abbreviations in the title and abstract should be avoided. Abbreviations should be defined on their first appearance in the text; those not accepted by international bodies should be avoided.

Title page

The title page should include:

- Title of the manuscript
- Short title which will be used as a running head
- Full name of each author
- Full location of department and institution where work was performed
- Name and address for correspondence, including fax number, telephone number, and e-mail address.
- Conflict of interest disclosure.
- Declaration of funding sources.
- Author Contributions according to the following criteria for authorship: conception and design; analysis and interpretation of the data; drafting of the article; critical revision of the article for important intellectual content; final approval of the article.

Abstract

For Original Articles, structured abstracts should be 250 words or less and include the following sections: Background, Methods, Results and Conclusion. Review articles should carry an unstructured abstract which should not exceed 200 words.

Key words

The abstract should be followed by a list of 3–5 keywords which will assist the cross-indexing of the article and which may be published separated by semicolons.

Main Text

For the main body of the text, the recommended structure of

the manuscript is:

- Introduction
- Materials and Methods
- Results
- Discussion

Define abbreviations at first mention in text and in each table and figure.

Introduction

State the objectives of the work and provide an adequate background, avoiding a detailed literature survey or a summary of the results.

Materials and Methods

Provide sufficient detail to allow the work to be reproduced. Methods already published should be indicated by a reference. This includes a description of the design, measurement and collection of data, type and source of subjects, inclusion and exclusion criteria and measures of outcome, number of subjects studied and why this number was chosen. Any deviation from the study protocol should be stated. Randomized controlled trials should adhere to the CONSORT guidelines that can be found at: <http://www.consort-statement.org>. Observational studies should also adhere to Strobe statement: <http://www.strobe-statement.org/>. Diagnostic accuracy studies should follow the Stard statement: <http://www.stard-statement.org/>. Systematic Reviews and Meta-Analyses should adhere to the PRISMA statement: <http://www.prisma-statement.org/>.

Statistical analysis

The statistical methods used should be relevant and clearly stated. Special or complex statistical methods should be explained and referenced. Describe statistical methods with enough detail to enable a knowledgeable reader with access to the original data to verify the reported results. When possible, quantify findings and present them with appropriate indicators of measurement error or uncertainty (such as confidence intervals). Avoid relying solely on statistical hypothesis testing, such as P values, which fail to convey important information about effect size. Define statistical terms, abbreviations, and symbols. Specify the software used.

Units

Follow internationally accepted rules and conventions: use the internal system of units (SI).

Results

Results should be clear and concise. Results should be explained and illustrated by using Tables and Figures. Do not duplicate information contained in tables and figures.

Discussion

Discussion should directly relate to the results of the study and should explore their significance. Do not provide a general review of the topic.

Conclusions

The conclusions should provide a summary of the key results and discuss the appropriateness and impact of this original work.

Acknowledgements

Collate acknowledgements in a separate section at the end of the article before the references. Acknowledgements should be made only to those who have made a substantial contribution to the study. Authors are responsible for obtaining written permission from people acknowledged by name in case readers infer their endorsement of data and conclusions.

References

Ensure that every reference cited in the text is also present in the reference list (and vice versa). References should be numbered in the order they appear in the text. Manuscripts should follow the style of the Vancouver agreement detailed in the International Committee of Medical Journal Editors' revised 'Uniform Requirements for Manuscripts Submitted to Biomedical Journals: Writing and Editing for Biomedical Publication', as presented at <http://www.ICMJE.org/>. In the text, references should be cited using Arabic numerals enclosed in square brackets [1]. The last names and initials of all authors should be referred to if they are up to six, otherwise only the first six are referred, with et al following. References should also include full title and source information. Journal names should be abbreviated according to the standard in the Index Medicus. No periods should be placed at the end of abbreviations of the journal.

Journal article, up to 6 personal author(s):

Example: Al-Habian A, Harikumar PE, Stocker CJ, Langlands K, Selway JL. Histochemical and immunohistochemical evaluation of mouse skin histology: comparison of fixation with neutral buffered formalin and alcoholic formalin. *J Histotechnol*. 2014;37(4):115-24.

Journal article, more than 6 personal author(s):

Example: Liaw S, Hasan I, Wade V, Canalese R, Kelaher M, Lau P, et al. Improving cultural respect to improve Aboriginal health in general practice: a multi-perspective pragmatic study. *Aust Fam Physician*. 2015;44(6):387-92.

Journal article/ Issue with a supplement

Example: Bonda C, Sharma P, LaFaver K. Clinical reasoning: a 28 year-old woman with lower extremity spasticity and microcytic anemia. *Neurology*. 2015;85(2) Suppl:e11-4.

Electronic journal article:

Example: Poling J, Kelly L, Chan C, Fisman D, Ulanova M. Hospital admission for community-acquired pneumonia in a First Nations population. *Can J Rural Med [Internet]*. 2014 Fall [cited 2015 Apr 27];19(4):135-41. Available from: <http://www.srpc.ca/14fal.html> by selecting PDF link in table of contents.

Book, personal author(s):

Example: Buckingham L. *Molecular diagnostics: fundamentals, methods and clinical applications*. 2nd ed. Philadelphia: F.A. Davis; c2012.

Book or pamphlet, organization as both author and publisher:

Example: College of Medical Radiation Technologists of Ontario. *Standards of practice*. Toronto: The College; 2011.

Book, editor(s):

Example: Kumar V, Abbas AK, Aster JC, editors. Robbins basic pathology. 16th ed. Philadelphia: Elsevier Saunders; c2013.

Poster presentation/session presented at a meeting or conference:

Example: Chasman J, Kaplan RF. The effects of occupation on preserved cognitive functioning in dementia. Poster session presented at: Excellence in clinical practice. 4th Annual Conference of the American Academy of Clinical Neuropsychology; 2006 Jun 15-17; Philadelphia, PA.

Tables

Tables should be typewritten, double-spaced, each one on a separate page and numbered consecutively with Arabic numerals in the order of their appearance in the text. Do not duplicate material presented in a figure. Tables should include a short but concise title. Tables should read vertically when possible. Place explanatory matter in footnotes, including any non-standard abbreviation. If data from another published or unpublished source are used, obtain permission and acknowledge fully.

Figure legends

Figure legends should be listed one after the other, as part of the main text, separate from the figure files. Each figure legend should have a brief title (in bold with figure number) followed by a description of each panel, and the symbols used. The statistical test used as well as the values of statistical significance (whether significant or not) should always be included in the figure legends. If a figure has been published previously, acknowledge the original source and submit written permission from the copyright holder to reproduce it. Authors will be required to pay for the extra cost of printing illustrations in color. However, there is an option to have their images in color in the electronic version of their manuscript and in grey scale in the printed version.

Figures

All figures for review should be submitted as a separate file in JPEG or TIFF format in grayscales or in RGB color mode with a resolution of at least 300 dpi. Number figures consecutively using Arabic numerals.

Photographs should be submitted as TIFF with a resolution of at least 300 pixels per inch; or Illustrator compatible EPS files with RGB color management or Photoshop or editable PDF files (grayscale or RGB).

Photographs of identifiable patients should be accompanied by written permission to publish from patient(s).

RGB figures will be presented in color in the electronic version and in grey scale in the printed version.

Ethical Considerations

An author should not publish manuscripts describing essentially the same research in more than one journal or primary publication. It must not be under consideration for publication elsewhere, and, if accepted, must not be published elsewhere in similar form, in any language. The International Committee of Medical Journal Editors has a full description about duplicate or redundant publication (<http://www.icmje.org>).

Authorship should be limited to those who have made a significant contribution to the conception, design, execution, or interpretation of the reported study.

The 'Achaiki Iatriki' editors endorse the principles of the Declaration of Helsinki and expect that all investigations involving humans will have been performed in accordance with these principles.

Authors should carefully protect patients' anonymity. Manuscripts reporting data from research conducted on humans must include a statement of assurance in the materials and methods section describing that: written informed consent was obtained from each patient included in the study and that the study protocol conforms to the ethical guidelines of the 1975 Declaration of Helsinki as reflected in a priori approval by the institution's human research committee.

Do not use patients' names, initials, or hospital numbers, especially in illustrative material.

For animal experimentation reported in the journal, it is expected that investigators will have observed the Interdisciplinary Principles and Guidelines for the Use of Animals in Research, Testing, and Education issued by the New York Academy of Sciences' Adhoc Committee on Animal Research.

Disclosures: Conflict of interest

All authors are required to provide a Declaration of Interest Statement recognizing and disclosing financial and other conflicts of interest that might bias their work. Particularly, they disclose any actual or potential conflict of interest including any financial, activities, additional affiliations, personal or other relationships with other people or organizations within three years of beginning the submitted work that could inappropriately influence, or be perceived to influence, their work. Further information at International Committee of Medical Journal Editors ("Uniform Requirements for Manuscripts Submitted to Biomedical Journals") -- February 2006

Disclosures: Financial disclosure

Authors are requested to identify who provided financial support for the conduct of the research and/or preparation of the article and to briefly describe the role of the sponsor(s), if any, in study design; in the collection, analysis and interpretation of data; in the writing of the report; and in the decision to submit the article for publication. If the funding source(s) had no such involvement then this should be stated.

Inform Consent

Patients have a right to privacy that should not be infringed without informed consent. Information such as patients' names, initials, or hospital numbers, should not be published in written descriptions, photographs, and pedigrees unless the information is essential for scientific purposes and the patient (or parent or guardian) gives written informed consent.

Identifying details should be omitted if they are not essential. Complete anonymity is difficult to achieve, however, and informed consent should be obtained if there is any doubt. For example, masking the eye region in photographs of patients is inadequate protection of anonymity. If identifying charac-

teristics are altered to protect anonymity, such as in genetic pedigrees, authors should provide assurance that alterations do not distort scientific meaning.

Further information at International Committee of Medical Journal Editors ("Uniform Requirements for Manuscripts Submitted to Biomedical Journals") -- February 2006

Human and Animal Rights

Manuscripts reporting experiments using humans or animals must include a statement giving assurance that all humans or animals received human care and that study protocols comply with the institution's guidelines. When reporting experiments on human subjects, authors should indicate whether the procedures followed were in accordance with the ethical standards of the responsible committee on human experimentation (institutional and national) and with the Helsinki Declaration of 1975, as revised in 2000. When reporting experiments on animals, authors should be asked to indicate whether the institutional and national guide for the care and use of laboratory animals was followed.

Further information at International Committee of Medical Journal Editors ("Uniform Requirements for Manuscripts Submitted to Biomedical Journals") -- February 2006

Copyright assignment

Upon acceptance of an article, authors will be asked to complete

a copyright assignment indicating that exclusive copyright in the paper is assigned to the Publisher.

MANUSCRIPT PROCESSING AND REVIEW

Submission

You can submit your manuscript either in Journal's website submission system or via email to achaiki.iatriki@gmail.com

Review process

Each manuscript submitted to ACHAIKI IATRIKI is assigned to a Section Editor who has expertise on the subject of the manuscript. The Section Editor initially evaluates the manuscript if it is appropriate and competitive for publication and sends the manuscript to 2-4 reviewers who are experts in the field.

PUBLICATION

Proofs

Proofs will be made available to the author(s) to be checked. It is the responsibility of the author(s) to make sure that the quality and accuracy of the manuscript, figures, and tables in the proofs is correct. At this stage, authors may make only minor corrections. Authors should return their proofs within 48 hours, by e-mail. At this point the author may order reprints, which are charged according to the number of reprints and the number of pages of the article.

Achaiki Iatriki

